



저작자표시-비영리-변경금지 2.0 대한민국

이용자는 아래의 조건을 따르는 경우에 한하여 자유롭게

- 이 저작물을 복제, 배포, 전송, 전시, 공연 및 방송할 수 있습니다.

다음과 같은 조건을 따라야 합니다:



저작자표시. 귀하는 원저작자를 표시하여야 합니다.



비영리. 귀하는 이 저작물을 영리 목적으로 이용할 수 없습니다.



변경금지. 귀하는 이 저작물을 개작, 변형 또는 가공할 수 없습니다.

- 귀하는, 이 저작물의 재이용이나 배포의 경우, 이 저작물에 적용된 이용허락조건을 명확하게 나타내어야 합니다.
- 저작권자로부터 별도의 허가를 받으면 이러한 조건들은 적용되지 않습니다.

저작권법에 따른 이용자의 권리는 위의 내용에 의하여 영향을 받지 않습니다.

이것은 [이용허락규약\(Legal Code\)](#)을 이해하기 쉽게 요약한 것입니다.

[Disclaimer](#)

More than 6 times of pretransplant therapeutic plasma
exchange increase the recurrence of hepatocellular
carcinoma in ABO incompatible living donor liver
transplantation

Young Jin Yoo

The Graduate School
Yonsei University
Department of Medicine

More than 6 times of pretransplant therapeutic plasma
exchange increase the recurrence of hepatocellular
carcinoma in ABO incompatible living donor liver
transplantation

A Master's Thesis Submitted
to the Department of Medicine
and the Graduate School of Yonsei University
in partial fulfillment of the
requirements for the degree of
Master of Medical Science

Young Jin Yoo

December 2024

**This certifies that the Master's Thesis
of Young Jin Yoo is approved**

Thesis Supervisor _____
Dong Jin Joo

Thesis Committee Member _____
Jun Young Park

Thesis Committee Member _____
Sinyoung Kim

**The Graduate School
Yonsei University
December 2024**

Acknowledgements

I would like to express my deepest gratitude to my advisor, Professor Dong Jin Joo, for guiding me in initiating and completing this research. He introduced me to the field of transplant surgery and inspired me profoundly with his relentless dedication to research, tireless problem-solving, and active engagement in clinical practice. It was through his influence that I was able to embark on and complete this study. I am also sincerely thankful to Professor Jun Young Park, who served as the chair of the thesis committee, for his fair and objective evaluation, which greatly contributed to improving the quality of this thesis. Additionally, I extend my heartfelt thanks to Professor Sinyoung Kim, a committee member, for enriching the content and details of this thesis through his diverse perspectives beyond clinical practice.

I am very grateful to Professor Kyu Ha Heo, Chief of the Division of Transplant Surgery, and Professor Myoung Soo Kim, Director of the Research Institute for Transplantation, for their dedication to advancing the field of transplant surgery, which has made this research possible. My special thanks also go to Professor Jae Geun Lee, Professor Deok-Gie Kim, and Professor Eun-Ki Min of the liver transplant division for their active involvement in both clinical practice and research, which has greatly supported this study.

I would especially like to thank my parents for their support. They provided endless love and the chance to continue studying. Lastly, I would like to share my pleasure and happiness upon finishing this work with my lovely wife and daughter. This thesis could not have been completed without the support of my wife Ji Hye Hong and my daughter Somin. Their encouragement has been indispensable in enabling me to pursue active research and clinical work.

30th of December, 2024

TABLE OF CONTENTS

LIST OF FIGURES	ii
LIST OF TABLES	iii
ABSTRACT IN ENGLISH	iv
1. INTRODUCTION.....	1
2. METHODS.....	2
2.1. Study material	2
2.2. Data collection and outcomes	4
2.3. Pretransplant desensitization for ABO incompatibility	4
2.4. Statistical methods	7
2.5. Statement of Ethics	7
3. RESULTS	8
3.1. Baseline characteristics.....	8
3.2. Detailed information on recipient of ABOi LDLT	10
3.3. HCC outcomes	12
3.4. Subgroup analysis for HCC recurrence	15
4. DISCUSSION	18
5. CONCLUSIONS	22
REFERENCES	23
APPENDICES	28
ABSTRACT IN KOREAN	32

LIST OF FIGURES

<Fig 1> Study population	2
<Fig 2> Cubic spline model for assessing proper pretransplant TPE number cutoff	3
<Fig 3> The modified desensitization protocol at YUHS for ABOi LDLT, since 2012.....	5
<Fig 4> The modified desensitization protocol at YUHS for ABOi LDLT, since 2019.....	6
<Fig 5> Kaplan-Meier curve of RFS and HCC recurrence according to ABO incompatibility and plasma exchange numbers	12
<Fig 6> Kaplan-Meier curve of RFS and HCC recurrence in ABOi subgroup, according to plasma exchange numbers	13
<Fig 7> HCC recurrence in ABOi subgroup, according to plasma exchange numbers and MORAL score	17
<Fig 8> Therapeutic plasma exchange and tumor progression flowchart	21

LIST OF TABLES

<Table 1> Baseline characteristics of patients, according to ABO incompatibility and the number of pretransplant therapeutic plasma exchange	9
<Table 2> Details for ABO incompatibility and desensitization of ABO incompatible group patients, according to therapeutic plasma exchange number	11
<Table 3> Multivariable Cox analysis for recurrence free survival and hepatocellular carcinoma recurrence	14
<Table 4> Subgroup analysis of 5-year hepatocellular carcinoma recurrence according to therapeutic plasma exchange numbers in ABO incompatible group	16
<Table 5> Previous studies regarding pretransplant desensitization and cancer risk	19

Abstract in English

More than 6 times of pretransplant therapeutic plasma exchange increase the recurrence of hepatocellular carcinoma in ABO incompatible living donor liver transplantation

Background

Previous studies have reported comparable oncologic outcome between ABO incompatible (ABOi) living donor liver transplantation (LDLT) and ABO compatible (ABOc) LDLT in hepatocellular carcinoma (HCC) patients. We aimed to analyze the relationship between the number of therapeutic plasma exchanges (TPE) and HCC outcomes in ABOi LDLT.

Methods

In this single center retrospective study, 428 LDLT recipients with HCC were categorized into three groups according to ABO incompatibility and number of pretransplant TPE, of which cutoff was more than 6 times, determined from cubic spline model for recurrence free survival (RFS): ABOc (n=323), ABOi/TPE<6 (n=75), and ABOi/TPE≥6 (n=30). The RFS and HCC recurrence rates were compared after adjusting for other risk factors for HCC outcomes.

Results

The three groups showed similar characteristics in most demographics, tumor markers and pathologies. The median initial isoagglutinin (IA) titer was 1:64 (range negative-1:512) in the ABOi/TPE<6 group and 1:512 (range 1:128-1:4096) in the ABOi/TPE≥6 group. Five year RFS was significantly lower (75.7% in the ABOc vs. 72.7% in the ABOi/TPE<6 vs. 50.0% in the ABOi/TPE≥6, P=0.005) and HCC recurrence was significantly higher in ABOi/TPE≥6 group than in the other groups (16.4% vs. 17.0% vs. 39.4%, P=0.014). In multivariable Cox regression analysis, ABOi/TPE≥6 was an independent risk factor for RFS (aHR 1.99 (1.02-3.86), P=0.042) and HCC recurrence (aHR 2.42 (1.05-5.57), P=0.037).

Conclusion

More than six pretransplant TPE have the potential for higher HCC recurrence after ABOi LDLT.

A strategy to reduce the number of TPE to less than six would be needed when planning ABOi LDLT for HCC patients, ensuring similar immunologic risk.

