

## Enamel Matrix Derivate for Periodontal Regeneration in the Interproximal Periodontal Defect Model

Kyu An Lee, Hyun-Chang Lim, Min-Soo Kim, Jung-Seok Lee, Seong-Ho Choi, and Ui-Won Jung\*

Department of Periodontology, Research Institute for Periodontal Regeneration, College of Dentistry, Yonsei University, Seoul, Korea  
(Received September 16, 2013 / Revised October 10, 2013 / Accepted October 15, 2013)

The aim of this study was to evaluate the effect of enamel matrix derivate (EMD) with combination of composite bovine-derivate xenograft on the periodontal regeneration in the interproximal periodontal defect model. The interproximal periodontal defects (IPDs) were surgically prepared between the first and second maxillary premolar, and the second and third maxillary premolar in four beagle dogs. EMD, collagenized bovine hydroxyapatite (CBHA), combination of two material, and sham surgery were allocated for each IPD. After eight weeks, the animals were sacrificed and the defects were analyzed by radiographic, histologic, and histometric methods. Regenerated woven bone was observed and cementoid was created along the adjacent root surfaces with proliferation of cementoblasts in every group. In the combination of EMD and CBHA group, Sharpey's fiber was observed beyond the crest of new bone and along the newly formed cementum, and apical migration of junctional epithelium appeared to be blocked by new cementum. In the BC and EMD+CBHA groups, the residual bovine hydroxyapatite particles were found in the periodontal defect. No direct contact was observed between residual particles and tooth surfaces. No remarkable difference was found between the histometric results among the groups. Within the limitation of this study, EMD, CBHA, and combination of two materials showed similar periodontal regeneration in the interproximal periodontal defect model. Further investigation on combination with barrier membrane may be required for improvement of the regenerative potential.

**Key words:** enamel matrix proteins, xenograft bioprosthesis, periodontal disease, regeneration

### Introduction

For periodontal regeneration, guided tissue regeneration (GTR) has been widely used with the concept of selective repopulation of periodontal ligament cells.<sup>1,2</sup> Histologically GTR has shown the formation of new cementum, periodontal ligament, and bone in animal studies in various defect types.<sup>3-9</sup> Human studies have also reported regeneration of cementum and periodontal ligament in histologic analysis.<sup>1,2,10,11</sup> Clinical attachment gain and probing depth reduction were observed following GTR in human clinical trials.<sup>12-16</sup> Therefore, GTR is considered as a reliable method for the regeneration of periodontal tissue.

However, alternative approaches for periodontal regeneration have been investigated for avoidance of complication and difficulty of GTR. Major complications of GTR include exposure of barrier membranes. When the membrane was exposed, cell occlusion is impaired and epithelial downgrowth is not blocked perfectly. In addition, GTR requires advanced flap management and suture techniques. The outcomes of GTR are thus often affected by technique sensitivity.

Enamel matrix derivate (EMD) has been introduced to facilitate the periodontal regeneration. Amelogenin is a protein that plays an important role in the development of enamel structure. EMD refers to purified hydrophobic amelogenins,<sup>17</sup> which promotes periodontal regeneration by mimicking the development of periodontium.<sup>18</sup> For the vehicle of EMD propylene glycol alginate (PGA) showed superior results to other alternatives such as hydroxyethyl cellulose.<sup>19</sup>

EMD showed formation of new cementum, periodontal ligament, and bone when applied in animal intrabony,<sup>18</sup> furcation,<sup>20</sup> and dehiscence type defects.<sup>21,22</sup> In human clinical studies Emdogain showed bone regeneration, clinical attachment gain, and probing depth reduction in intrabony defects<sup>23-27</sup> and class II furcation defects.<sup>28,29</sup>

Lack of space maintenance, however, prevents the periodontal regeneration by EMD in non-contained defects. The amount of radiographic and clinical gain followed by application of EMD is significantly less in the 1-wall intrabony defects than 2- or 3-wall defects.<sup>30</sup> In class III furcation defects EMD failed to achieve complete healing of the defect and there was no significant difference in clinical outcomes between EMD and GTR.<sup>31</sup>

Combination of bone material with EMD has been con-

\*Corresponding author: drjew@yuhs.ac

ducted for improvement of space maintenance and regeneration potentials. The effectiveness of combination of bone material with EMD shows inconsistent outcomes and needs more investigation. For space maintenance particulated bone material was used in combination with EMD. Bovine hydroxyapatite (BH) was widely used due to superior biocompatibility. In vitro EMD enhanced the cell activity on residual bovine bone particles.<sup>32)</sup> However, the effectiveness of combination remained still controversial in human studies.<sup>33-37)</sup> In animal intrabony defects EMD combined with BH enhanced the formation of new connective tissue and bone,<sup>38)</sup> and similar results were shown in a human study.<sup>39)</sup> However, other studies failed to show that combination of EMD with other agents can improve the final results.<sup>37,40,41)</sup>

A collagenized bovine hydroxyapatite (CBHA) was also introduced in periodontal regeneration. CBHA consists of mainly deproteinized bovine cancellous bone granules (90%), which are embedded in highly purified collagen matrix (10%). In intrabony defects CBHA itself showed superior clinical gain in human clinical studies.<sup>42-44)</sup> New bone, cementum, and periodontal ligament were observed in histologic analysis of human intrabony defects.<sup>45)</sup> Grafted in extraction sockets, CBHA has exhibited less shrinkage of residual ridge,<sup>46,47)</sup> which implies the potential of preservation of residual ridge. However, regenerative potential of combination therapy with CBHA and EMD has not been clarified.