Key words : ABO incompatible living donor liver transplantation, Hepatocellular carcinoma, Plasma exchange, Oncologic outcome

I. INTRODUCTION

Liver transplantation (LT) is an effective, and sometimes the only, treatment option for unresectable hepatocellular carcinoma (HCC). However, owing to organ shortages, not all patients can receive timely LT. Consequently, the demand for living donor liver transplantation (LDLT) for HCC is increasing worldwide, and numerous studies have reported comparable oncological outcomes between LDLT and deceased donor liver transplantation (DDLT). (Akamatsu et al., 2014; Azoulay et al., 2017; Goldaracena & Barbas, 2019; Lai et al., 2021; Lee, 2015; Ninomiya et al., 2015; Ogawa & Takada, 2016)

When an ABO incompatible (ABOi) living donor is the only available option, ABO incompatible living donor liver transplantation (ABOi LDLT) with proper desensitization becomes a viable choice. (Egawa et al., 2014; Gordon et al., 1986; Gugenheim et al., 1990; Kim et al., 2013; C.-F. Lee et al., 2015; Lee et al., 2014; Matsuno et al., 2008; Tanabe et al., 2002) Despite the need for pretransplant antibody treatment and an increased risk of posttransplant infections, ABOi LDLT has been reported as a feasible treatment for patients with end-stage liver disease, offering substantial survival benefits even for those with high Model for End-Stage Liver Disease (MELD) scores. (Egawa et al., 2014; Kim et al., 2013; Lee et al., 2022; Yim et al., 2023) Additionally, several Korean centers have reported that ABOi LDLT has a similar impact on HCC outcomes compared to ABO compatible (ABOc) LDLT (ABOc LDLT). (Kang et al., 2019; Kim et al., 2018; Kim et al., 2013; Kim et al., 2019; Yoon et al., 2018)

Despite these reports, ABOi LDLT necessitates more potent immunosuppression, including B-cell depleting agents, therapeutic plasma exchange (TPE), and higher maintenance immunosuppressants, which raises concerns about potentially adverse oncologic outcomes. (S. D. Lee et al., 2015; Miyagi et al., 2012) Furthermore, ABOi LDLT requires additional pretransplant TPE sessions as the titer of blood group antibodies increases. However, there are no published studies examining the differences in HCC outcomes based on the degree of desensitization required.

Therefore, this study aimed to analyze the effect of the number of pretransplant TPE sessions, a critical component of pretransplant treatment, on HCC outcomes in ABOi LDLT.

II. METHODS

2.1. Study material

In this retrospective cohort study, we analyzed single center data from 466 patients who underwent LDLT for HCC between January 2011, when ABOi LDLT was initiated at our institution, and December 2022. The baseline characteristics and details of explant pathology were retrieved from a prospectively collected institutional database. The exclusion criteria were as follows: mixed cholangiocellular carcinoma on pathology (n=29), liver cancer other than HCC (n=2), LDLT from a dual living donor (n=3), and missing data (n=4) (Figure 1).

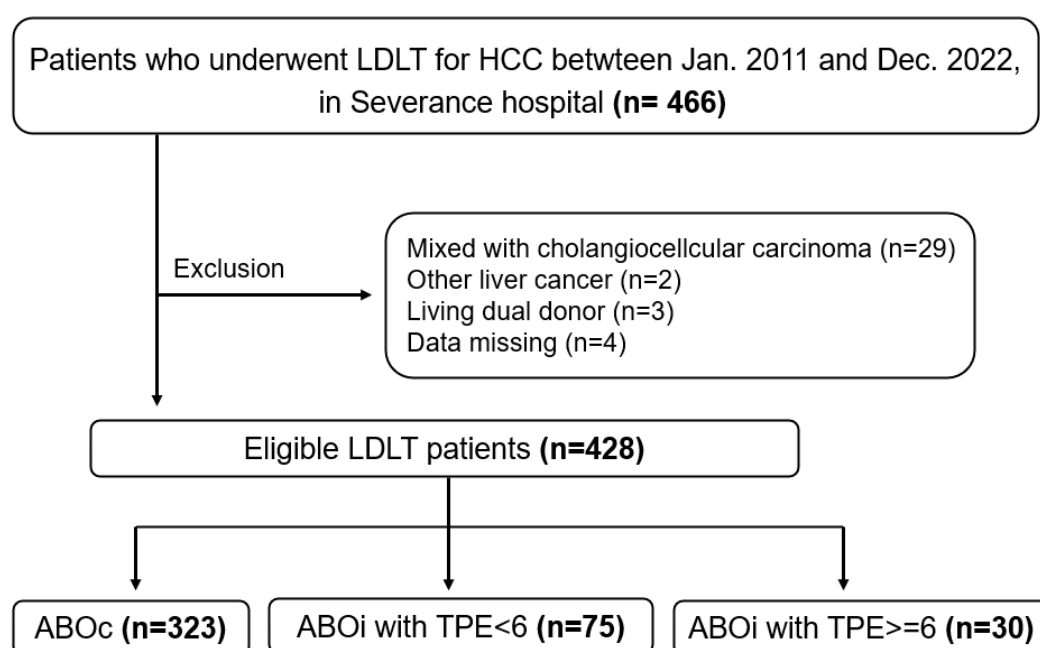


Figure 1. Study population

LDLT, living donor liver transplantation; HCC, hepatocellular carcinoma; ABOc, ABO compatible; ABOi, ABO incompatible; TPE, therapeutic plasma exchange.

A total of 428 eligible patients were categorized according to ABO incompatibility and the number of pretransplant TPE sessions: ABO compatible (ABOc group, n=323, 75.5%), ABO incompatible with fewer than 5 TPE sessions (ABOi/TPE ≤ 5 group, n=75, 17.5%), and ABO incompatible with 6 or more TPE sessions (ABOi/TPE ≥ 6 group, n=30, 7.5%). The cutoff for the number of TPE sessions (6 times) was determined based on the spline curve for recurrence free survival (RFS), where the hazard began to significantly increase (Figures 2).

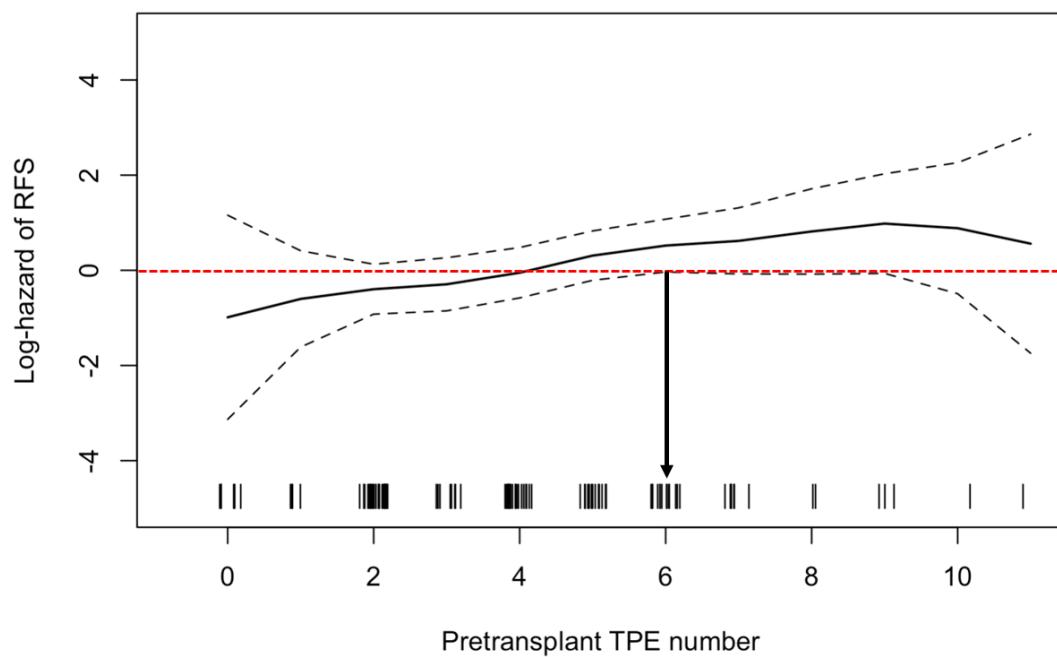


Figure 2. Cubic spline model for assessing proper pretransplant TPE number cutoff
 RFS; recurrence free survival, TPE; plasma exchange.

2.2. Data collection and outcomes

All relevant information regarding recipients, donors, and LDLT surgery was retrieved from the institutional database. The underlying liver diseases associated with HCC included hepatitis B, hepatitis C, and non-B/non-C. Detailed information on explant pathology and tumor markers, such as alpha-fetoprotein (AFP) and protein induced by vitamin K absence or antagonist-II (PIVKA-II) at the time of LDLT, was obtained. Additionally, data on pretransplant locoregional and systemic treatments, as well as previous hepatectomies, were collected for patients with HCC. RFS and HCC recurrence (time to recurrence) were the primary outcomes.

2.3. Pretransplant desensitization for ABO incompatibility

Our institutional protocol for desensitization in ABOi LDLT mainly consisted of rituximab and TPE, as described previously. (Choi et al., 2024; J. Lee et al., 2015) A recently revised version of this protocol is provided in Figure 3 and Figure 4 (Desensitization protocol for ABOi LDLT).

For the initial and target isoagglutinin (IA) titers, higher IgM or IgG anti-A/B titers were employed. The number of preoperative TPE sessions was determined based on the initial IA titer, the response to TPE, and the decrease in the ABO titer. Splenectomy and postoperative TPE were performed in patients at high risk of rejection, specifically those with an IA titer greater than 1:64 at the time of LT. Additional rounds of TPE were conducted postoperatively in cases of clinical rejection or IA titer rebound, defined as a resurgence to 1:64 and a minimum two-fold increase. Following TPE, intravenous immunoglobulin (IVIG) was administered at a dose of 500–800 mg/kg on an individualized basis, depending on ABO antibody levels and infection risk.

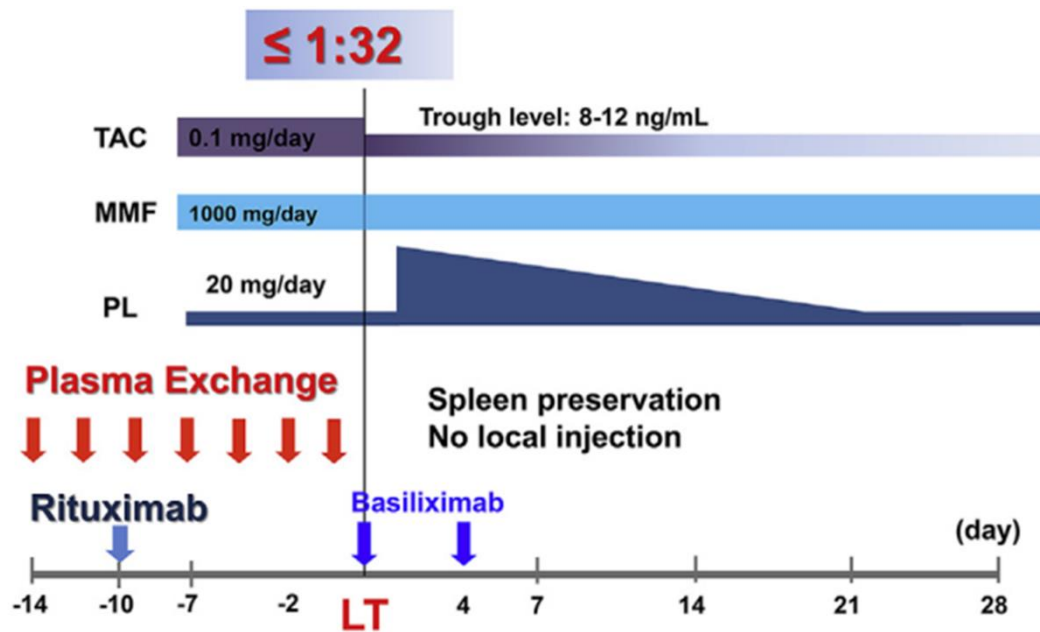


Figure 3. The modified desensitization protocol at YUHS for ABOi LDLT, since 2012

YUHS, Yonsei University Health System; ABOi, ABO incompatible; LDLT, living donor liver transplantation; TAC, tacrolimus; MMF, mycophenolate mofetil; PL, prednisolone; LT, liver transplantation.

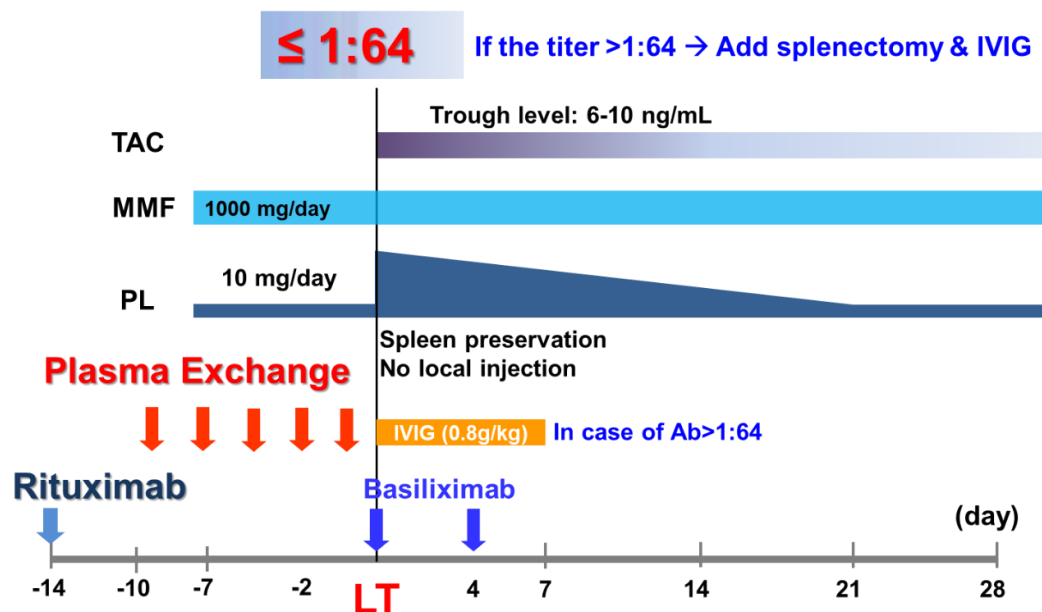


Figure 4. The modified desensitization protocol at YUHS for ABOi LDLT, since 2019

YUHS, Yonsei University Health System; ABOi, ABO incompatible; LDLT, living donor liver transplantation; IVIG, intravenous immunoglobulin; LT, liver transplantation; TAC, tacrolimus; MMF, mycophenolate mofetil; PL, prednisolone; PGE-1, prostaglandin E1.

2.4. Statistical analysis

Depending on the type of variable, data were presented either as numbers (percentages) or as medians (interquartile range [IQR]). The Mann-Whitney U test or chi-square test was used to compare continuous and categorical variables, respectively, when appropriate. HCC outcomes were analyzed using Kaplan-Meier curves and log-rank tests. Multivariable Cox regression was performed to evaluate HCC outcomes in the entire population, including covariates with significant P values <0.1 from the univariable analysis. In the risk analysis of HCC recurrence, non-HCC death was considered a competing risk, utilizing the Fine and Gray method (Fine & Gray, 1999) for competing risk regression. In the ABOi LDLT groups, the 5-year estimates of HCC recurrence were compared based on the number of TPE sessions (<6 vs. ≥ 6) across various subgroups categorized by tumor burden, which reflects the tumor size, tumor number, and AFP and PIVKA-II levels (Duvoux et al., 2012; Lee et al., 2016; Mazzaferro et al., 2009; Mazzaferro et al.), as well as ABO antibody strength, postoperative rebound of IA titer and TPE, and splenectomy status. Subgroup analyses were conducted in a univariate manner due to the small size of each group. All statistical analyses were performed using the R statistical package, version 4.3.0 for macOS (<http://cran.r-project.org/>), with the significance threshold set at $P < 0.05$.