The aim of this study was to evaluate the effect of EMD with application of CBHA on the periodontal regeneration in the interproximal periodontal defect model.

## Materials and Method

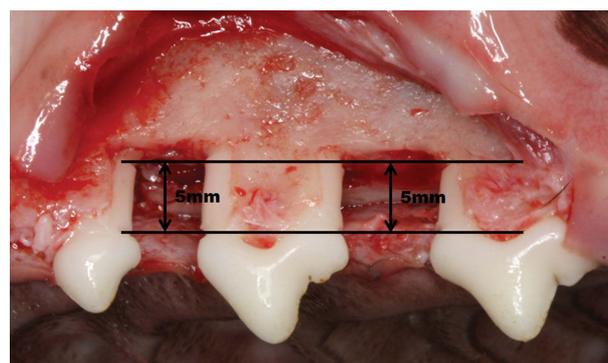
### Animals

Four 15-months-old beagle dogs with healthy periodontium, each weighing 10-15 kg, were used. Animal selection, management, preparation, and the surgical protocol followed the protocol that was approved by the Institutional Animal Care and Use Committee, Yonsei Medical Center, Seoul, Korea.

### Surgical protocol

The surgical procedure was performed under general anesthesia with subcutaneous injection of atrophine 0.05 mg/kg and intravenous injection of compound of xylazine (Rompun®, Bayer Korea, Seoul, Korea) 2 mg/kg and ketamine hydrochloride (Ketalar®, Yuhan Co., Seoul, Korea) 10 mg/kg. After intubated, 2% enflurane was administered and the disinfection of surgical sites was performed. Routine dental infiltration anesthesia was performed with 2% lidocaine hydrochloride including epinephrine 1:100,000 (Kwangmyung Pharm., Seoul, Korea). Mucoperiosteal flap was reflected after a crevicular and vertical incision at the maxillary premolar area.

The interproximal periodontal defects (IPDs) were surgically



**Figure 1.** The interproximal periodontal defect (IPD) prepared between maxillary premolars.

prepared in each side between the first and second maxillary premolar (P1 and P2), and the second and third maxillary premolar (P2 and P3) with a fissure bur and a chisel with continuous saline irrigation for prevention of overheating. The width of the defect was 3 mm, measured from the center of the defect to buccal and palatal bone. The apical notch was created on the roots of the teeth adjacent to the defects to ensure the height of the defect to be 5 mm, measured from the apical reference notch to the cemento-enamel junction (Figure 1).

The root planning of teeth were performed with root planning bur and Gracey curettes to remove old cementum. One of following material was randomly allocated for each IPD: Bio-oss Collagen® (Geistlich Sons Ltd., Wolhusen, Switzerland), Emdogain® (Straumann, Basel, Switzerland), combination of both materials, and sham surgery. After application of material, the flap was positioned coronally and primary tension-free wound closure was accomplished. The flap was sutured with 5-0 resorbable suture material (Polyglactin 910, braided absorbable suture, Ethicon, Johnson & Johnson Int., Edinburgh, U.K.) with interrupted suture.

### Post-surgery management

Post-surgery management included intravenous injection of antibiotics (Cefazoline Sodium 20 mg/kg, Yuhan Co., Seoul, Korea) and topical application of chlorhexidine solution (Hexamedine 0.2%, Bukwang pharmaceutical Co., Seoul, Korea) daily for the prevention of infection. The sutures were removed at approximately 10 days post-surgery. The animals were euthanized after 8 weeks following the surgery by intravenous injection of anesthesia drug overdose.

### Histologic processing and radiographic analysis

Block sections including segments of the defects at the surgical sites were dissected at sacrifice. The sections were fixed in 10% buffered formalin for ten days and rinsed with water. Then the blocks were scanned with a micro-CT (SkyScan 1072®, SkyScan, Aartselaar, Belgium) at a resolution of 35  $\mu$ m (100 kV, 100  $\mu$ A). The scanned data were converted into a

Digital Imaging and Communications in Medicine (DICOM) format and the surgical sites were three-dimensionally reconstructed with OnDemand 3D® software (Cybermed, Seoul, Korea). The residual material was selected in each axial cross section, and combined into red-tone in 3-D reconstruction view.

### Histologic analysis

The sections were decalcified in 5% formic acid for 2 weeks after rinsed in sterile water, and they were dehydrated in a graded ethanol series and embedded in paraffin. Serial sections, 5 µm thick, were cut in a mesial-distal direction at intervals of 80 µm. Histologic specimens were observed under a light microscope (LEICA DM-LB, LEICA, WETZLAR, Germany) for evaluation of soft and hard tissue healing patterns, which include the degree of inflammation, bone and attachment gain.

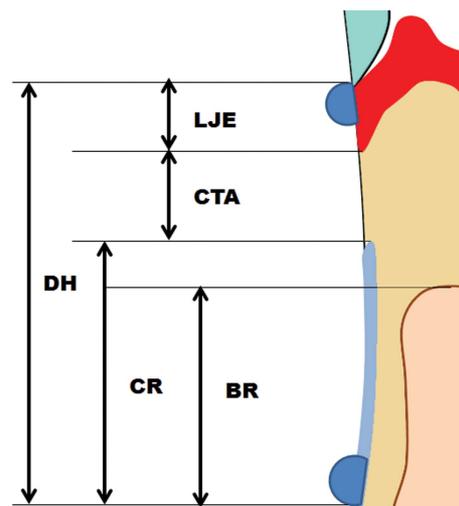
### Histometric analysis

Following parameters were analyzed by histometric analysis (Figure 2):

- Defect height (DH): distance from the cemento-enamel junction (CEJ) to the apical reference notch
- Long junctional epithelial attachment (LJE): distance from the CEJ to the apical extension of the junctional epithelium
- Connective tissue attachment (CTA): distance from the apical extension of the junctional epithelium to the coronal extension of new cementum
- Cementum regeneration (CR): distance from the coronal extension of new cementum or cementum-like substance to the apical reference notch
- Bone regeneration (BR): distance from the apical reference notch to the coronal extension of newly formed bone

### Statistical analysis

The means and standard deviations of the measurements



**Figure 2.** The schematic diagram of histometric measurement parameters. DH: Defect height; LJE: Long junctional epithelium; CTA: Connective tissue attachment; CR: Cementum regeneration; BR: Bone regeneration.

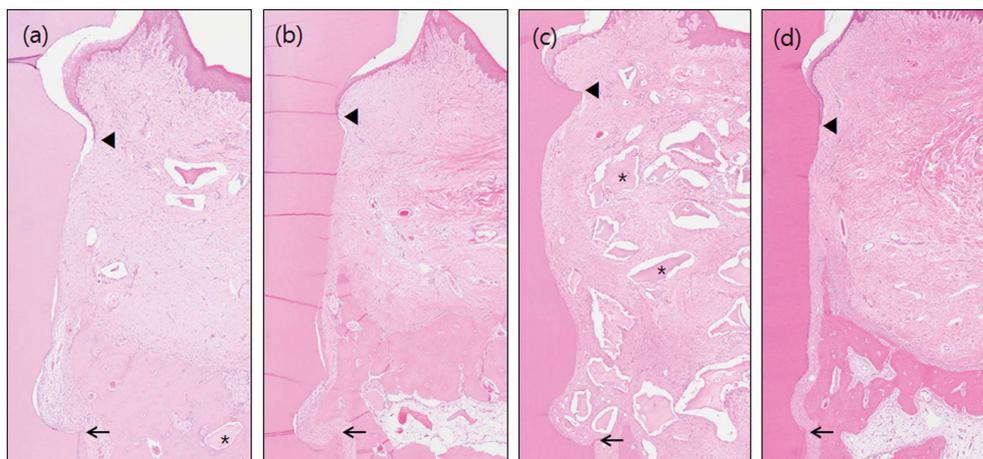
were analyzed descriptively for each group.

## Results

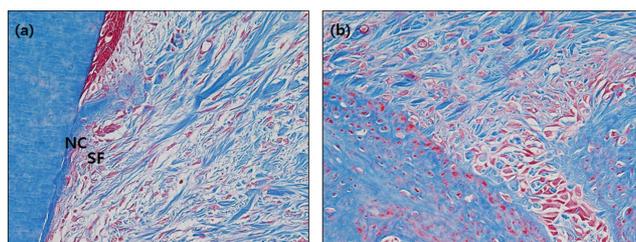
### Clinical observations and histologic analysis

The wound healing of defects was clinically uneventful except one defect. On the right side of dog #3, the sutures were loosened after the surgery and material was lost.

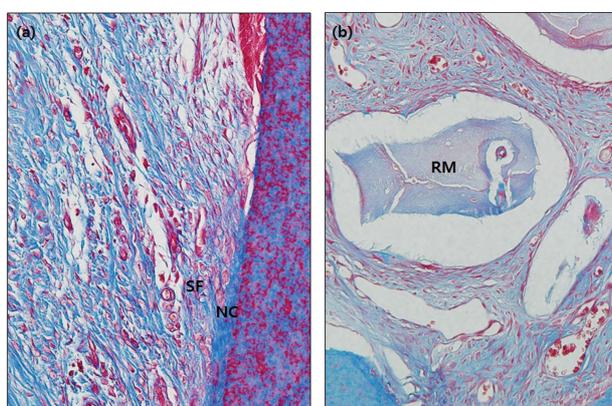
In common, regenerated woven bone was observed, and cementoid was created along the adjacent root surfaces with proliferation of cementoblasts. In some specimens, root resorption and associated osteoclasts were observed. In CBHA and EMD+CBHA groups, the residual bovine bone particles were found in the periodontal defect. No direct contact was



**Figure 3.** Histologic analysis (magnification  $\times 40$ ). New bone was formed beyond the bottom of the apical notch (arrows). Migration of junctional epithelium (arrowheads), connective tissue and vascularization was observed. Residual material (asterisks) was shown in CBHA and EMD+CBHA groups. (a) EMD+CBHA (b) EMD (c) CBHA (d) Control.