2.5. Statement of ethics

This study was performed in accordance with the Declaration of Helsinki and the Declaration of Istanbul and was approved by the Institutional Review Board of Severance Hospital, Yonsei University Health System (IRB number 4-2024-0977). The requirement of informed consent was waived due to the retrospective nature of the study.

III. RESULTS

3.1. Baseline characteristics

No significant difference was noted in most baseline patient characteristics (Table 1). The distribution of LT years was also not statistically significant ($P = 0.069$); however, a higher proportion of transplants in the ABOi groups occurred between 2016 and 2019. Most patients had hepatitis B as the underlying cause of HCC across all groups, with no statistical significance (76.8% in ABOc, 69.3% in ABOi/TPE<6, and 86.7% in ABOi/TPE \geq 6, $P = 0.401$). Notably, the ABOi/TPE \geq 6 group required a significantly higher number of red blood cell transfusions (median 4.5 packs) than the ABOc and ABOi/TPE<6 groups (median 2 packs, $P = 0.014$). No significant differences were noted in the pretransplant AFP and PIVKA-II levels. Additionally, history of hepatectomy, locoregional therapy (LRT), and systemic treatment were similar across the groups.

Most characteristics from explant pathology were similar across the groups, including the incidence of portal vein tumor thrombosis (PVTT), total necrosis, number of viable tumors, maximum tumor size, microvascular invasion, and poor differentiation. However, the presence of satellite nodules was significantly higher in the ABOi/TPE \geq 6 group (26.7%) than in the other groups (10.8% in ABOc and 10.7% in the ABOi/TPE<6, $P = 0.035$).

Table 1. Baseline characteristics of patients, according to ABO incompatibility and the number of pretransplant therapeutic plasma exchange

	ABOc (n=323)	ABOi/TPE<6 (n=75)	ABOi/TPE≥6 (n=30)	<i>P</i>
Age, years	56.8 ± 7.0	57.1 ± 6.9	55.6 ± 7.3	0.608
Sex, female	58 (18.0)	17 (22.7)	7 (23.3)	0.539
BMI	23.8 (22.3-26.1)	24.9 (23.4-26.3)	24.0 (21.9-25.9)	0.065
LT year				0.069
2011-2015	112 (34.7)	15 (20.0)	7 (23.3)	
2016-2019	104 (32.2)	33 (44.0)	14 (46.7)	
2020-2022	107 (33.1)	27 (36.0)	9 (30.0)	
Underlying liver disease for HCC				0.401
Hepatitis B	248 (76.8)	52 (69.3)	26 (86.7)	
Hepatitis C	23 (7.1)	8 (10.7)	1 (3.3)	
Non-B, Non-C	52 (16.1)	15 (20.0)	3 (10.0)	
Hypertension	74 (22.9)	21 (28.0)	9 (30.0)	0.490
Diabetes mellitus	97 (30.0)	27 (36.0)	12 (40.0)	0.367
Pretransplant MELD	10 (8-14)	10 (8-13)	11.5 (8-14)	0.674
Donor age, years	31 (24-40)	34 (26-40.5)	35 (25-41)	0.203
Donor sex, female	130 (40.2)	26 (34.7)	11 (36.7)	0.647
GRWR [†] < 0.8	27 (8.4)	5 (6.7)	2 (6.7)	0.856
Macrovesicular steatosis ≥ 10%	46 (15.2)	9 (12.3)	2 (6.9)	0.422
Cold ischemic time, min	126 (106-150)	126 (102-152.5)	128.5 (96-180)	0.884
Transfusion RBC, packs	2 (0-6)	2 (0-7.5)	4.5 (2-9)	0.014
AFP at LT, ng/mL	6.6 (3.3-23.1)	6.4 (3.3-14.0)	4.3 (2.2-27.2)	0.545
PIVKA at LT, mAU/mL	38 (22-112)	38 (23.5-141)	47 (20-232)	0.666
Hepatectomy history	62 (19.2)	13 (17.3)	7 (23.3)	0.779
Pretransplant LRT	246 (76.2)	59 (78.7)	23 (76.7)	0.899
Systemic treatment	45 (13.9)	9 (12.0)	5 (16.7)	0.812
Explant pathology				
Total necrosis	57 (17.6)	13 (17.3)	4 (13.3)	0.836
Viable tumor number	1 (1-3)	2 (1-3)	2 (1-3)	0.485
Maximum tumor size, cm	1.7 (1.0-3.0)	1.8 (0.8-3.0)	2.4 (1.3-3.7)	0.139

Microvascular invasion	76 (23.5)	20 (26.7)	9 (30.0)	0.656
Poor differentiation	107 (33.1)	22 (29.3)	13 (43.3)	0.388
Satellite nodule	35 (10.8)	8 (10.7)	8 (26.7)	0.035
PVTT	5 (1.5)	2 (2.7)	1 (3.3)	0.673

Results presented as number (percentage) or median (interquartile range) values.

† Graft weight was directly measured during operation.

Abbreviations: ABOc, ABO compatible; ABOi, ABO incompatible; AFP, alpha-feto protein; BMI, body mass index; GRWR, graft recipient weight ratio; HCC, hepatocellular carcinoma; LRT, locoregional treatment; LT, liver transplantation; MELD, model for end-stage liver disease; PIVKA, protein induced by vitamin K antagonist-II; PVTT, portal vein tumor thrombosis; TPE, therapeutic plasma exchange.

3.2. Detailed information on recipient of ABOi LDLT

Almost all patients who underwent ABOi LDLT received rituximab and at least one cycle of TPE for desensitization. Table 2 presents details regarding ABO incompatibility and desensitization protocols for patients in the ABOi group, categorized by the number of pretransplant TPE sessions. A significantly higher proportion of A to O transplants was observed in the ABOi/TPE \geq 6 group (56.7%) than in the ABOi/TPE<6 group (17.3%, $P < 0.001$). The median IA titer was significantly higher in the ABOi/TPE \geq 6 group than in the ABOi/TPE<6 group at initial assessment (1:64 vs. 1:512, $P < 0.001$), at the time of LT (1:8 vs. 1:32, $P < 0.001$), and after LT (1:16 vs. 1:32, $P < 0.001$).

Additionally, a significantly higher proportion of patients in the ABOi/TPE \geq 6 group underwent splenectomy (5.3% vs. 23.3%, $P = 0.018$), pretransplant IVIG (10.7% vs. 53.3%, $P < 0.001$), posttransplant IVIG (8.0% vs. 30.0%, $P = 0.009$), and posttransplant TPE (18.7% vs. 36.7%, $P = 0.049$).

Table 2. Details for ABO incompatibility and desensitization of ABO incompatible group patients, according to therapeutic plasma exchange number

	ABOi/TPE<6 (n=75)	ABOi/TPE≥6 (n=30)	<i>P</i>
ABO type			<0.001
<i>A</i>	36 (48.0)	2 (6.7)	
<i>B</i>	16 (21.3)	1 (3.3)	
<i>O</i>	23 (30.7)	27 (90.0)	
Donor ABO type			0.008
<i>A</i>	23 (30.7)	17 (56.7)	
<i>AB</i>	25 (33.3)	2 (6.7)	
<i>B</i>	27 (36.0)	11 (36.7)	
A to O	13 (17.3)	17 (56.7)	<0.001
IA titer at initial ^a	1:64 (1:2-1:512)	1:512 (1:16-1:4096)	<0.001
IA titer at LT ^b	1:8 (1:1-1:256)	1:32 (1:1-1:128)	<0.001
Pretransplant TPE number	3 (2-4)	6.5 (6-7)	<0.001
Pretransplant IVIG	8 (10.7)	16 (53.3)	<0.001
Rituximab	73 (97.3)	30 (100.0)	0.910
Rituximab conventional dose ^c	54 (72.0)	26 (86.7)	0.078
Pretransplant duration of MMF	7 (4-8)	7 (0-8)	0.645
Pretransplant MMF total dose, mg	3500 (250-4000) ^d	3500 (750-7500) ^e	0.284
Splenectomy	4 (5.3)	7 (23.3)	0.018
Posttransplant IA titer rebound ^f	19 (25.3)	8 (26.7)	0.986
Posttransplant maximum IA titer ^g	1:16 (1:1-1:1024)	1:32 (1:2-1:2048)	0.001
Posttransplant TPE ^h	14 (18.7)	11 (36.7)	0.049
Posttransplant IVIG	6 (8.0)	9 (30.0)	0.009

Results presented as number (percentage) or median (range) values.

^a The number of min/max range patients was n=1 (Titer 1:2), n=2 (Titer 1:512), n=6 (Titer 1:16), and n=3 (Titer 1:4096)

^b The number of min/max range patients was n=7 (Titer 1:1), n=1 (Titer 1:256), n=6 (Titer 1:1), and n=2 (Titer 1:128)

^c 375±25 mg per body surface area(m²).

^d The number of min/max range patients was n=1 (250mg), and n=1 (5000mg). Interquartile range was 3500-4000mg

^e The number of min/max range patients was n=1 (750mg), and n=2 (7500mg). Interquartile range was 2000-4000mg

^f Defined as IA titer increased to more than 1:64 after transplantation.

^g The number of min/max range patients was n=4 (Titer 1:1), n=1 (Titer 1:1024), n=8 (Titer 1:2), and n=1 (Titer 1:2048)

^h Posttransplant TPE number ranges 0-10 in ABOi/TPE<6 group, and 0-15 in ABOi/TPE≥6 group.

Abbreviations: ABOi, ABO incompatible; IA, isoagglutinin; IVIG, intravenous immunoglobulin; LT, liver transplantation; MMF, mycophenolate mofetil; TPE, therapeutic plasma exchange;

3.3. HCC outcomes

As shown in the Kaplan-Meier curves in Figure 1, a significant difference was observed in RFS between the ABOc group and the ABOi/TPE \geq 6 group (5-year survival: 75.7% in the ABOc group vs. 50.0% in the ABOi/TPE \geq 6 group, $P=0.005$). Additionally, the HCC recurrence rates also differed significantly (5-year survival: 16.4% vs. 39.4%, $P = 0.014$).

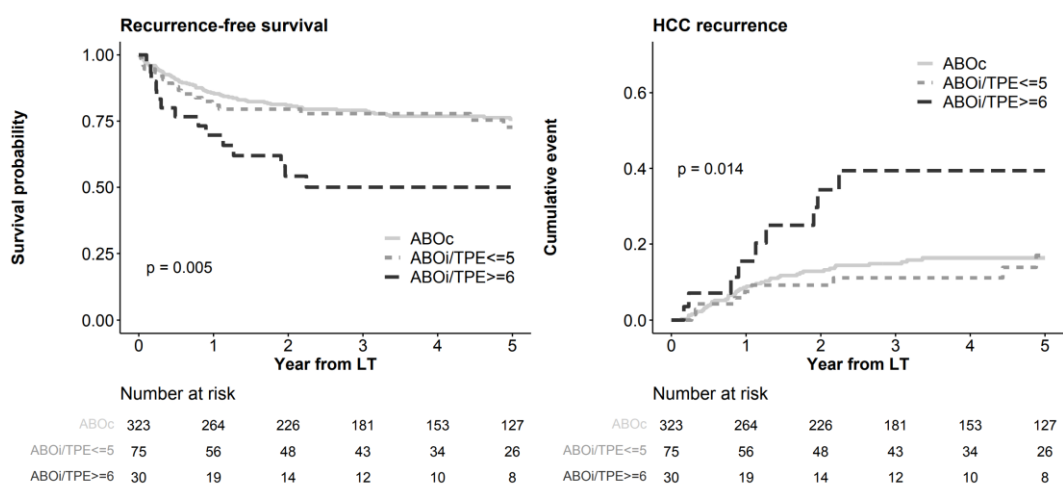


Figure 5. Kaplan-Meier curve of RFS and HCC recurrence according to ABO incompatibility and plasma exchange numbers.

RFS, recurrence free survival; HCC, hepatocellular carcinoma; ABOc, ABO compatible; ABOi, ABO incompatible; TPE, therapeutic plasma exchange; LT, liver transplantation.

To further evaluate the impact of TPE on oncologic outcomes, we categorized the ABO incompatibility group into subgroups based on the number of TPE sessions: ≤ 3 sessions (5-year RFS: 76.2%, 5-year HCC recurrence: 17.4%), 4-5 sessions (5-year RFS: 68.5%, 5-year HCC recurrence: 16.5%), and ≥ 6 sessions (5-year RFS: 50.0%, 5-year HCC recurrence: 39.4%).

Although these results were not statistically significant, a trend related to the number of TPE sessions was observed ($P = 0.056$ for RFS and $P = 0.051$ for HCC recurrence, Figure 6).

In the multivariable Cox analyses (Table 3), the ABOi/TPE ≥ 6 group was significantly associated with RFS (hazard ratio [HR] = 1.99, 95% confidence interval [CI]: 1.02-3.86, $P = 0.042$) and HCC recurrence (HR = 2.42, 95% CI: 1.05-5.57, $P = 0.037$).

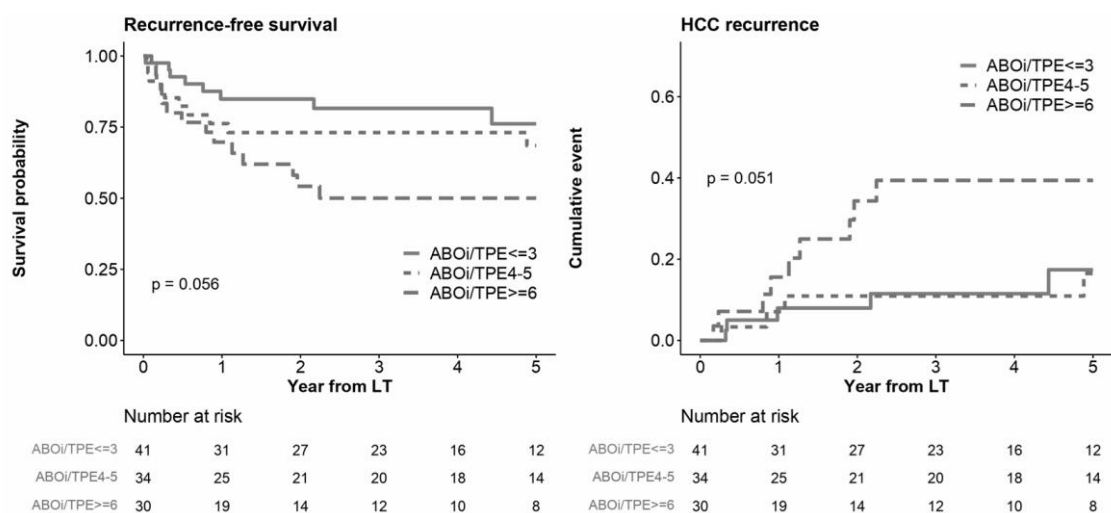


Figure 6. Kaplan-Meier curve of RFS and HCC recurrence in ABOi subgroup, according to plasma exchange numbers.

RFS, recurrence free survival; HCC, hepatocellular carcinoma; ABOi, ABO incompatible; TPE, therapeutic plasma exchange; LT, liver transplantation.