**Figure 4.** Histologic finding in EMD+CBHA group. Masson's trichrome stain (magnification  $\times 400$ ) (a) New cementum (NC) was formed along the root surface, and Sharpey's fiber (SF) was embedded into the newly formed cementum. Apical migration of junctional epithelium appeared to be blocked by NC. (b) New bone was formed, and inflammatory reaction was not observed.



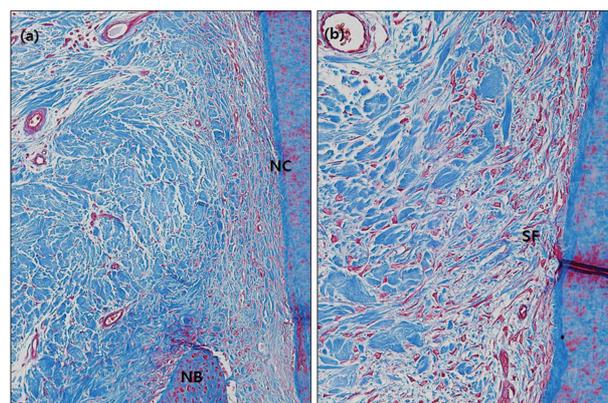
**Figure 5.** Histologic finding in CBHA group. Masson's trichrome stain (a) New cementum (NC) was formed along the root surface, and Sharpey's fiber (SF) was embedded into the newly formed cementum (magnification  $\times 400$ ). (b) Residual material (RM) was observed, without direct fibrous encapsulation or similar structures. Inflammatory reaction or other adverse effects were not observed (magnification  $\times 200$ ).

observed between residual particles and tooth surfaces. No remarkable difference was found between the occurrences of root resorption among the groups (Figure 3).

In Masson's trichrome stain, Sharpey's fiber was embedded into newly formed cementum. In EMD+CBHA groups, Sharpey's fiber was observed beyond the crest of new bone and along the newly formed cementum, and apical migration of junctional epithelium appeared to be blocked by new cementum (Figure 4). In CBHA groups, residual material was observed without direct fibrous encapsulation or similar structures. Sharpey's fiber was embedded into newly formed cementum, and inflammatory reaction or other adverse effects were not observed (Figure 5). Regenerated bone and cementum were observed in common including EMD group (Figure 6).

### Histometric analysis

Table 1 shows the histometric results of DH, LJE, CTA, CR, and BR. Neither difference nor tendency was observed in any



**Figure 6.** Histologic finding in EMD group. Masson's trichrome stain (a) New bone (NB) formation was observed, and new cementum (NC) was formed along the root surface (magnification  $\times 40$ ). (b) Sharpey's fiber (SF) was embedded into the newly formed cementum (magnification  $\times 400$ ).

**Table 1.** Histometric analysis

	EMD (n = 2)	EMD+Bio-oss (n = 2)	Bio-Oss (n = 5)	Control (n = 4)
LJE	1.22 $\pm$ 0.32	1.93 $\pm$ 0.37	1.82 $\pm$ 0.72	1.10 $\pm$ 0.24
CTA	0.39 $\pm$ 0.14	0.37 $\pm$ 0.35	0.23 $\pm$ 0.20	0.36 $\pm$ 0.30
CR	2.92 $\pm$ 0.23	2.26 $\pm$ 0.31	2.75 $\pm$ 0.74	2.92 $\pm$ 0.51
BR	1.75 $\pm$ 0.04	1.80 $\pm$ 0.09	2.06 $\pm$ 0.69	1.81 $\pm$ 0.15
DH	4.53 $\pm$ 0.25	4.55 $\pm$ 0.33	4.81 $\pm$ 0.18	4.38 $\pm$ 0.35

EMD, enamel matrix derivate; LJE, long junctional epithelial attachment; CTA, connective tissue attachment; CR, cementum regeneration; BR, bone regeneration; DH, defect height.

parameters among groups.

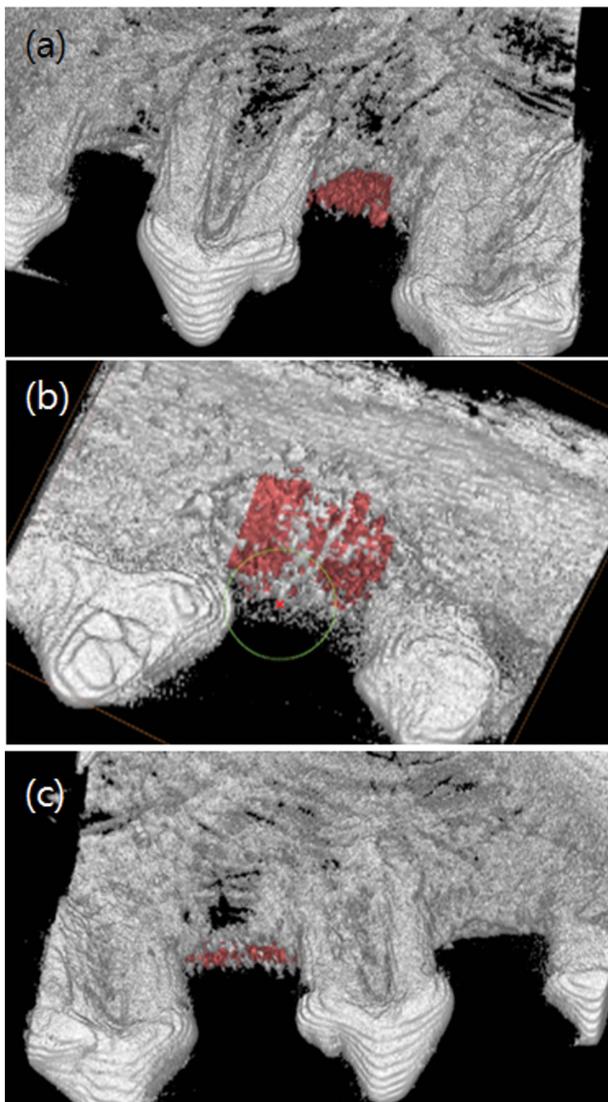
### Radiographic analysis

Residual material was color-coded in three-dimensional reconstruction view. No root resorption was observed in any groups and new bone was formed beyond the bottom notch of the defect, which enhances the histologic findings. In every defect where CBHA was applied with or without EMD, residual material was observed in the interproximal defect (Figure 7).

### Discussion

This study has shown the periodontal regeneration of combination therapy using EMD with CBHA in the periodontal interproximal defect model. All groups exhibited similar regeneration pattern and failed to reveal any differences when EMD and CBHA were combined.

The regenerative potential depends on the configuration of the defects. Previously, intrabony defects have been most investigated to assess the effect of EMD. The potential of regeneration depends on the number of bony walls because of wound stability and vascularization.<sup>48,49</sup> In contained defects



**Figure 7.** Residual material was color-coded into red-tone in (a, b) CBHA group (c) EMD+CBHA group.

such as 3-wall defects EMD alone showed significantly superior periodontal regeneration compared with control group and comparable regeneration with GTR procedure. However, non-contained defects have not shown consistent results. While in some studies EMD showed significantly higher periodontal regeneration than control in 1-wall and 2-wall defects,<sup>24,50-52</sup> other studies failed to show significant improvement of the probing depth and clinical attachment gain.<sup>30,53</sup> Histologic analysis in humans showed the evidence of periodontal regeneration in deep intrabony pockets but on an inconsistent basis.<sup>54</sup> Other types of non-contained defects have yielded much less favorable regeneration than in the intrabony defects. In the supraalveolar defects, the regeneration of new cementum was observed when EMD was applied in rats, but the authors failed to show the significant improvement over the control.<sup>55</sup> In the dehiscence defects of beagle

dogs, new bone formation was limited in the dehiscence portion, even though the newly formed cementum and connective tissue regeneration was observed in the EMD group.

The periodontal interproximal defect used in this study can be considered as a more unfavorable defect than a dehiscence or supraalveolar defect. Neither buccal nor lingual wall is not present, and two opposing root surfaces lack vascularization. Therefore, the periodontal interproximal defect can be recognized as a defect without a bony wall.<sup>56</sup> Since the combination therapy in the supraalveolar and dehiscence defect failed to show superior results over control, the more unfavorable defect can hardly exhibit better periodontal regeneration.

The combination therapy of BH and EMD provided the promising results in several studies but on an inconsistent basis. In vitro the combination of EMD with BH enhanced the attachment, proliferation, and differentiation of periodontal ligament cells and osteoblasts on BH particles.<sup>32</sup> In the human intrabony defects EMD showed the formation of connective tissue attachment in human histologic analysis although the configuration of the defect was not exactly specified.<sup>39</sup> In addition, several studies have supported that BH helps to augment the effect of EMDs in clinical attachment gain and probing depth reduction.<sup>33</sup> On the contrary, other studies reported that combination of EMD with BH failed to provide significant differences compared with bovine bone material alone in terms of attachment gain, probing depth, and BR<sup>37,40</sup> in the intrabony defects.

This study reported the results using CBHA with EMD in the periodontal interproximal without membrane. To improve the result of combination therapy, further studies with barrier membrane may be required. Animal studies presented complete healing only when barrier membrane was applied regardless of application of EMD or BH.<sup>57</sup> In the non-contained defects, the combination of BH with a bilayered collagen barrier demonstrated greater bone and cementum regeneration in human radiographic and histologic analysis.<sup>58</sup> Similar findings have been reported by several studies.<sup>59,60</sup> In a human clinical study, EMD alone showed less probing depth reduction and clinical attachment gain than guide tissue regeneration using non-resorbable titanium-reinforced membrane in non-contained defects.<sup>30</sup> Therefore, application of barrier membrane combined with EMD, BH and/or CBHA in the interproximal periodontal defect model might result in better results in radiographic or histologic analysis.

## Conclusion

Within the limitation of this study, EMD, CBHA, and combination of two materials showed similar periodontal regeneration in the interproximal periodontal defect model. Further investigation on combination with barrier membrane may be required for improvement of the regenerative potential.