Table 3. Multivariable Cox analysis for recurrence free survival and hepatocellular carcinoma recurrence

Variables	Recurrence free survival HR (95% CI)	<i>P</i>	HCC recurrence [†] HR (95% CI)	<i>P</i>
ABOi group				
ABOc	Reference		Reference	
ABOi/TPE<6	1.08 (0.63-1.85)	0.777	0.97 (0.46-2.01)	0.928
ABOi/TPE≥6	1.99 (1.02-3.86)	0.042	2.42 (1.05-5.57)	0.037
Age, years	-	-	0.96 (0.92-1.00)	0.048
BMI	0.95 (0.89-1.01)	0.102	-	-
Pretransplant MELD	1.06 (1.03-1.09)	<0.001	-	-
Cold ischemic time, min	1.00 (1.00-1.01)	0.622	-	-
Transfusion RBC, pack	1.02 (1.01-1.04)	0.002	-	-
Log_AFP at LT	1.11 (0.98-1.25)	0.093	1.09 (0.94-1.25)	0.260
Log_PIVKA at LT	1.03 (0.90-1.18)	0.659	1.14 (0.98-1.34)	0.091
Pretransplant LRT, yes	2.91 (1.45-5.84)	0.003	7.00 (2.02-24.26)	0.002
Systemic treatment, yes	2.20 (1.37-3.53)	0.001	2.10 (1.16-3.82)	0.015
Viable tumor number	1.02 (1.01-1.04)	0.007	1.04 (1.01-1.07)	0.004
Maximum tumor size, cm	0.90 (0.82-0.98)	0.019	0.92 (0.82-1.03)	0.150
Microvascular invasion, yes	1.77 (0.97-3.22)	0.062	2.07 (1.01-4.24)	0.046
Poor differentiation, yes	1.30 (0.82-2.05)	0.268	1.69 (0.95-3.02)	0.076
Satellite nodule, yes	1.67 (0.90-3.10)	0.101	1.83 (0.92-3.63)	0.085
PVTT, yes	2.83 (0.98-8.16)	0.054	2.49 (0.57-10.86)	0.226

Variables which result $p < 0.1$ in univariable Cox analysis were included and represented at multivariable Cox analysis. Full univariate and multivariate results are represented at Appendix Table 1 and Appendix Table 2.

[†] Multivariable analysis for HCC recurrence was performed treating non-HCC death as competing risk.

Abbreviations: ABOc, ABO compatible; ABOi, ABO incompatible; AFP, alpha-feto protein; BMI, body mass index; CI, confidence interval; HCC, hepatocellular carcinoma; HR, hazard ratio; LRT, locoregional treatment; MELD, model for end-stage liver disease; PIVKA, protein induced by vitamin K antagonist-II; PVTT, portal vein tumor thrombosis; TPE, therapeutic plasma exchange.

3.4. Subgroup analysis for HCC recurrence

In the subgroup analysis (Table 4), the 5-year HCC recurrence rates were higher across all subgroups in the ABOi/TPE \geq 6 group. Although this trend is numerically apparent, the small sample size limits the ability to confirm statistical significance. Interestingly, among patients with a tumor marker-based MoRAL score \geq 100, the recurrence rate was significantly higher in the ABOi/TPE \geq 6 group (56.7%) than in the ABOi/TPE $<$ 6 group with a MoRAL score \geq 100 (16.1%, $P = 0.017$). However, patients with a MoRAL score $<$ 100 exhibited similar 5-year HCC recurrence rates between the two groups (17.6% vs. 20.5%, $P = 0.75$). Figure 7 illustrates HCC recurrence based on the number of TPE sessions and the MoRAL score. As observed, a marked difference was evident between the ABOi/TPE \geq 6 group with a high MoRAL score and the other groups ($P = 0.0042$).

Regarding the tumor burden criteria, the ABOi/TPE $<$ 6 group of patients within the Milan criteria exhibited a significantly lower 5-year HCC recurrence rate (9.9%) than the ABOi/TPE \geq 6 group (35.2%, $P = 0.025$). Additionally, the 5-year HCC recurrence rate was significantly lower in the ABOi/TPE $<$ 6 group of patients within the Up-to-7 criteria (16.1%) than in the ABOi/TPE \geq 6 group (35.5%, $P = 0.033$).

In the subgroup analysis based on immunological classification, patients with an initial IA titer \leq 1:128 demonstrated a significantly higher recurrence rate in the ABOi/TPE \geq 6 group (50.0%) than in the ABOi/TPE $<$ 6 group (17.9%, $P = 0.035$). However, patients with an IA titer \geq 1:32 at LT had a significantly higher recurrence rate in the ABOi/TPE \geq 6 group (43.5%) than in the ABOi/TPE $<$ 6 group (0.0%, $P = 0.026$).

Table 4. Subgroup analysis of 5-year hepatocellular carcinoma recurrence according to therapeutic plasma exchange numbers in ABO incompatible group

Subgroups	Patient number		5 year HCC recurrence		<i>P</i>
	ABOi/TPE<6	ABOi/TPE≥6	ABOi/TPE<6	ABOi/TPE≥6	
	(n=75)	(n=30)	(n=75)	(n=30)	
Milan criteria					
Within	44	15	9.9%	35.2%	0.025
Above	31	15	27.2%	43.8%	0.267
Up-to-7					
Within	65	22	16.1%	35.5%	0.033
Above	10	8	27.1%	47.5%	0.569
French risk score					
≤2	56	20	14.8%	31.8%	0.082
>2	19	10	21.3%	55.0%	0.117
MoRAL score					
<100	49	16	17.6%	20.5%	0.750
≥100	26	14	16.1%	56.7%	0.017
IA titer at initial					
≤1:128	65	6	17.9%	50.0%	0.035
≥1:256	10	24	0.0%	64.1%	0.177
IA titer at LT					
≤1:16	61	11	19.4%	31.8%	0.287
≥1:32	14	19	0.0%	43.5%	0.026
IA titer rebound					
No	56	22	12.9%	28.6%	0.073
Yes	19	8	28.9%	66.7%	0.103
Post LT TPE					
No	61	19	12.1%	24.0%	0.203
Yes	14	11	37.3%	59.1%	0.246
Splenectomy					
No	71	23	17.9%	36.9%	0.075
Yes	4	7	0.0%	46.4%	0.137

Abbreviations: ABOi, ABO incompatible; HCC, hepatocellular carcinoma; IA, isoagglutinin; LT, liver transplantation; MoRAL, Model of Recurrence After Liver Transplant; TPE, therapeutic plasma exchange.

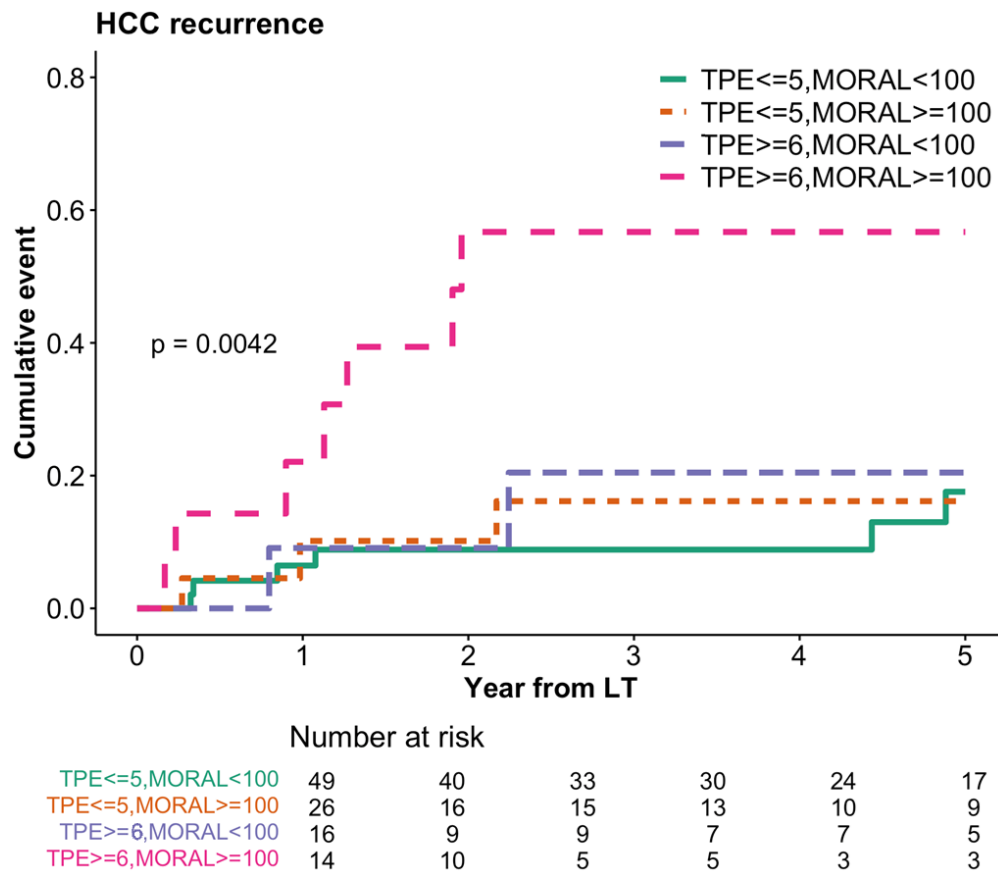


Figure 7 - HCC recurrence in ABOi subgroup, according to plasma exchange numbers and MORAL score.

HCC, hepatocellular carcinoma; TPE, therapeutic plasma exchange; MORAL, Model Of Recurrence After Liver transplant; LT, liver transplantation.

IV. DISCUSSION

The study evaluated the impact of pretransplant TPE sessions on HCC recurrence in patients undergoing ABOi LDLT and to determine if limiting TPE sessions to fewer than six can enhance oncologic outcomes. We found that in the ABOi LDLT group, patients who underwent more than six pretransplant TPE sessions exhibited significantly worse HCC RFS and recurrence outcomes, with a similar trend observed in the subgroup analysis. Interestingly, the MoRAL score, which includes biomarkers, revealed that poorer oncologic outcomes were particularly pronounced in the high MoRAL score group. This suggests that in patients requiring a greater number of TPE sessions, biomarkers, in addition to tumor size, may play a crucial role in influencing HCC outcomes. The strengths of our study include a well-organized dataset and a standardized desensitization protocol within the context of ABOi LDLT.

Globally, there has been a growing demand for LT as a definitive treatment for HCC, particularly for LDLT and ABOi LDLT due to organ shortages. (Matsuno et al., 2008; Rela & Rammohan, 2021) In many countries outside East Asia, there is greater availability of deceased donors (DD), resulting in a predominant reliance on DDLT. (Hibi et al., 2020; Lee et al., 2012; Rela & Rammohan, 2021) Consequently, these regions have limited cases and data regarding ABOi LDLT and the frequent use of TPE. In contrast, due to extreme shortages of deceased donors in Korea, LDLT is commonly performed for HCC. (Choi, 2022; Hibi et al., 2020) Paradoxically, this societal impact of deceased donor shortages has contributed to the accumulation of extensive data on ABOi LDLT, particularly in cases with high ABO antibody titers and a greater number of TPE sessions.

TPE is an intervention that involves the extracorporeal removal, return, or exchange of blood plasma or its components. (Kaplan, 2008; Schwartz et al., 2016) The fundamental mechanism of this procedure is accomplished through centrifugation or filtration using semipermeable membranes. (Gerhardt et al., 1992; Siami & Siami, 2001) In ABOi LDLT, the primary purpose of TPE is to remove IA. However, because this procedure is not selective, other immune-related factors in the blood are also removed, which presents a theoretical concern. Consequently, TPE is typically used as a primary or adjunctive treatment for conditions such as neurological diseases—including multiple sclerosis, amyotrophic lateral sclerosis, and myasthenia gravis—as well as autoimmune diseases like systemic lupus erythematosus and Kawasaki disease. Recent studies in this field have indicated that TPE promotes the differentiation and function of regulatory T cells.

(Barath et al., 2007; Fiorini et al., 1982; Jamshidian & Gharagozloo, 2012; Koizumi et al., 2019; Mehdi pour et al., 2021; Thonhoff et al., 2016; Zhang et al., 2014)

Upon reviewing prior studies, it was noted that desensitization through pretransplant TPE or induction therapy in immunologically high-risk groups is associated with an increased cancer risk in certain malignancies (Table 5). (S. D. Lee et al., 2015; Miyagi et al., 2012; Motter et al., 2023; Yang et al., 2016) Although specific studies on ABOi LDLT are lacking, and the existing literature did not establish consistent protocols for TPE in kidney transplantation, direct comparisons with our study are challenging. Nevertheless, these findings underscore the relevance of desensitization and induction therapy concerning cancer risk, which was considered in our research.

Table 5. Previous studies regarding pretransplant desensitization and cancer risk.

Study	Yang, C.Y., et al. †	Motter, J.D., et al. ‡
Country	Taiwan	USA
Study period	2007-2013	1997-2016
Transplantation	Kidney	Kidney
Compared groups	DSA+ (n=22) vs. DSA- (n=152)	ABOi LDKT (n=858) vs. ABOc LDKT (n=12239)
TPE number	At least 4 cycles in DSA+ group	Not provided
Cancer type	Urothelial, endometrial, colon, and thyroid cancer	Colorectal cancer
Cancer incidence	DSA+ 19.6% vs. DSA- 8.5% for 5 year (HR =7.81, p=0.028)	ABOi 0.6% vs. ABOc 0.3% (HR=3.27, p=0.002)
Hypothesis for higher cancer incidence	Desensitization therapy for DSA+ including TPE might increase cancer	Desensitization therapy might increase cancer

† Yang, C.Y., et al., *Renal transplantation across the donor-specific antibody barrier: Graft outcome and cancer risk after desensitization therapy*. J Formos Med Assoc, 2016. **115**(6): p. 426-33.

‡ Motter, J.D., et al., *Cancer Risk Following HLA-Incompatible Living Donor Kidney Transplantation*. Transplant Direct, 2023. **9**(8): p. e1505.

Abbreviations: ABOc, ABO compatible; ABOi, ABO incompatible; DSA, donor specific antibody; TPE, therapeutic plasma exchange; HR, hazard ratio; LDKT, living donor kidney transplantation; TPE, therapeutic plasma exchange.

Recent trends suggest that ABOi LDLT outcomes, including HCC cancer outcome and oncologic survival benefits, are comparable to those of ABO compatible (ABOc) LT. (Kim et al., 2018; Kim et al., 2019; Yoon et al., 2018) However, these studies did not account for the cumulative and long-term effects of TPE, which prompted the initiation of our research.

In our study, the data indicated that ABOi patients requiring six or more pretransplant TPE sessions exhibited significantly poorer RFS and higher rates of HCC recurrence than ABOc patients. Additionally, our subgroup analysis shows that a higher number of pretransplant TPE sessions (≥ 6) was associated with a statistically significant increase in the 5-year HCC recurrence rate across several subgroups, including those within the Milan and Up-to-7 criteria, those with a high MoRAL score, and those with lower initial IA titers and higher IA titers at the time of LT.

Notably, within the size-based criteria, the ABOi/TPE ≥ 6 group exhibited a significantly higher recurrence rate. In contrast, regarding the tumor marker-based MoRAL score, a higher recurrence rate was observed in the ABOi/TPE ≥ 6 group only among patients with a score above 100. This suggests that among patients with a lower size-based tumor burden and a higher biologic-based tumor burden, those requiring more TPE sessions tended to experience poorer oncologic outcomes. Furthermore, this implies that the number of TPE sessions may be a more critical factor than the IA titer in influencing these outcomes.