## Acknowledgement

This study was supported by a grant of the Korea Health technology R&D Project, Ministry of Health & Welfare, Republic of Korea. (A120822)

## References

1. S. Nyman, J. Lindhe, T. Karring et al., "New attachment following surgical treatment of human periodontal disease," *J. Clin. Periodontol.*, **9**, 290-296 (1982).
2. J. Gottlow, S. Nyman, J. Lindhe et al., "New attachment formation in the human periodontium by guided tissue regeneration. Case reports," *J. Clin. Periodontol.*, **13**, 604-616 (1986).
3. R. G. Caffesse, B. A. Smith, W. A. Castelli et al., "New attachment achieved by guided tissue regeneration in beagle dogs," *J. Periodontol.*, **59**, 589-594 (1988).
4. J. Caton, C. Wagener, A. Polson et al., "Guided tissue regeneration in interproximal defects in the monkey," *Int. J. Periodontics Restorative Dent.*, **12**, 266-277 (1992).
5. A. Sculean, N. Donos, M. Brex et al., "Treatment of intrabony defects with guided tissue regeneration and enamel-matrix-proteins. An experimental study in monkeys," *J. Clin. Periodontol.*, **27**, 466-472 (2000).
6. R. G. Caffesse, L. E. Dominguez, C. E. Nasjleti et al., "Furcation defects in dogs treated by guided tissue regeneration (GTR)," *J. Periodontol.*, **61**, 45-50 (1990).
7. R. Niederman, E. D. Savitt, J. D. Heeley et al., "Regeneration of furca bone using Gore-Tex periodontal material," *Int. J. Periodontics Restorative Dent.*, **9**, 468-480 (1989).
8. J. Gottlow, T. Karring, and S. Nyman, "Guided tissue regeneration following treatment of recession-type defects in the monkey," *J. Periodontol.*, **61**, 680-685 (1990).
9. T. J. Sigurdsson, R. Hardwick, G. C. Bogle et al., "Periodontal repair in dogs: space provision by reinforced ePTFE membranes enhances bone and cementum regeneration in large supraalveolar defects," *J. Periodontol.*, **65**, 350-356 (1994).
10. A. Sculean, A. Stavropoulos, P. Windisch et al., "Healing of human intrabony defects following regenerative periodontal therapy with a bovine-derived xenograft and guided tissue regeneration," *Clin. Oral Investig.*, **8**, 70-74 (2004).
11. P. Cortellini, C. Clauser, and G. P. Prato, "Histologic assessment of new attachment following the treatment of a human buccal recession by means of a guided tissue regeneration procedure," *J. Periodontol.*, **64**, 387-391 (1993).
12. P. Cortellini, G. Pini Prato, and M. S. Tonetti, "Periodontal regeneration of human intrabony defects with titanium reinforced membranes. A controlled clinical trial," *J. Periodontol.*, **66**, 797-803 (1995).
13. P. Cortellini, G. Pini Prato, and M. S. Tonetti, "Periodontal regeneration of human intrabony defects with bioresorbable membranes. A controlled clinical trial," *J. Periodontol.*, **67**, 217-223 (1996).
14. M. S. Tonetti, P. Cortellini, J. E. Suvan et al., "Generalizability of the added benefits of guided tissue regeneration in the treatment of deep intrabony defects. Evaluation in a multi-center randomized controlled clinical trial," *J. Periodontol.*, **69**, 1183-1192 (1998).
15. N. Blumenthal and J. Steinberg, "The use of collagen membrane barriers in conjunction with combined demineralized bone-collagen gel implants in human intrabony defects," *J. Periodontol.*, **61**, 319-327 (1990).
16. P. Ratka-Kruger, E. Neukranz, and P. Raetzke, "Guided tissue regeneration procedure with bioresorbable membranes versus conventional flap surgery in the treatment of intrabony periodontal defects," *J. Clin. Periodontol.*, **27**, 120-127 (2000).
17. S. Gestrelus, C. Andersson, D. Lidstrom et al., "In vitro studies on periodontal ligament cells and enamel matrix derivative," *J. Clin. Periodontol.*, **24**, 685-692 (1997).
18. L. Hammarstrom, L. Heijl, and S. Gestrelus, "Periodontal regeneration in a buccal dehiscence model in monkeys after application of enamel matrix proteins," *J. Clin. Periodontol.*, **24**, 669-677 (1997).
19. S. Gestrelus, C. Andersson, A. C. Johansson et al., "Formulation of enamel matrix derivative for surface coating. Kinetics and cell colonization," *J. Clin. Periodontol.*, **24**, 678-684 (1997).
20. P. F. Regazzini, A. B. Novaes, Jr., P. T. de Oliveira et al., "Comparative study of enamel matrix derivative with or without GTR in the treatment of class II furcation lesions in dogs," *Int. J. Periodontics Restorative Dent.*, **24**, 476-487 (2004).
21. E. A. Sallum, S. P. Pimentel, J. B. Saldanha et al., "Enamel matrix derivative and guided tissue regeneration in the treatment of dehiscence-type defects: a histomorphometric study in dogs," *J. Periodontol.*, **75**, 1357-1363 (2004).
22. K. Alhezaimi, T. Al-Shalan, R. O'Neill et al., "Connective tissue-cementum regeneration: a new histologic regeneration following the use of enamel matrix derivative in dehiscence-type defects. A dog model," *Int. J. Periodontics Restorative Dent.*, **29**, 425-433 (2009).
23. A. Parashis and K. Tsiklakis, "Clinical and radiographic findings following application of enamel matrix derivative in the treatment of intrabony defects. A series of case reports," *J. Clin. Periodontol.*, **27**, 705-713 (2000).
24. L. Heijl, G. Heden, G. Svardstrom et al., "Enamel matrix derivative (EMDOGAIN) in the treatment of intrabony periodontal defects," *J. Clin. Periodontol.*, **24**, 705-714 (1997).
25. G. Heden, "A case report study of 72 consecutive Emdogain-treated intrabony periodontal defects: clinical and radiographic findings after 1 year," *Int. J. Periodontics Restorative Dent.*, **20**, 127-139 (2000).
26. G. Heden, J. Wennstrom, and J. Lindhe, "Periodontal tissue alterations following Emdogain treatment of periodontal sites with angular bone defects. A series of case reports," *J. Clin. Periodontol.*, **26**, 855-860 (1999).
27. M. S. Tonetti, N. P. Lang, P. Cortellini et al., "Enamel matrix proteins in the regenerative therapy of deep intrabony defects," *J. Clin. Periodontol.*, **29**, 317-325 (2002).
28. S. Jepsen, B. Heinz, K. Jepsen et al., "A randomized clinical trial comparing enamel matrix derivative and membrane treatment of buccal Class II furcation involvement in mandibular molars. Part I: Study design and results for primary outcomes," *J. Periodontol.*, **75**, 1150-1160 (2004).
29. J. Meyle, J. R. Gonzales, R. H. Bodeker et al., "A randomized clinical trial comparing enamel matrix derivative and membrane treatment of buccal class II furcation involvement in mandibular molars. Part II: secondary outcomes," *J. Periodontol.*, **75**, 1188-1195 (2004).
30. V. I. Siciliano, G. Andreuccetti, A. I. Siciliano et al., "Clinical outcomes after treatment of non-contained intrabony defects with enamel matrix derivative or guided tissue regeneration: a 12-month randomized controlled clinical trial," *J. Periodontol.*, **82**, 62-71 (2011).
31. N. Donos, L. Glavind, T. Karring et al., "Clinical evaluation of an enamel matrix derivative and a bioresorbable membrane in the treatment of degree III mandibular furcation involvement: a

- series of nine patients," *Int. J. Periodontics Restorative Dent.*, **24**, 362-369 (2004).
32. R. J. Miron, D. D. Bosshardt, E. Hedbom et al., "Adsorption of enamel matrix proteins to a bovine-derived bone grafting material and its regulation of cell adhesion, proliferation, and differentiation," *J. Periodontol.*, **83**, 936-947 (2012).
  33. V. Lekovic, P. M. Camargo, M. Weinlaender et al., "A comparison between enamel matrix proteins used alone or in combination with bovine porous bone mineral in the treatment of intrabony periodontal defects in humans," *J. Periodontol.*, **71**, 1110-1116 (2000).
  34. D. Velasquez-Plata, E. T. Scheyer, and J. T. Mellonig, "Clinical comparison of an enamel matrix derivative used alone or in combination with a bovine-derived xenograft for the treatment of periodontal osseous defects in humans," *J. Periodontol.*, **73**, 433-440 (2002).
  35. P. M. Camargo, V. Lekovic, M. Weinlaender et al., "The effectiveness of enamel matrix proteins used in combination with bovine porous bone mineral in the treatment of intrabony defects in humans," *J. Clin. Periodontol.*, **28**, 1016-1022 (2001).
  36. V. Lekovic, P. M. Camargo, M. Weinlaender et al., "The use of bovine porous bone mineral in combination with enamel matrix proteins or with an autologous fibrinogen/fibronectin system in the treatment of intrabony periodontal defects in humans," *J. Periodontol.*, **72**, 1157-1163 (2001).
  37. E. T. Scheyer, D. Velasquez-Plata, M. A. Brunsvold et al., "A clinical comparison of a bovine-derived xenograft used alone and in combination with enamel matrix derivative for the treatment of periodontal osseous defects in humans," *J. Periodontol.*, **73**, 423-432 (2002).
  38. S. Yamamoto, H. Masuda, Y. Shibukawa et al., "Combination of bovine-derived xenografts and enamel matrix derivative in the treatment of intrabony periodontal defects in dogs," *Int. J. Periodontics Restorative Dent.*, **27**, 471-479 (2007).
  39. A. Sculean, P. Windisch, T. Keglevich et al., "Clinical and histologic evaluation of human intrabony defects treated with an enamel matrix protein derivative combined with a bovine-derived xenograft," *Int. J. Periodontics Restorative Dent.*, **23**, 47-55 (2003).
  40. A. Sculean, G. C. Chiantella, P. Windisch et al., "Clinical evaluation of an enamel matrix protein derivative (Emdogain) combined with a bovine-derived xenograft (Bio-Oss) for the treatment of intrabony periodontal defects in humans," *Int. J. Periodontics Restorative Dent.*, **22**, 259-267 (2002).
  41. G. Zucchelli, C. Amore, L. Montebugnoli et al., "Enamel matrix proteins and bovine porous bone mineral in the treatment of intrabony defects: a comparative controlled clinical trial," *J. Periodontol.*, **74**, 1725-1735 (2003).
  42. M. L. Nevins, M. Camelo, S. E. Lynch et al., "Evaluation of periodontal regeneration following grafting intrabony defects with bio-oss collagen: a human histologic report," *Int. J. Periodontics Restorative Dent.*, **23**, 9-17 (2003).
  43. M. L. Nevins, M. Camelo, A. Rebaudi et al., "Three-dimensional micro-computed tomographic evaluation of periodontal regeneration: a human report of intrabony defects treated with Bio-Oss collagen," *Int. J. Periodontics Restorative Dent.*, **25**, 365-373 (2005).
  44. A. Sculean, G. C. Chiantella, P. Windisch et al., "Healing of intrabony defects following treatment with a composite bovine-derived xenograft (Bio-Oss Collagen) in combination with a collagen membrane (Bio-Gide PERIO)," *J. Clin. Periodontol.*, **32**, 720-724 (2005).
  45. G. A. Hartman, R. M. Arnold, M. P. Mills et al., "Clinical and histologic evaluation of anorganic bovine bone collagen with or without a collagen barrier," *Int. J. Periodontics Restorative Dent.*, **24**, 127-135 (2004).
  46. G. Cardaropoli, M. Araujo, R. Hayacibara et al., "Healing of extraction sockets and surgically produced - augmented and non-augmented - defects in the alveolar ridge. An experimental study in the dog," *J. Clin. Periodontol.*, **32**, 435-440 (2005).
  47. M. G. Araujo and J. Lindhe, "Ridge preservation with the use of Bio-Oss collagen: A 6-month study in the dog," *Clin. Oral Implants Res.*, **20**, 433-440 (2009).
  48. U. M. Wikesjo and K. A. Selvig, "Periodontal wound healing and regeneration," *Periodontol.* **2000**, **19**, 21-39 (1999).
  49. C. S. Kim, S. H. Choi, J. K. Chai et al., "Periodontal repair in surgically created intrabony defects in dogs: influence of the number of bone walls on healing response," *J. Periodontol.*, **75**, 229-235 (2004).
  50. O. Zetterstrom, C. Andersson, L. Eriksson et al., "Clinical safety of enamel matrix derivative (EMDOGAIN) in the treatment of periodontal defects," *J. Clin. Periodontol.*, **24**, 697-704 (1997).
  51. A. Sculean, N. Donos, A. Miliuskaite et al., "Treatment of intrabony defects with enamel matrix proteins or bioabsorbable membranes. A 4-year follow-up split-mouth study," *J. Periodontol.*, **72**, 1695-1701 (2001).
  52. A. Sculean, N. Donos, A. Blaes et al., "Comparison of enamel matrix proteins and bioabsorbable membranes in the treatment of intrabony periodontal defects. A split-mouth study," *J. Periodontol.*, **70**, 255-262 (1999).
  53. P. Proussaefs, J. Lozada, A. Kleinman et al., "The use of ramus autogenous block grafts for vertical alveolar ridge augmentation and implant placement: a pilot study," *Int. J. Oral Maxillofac. Implants*, **17**, 238-248 (2002).
  54. R. A. Yukna and J. T. Mellonig, "Histologic evaluation of periodontal healing in humans following regenerative therapy with enamel matrix derivative. A 10-case series," *J. Periodontol.*, **71**, 752-759 (2000).
  55. C. E. Nemcovsky, S. Zahavi, O. Moses et al., "Effect of enamel matrix protein derivative on healing of surgical supra-infrabony periodontal defects in the rat molar: a histomorphometric study," *J. Periodontol.*, **77**, 996-1002 (2006).
  56. U. W. Jung, Y. Y. Chang, Y. J. Um et al., "Interproximal periodontal defect model in dogs: a pilot study," *Oral Dis.*, **17**, 26-32 (2011).
  57. L. Ritter, J. Neugebauer, R. A. Mischkowski et al., "Evaluation of the course of the inferior alveolar nerve in the mental foramen by cone beam computed tomography," *Int. J. Oral Maxillofac. Implants*, **27**, 1014-1021 (2012).
  58. M. Camelo, M. L. Nevins, R. K. Schenk et al., "Clinical, radiographic, and histologic evaluation of human periodontal defects treated with Bio-Oss and Bio-Gide," *Int. J. Periodontics Restorative Dent.*, **18**, 321-331 (1998).
  59. J. T. Mellonig, "Human histologic evaluation of a bovine-derived bone xenograft in the treatment of periodontal osseous defects," *Int. J. Periodontics Restorative Dent.*, **20**, 19-29 (2000).
  60. M. Camelo, M. L. Nevins, S. E. Lynch et al., "Periodontal regeneration with an autogenous bone-Bio-Oss composite graft and a Bio-Gide membrane," *Int. J. Periodontics Restorative Dent.*, **21**, 109-119 (2001).