Unlike previous studies, we focused on the immunomodulatory effects of T-regulatory (T-reg) cells induced by TPE and their association with HCC recurrence. As discussed earlier, while it is well established that T-reg cells are effective in treating autoimmune and neurological disorders, there are theoretical concerns that this process may reduce patient resistance to cancer. (Miyara & Sakaguchi, 2007) The literature indicates that the activation of T-reg cells can increase the risk of cancers such as HCC, with CD4+CD25+FoxP3+ T cells playing a significant role in this risk. (Kalathil et al., 2013; Sakaguchi et al., 2008; Sun et al., 2014) Although the exact cytokines and mechanisms through which these cells interact with others remain unclear, their differentiation in the tumor microenvironment (TME) has been observed, (Adeegbe & Nishikawa, 2013; Fridman et al., 2012; Ohue & Nishikawa, 2019) suggesting a potential increase in poor long-term cancer outcomes in various malignancies, including HCC. This information is illustrated in Figure 8.

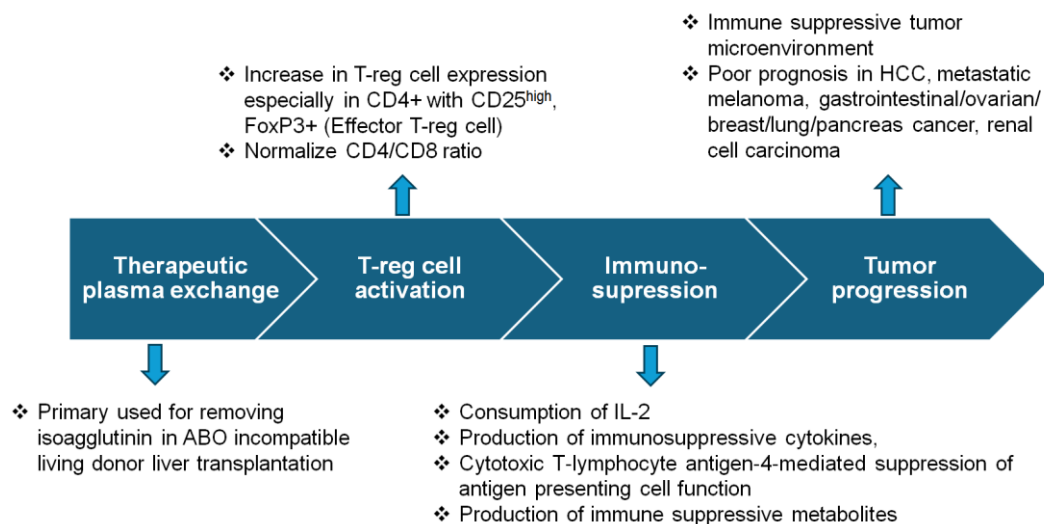


Figure 8. Therapeutic plasma exchange and tumor progression flowchart

This diagram outlines the impact of TPE on T-reg cell activation and its subsequent effects on immunosuppression and tumor progression. TPE is primarily employed to remove isoagglutinin in ABO incompatible living donor liver transplantation, which increases T-reg cell expression, particularly effector T-reg cell. This immune modulation contributes to IL-2 consumption and enhances the production of immunosuppressive cytokines, resulting in a more immunosuppressive tumor microenvironment and poorer prognostic outcomes in HCC.

T-reg, T-regulatory; FoxP3, forkhead box P3; IL, interleukin; HCC, hepatocellular carcinoma.

In summary, our hypothesis suggests that plasmapheresis induces the activation of T-reg cells, particularly CD4⁺ with CD25^{high}, FoxP3⁺ effector T-reg cells, leading to an immunosuppressive effect within the tumor microenvironment that may facilitate tumor progression in various malignancies, including HCC. While some aspects of this pathway remain unexplained in the current foundational research, further studies are warranted to elucidate these mechanisms. Notably, the cumulative effect of TPE in the context of ABOi LT has not been extensively studied, underscoring the significance of our research.

This study has some limitations, including its retrospective and non-randomized design, the low number of ABOi/TPE ≥ 6 patients from a single center (n=30), and the lack of fully established theoretical hypotheses or evidence to support our claims. However, despite these limitations, our study is significant, as it is the first to investigate the relationship between HCC outcomes and the number of preoperative TPE sessions. In the future, we aim to address these limitations by increasing the sample size and establishing a more robust theoretical framework.

V. CONCLUSIONS

This study demonstrated that the administration of more than six pretransplant TPE sessions in patients with HCC undergoing ABOi LDLT was associated with poorer oncologic outcomes. Based on our clinical findings and the theoretical association between TPE and HCC oncologic outcomes, we propose that limiting the number of TPE sessions to fewer than six may improve cancer outcomes in patients with HCC receiving ABOi LDLT. A strategy to reduce the number of TPE sessions to fewer than five should be implemented when planning ABOi LDLT for HCC patients, while ensuring similar immunologic risk.

References

1. Adeegbe, D. O., & Nishikawa, H. (2013). Natural and Induced T Regulatory Cells in Cancer [Review]. *Frontiers in Immunology*, 4. <https://doi.org/10.3389/fimmu.2013.00190>
2. Akamatsu, N., Sugawara, Y., & Kokudo, N. (2014). Living-donor vs deceased-donor liver transplantation for patients with hepatocellular carcinoma. *World J Hepatol*, 6(9), 626-631. <https://doi.org/10.4254/wjh.v6.i9.626>
3. Azoulay, D., Audureau, E., Bhangui, P., Belghiti, J., Boillot, O., Andreani, P., Castaing, D., Cherqui, D., Irtan, S., Calmus, Y., Chazouillères, O., Soubrane, O., Luciani, A., & Feray, C. (2017). Living or Brain-dead Donor Liver Transplantation for Hepatocellular Carcinoma: A Multicenter, Western, Intent-to-treat Cohort Study. *Annals of Surgery*, 266(6). https://journals.lww.com/annalsofsurgery/fulltext/2017/12000/living_or_brain_dead_donor_liver_transplantation.23.aspx
4. Barath, S., Soltész, P., Kiss, E., Aleksza, M., Zeher, M., Szegedi, G., & Sipka, S. (2007). The severity of systemic lupus erythematosus negatively correlates with the increasing number of CD4+CD25(high)FoxP3+ regulatory T cells during repeated plasmapheresis treatments of patients. *Autoimmunity*, 40(7), 521-528. <https://doi.org/10.1080/08916930701610028>
5. Choi, H. J. (2022). Current status and outcome of liver transplantation in South Korea. *Clin Mol Hepatol*, 28(1), 117-119. <https://doi.org/10.3350/cmh.2021.0381>
6. Choi, M. C., Min, E.-K., Yim, S. H., Kim, D.-G., Lee, J. G., Joo, D. J., & Kim, M. S. (2024). High Number of Plasma Exchanges Increases the Risk of Bacterial Infection in ABO-incompatible Living Donor Liver Transplantation. *Transplantation*, 108(8). https://journals.lww.com/transplantjournal/fulltext/2024/08000/high_number_of_plasma_exchanges_increases_the_risk.20.aspx
7. Duvoux, C., Roudot-Thoraval, F., Decaens, T., Pessione, F., Badran, H., Piardi, T., Francoz, C., Compagnon, P., Vanlemmens, C., Dumortier, J., Dharancy, S., Gugenheim, J., Bernard, P. H., Adam, R., Radenne, S., Muscari, F., Conti, F., Hardwigsen, J., Pageaux, G. P., . . . Cherqui, D. (2012). Liver Transplantation for Hepatocellular Carcinoma: A Model Including α -Fetoprotein Improves the Performance of Milan Criteria. *Gastroenterology*, 143(4), 986-994.e983. <https://doi.org/https://doi.org/10.1053/j.gastro.2012.05.052>
8. Egawa, H., Teramukai, S., Haga, H., Tanabe, M., Mori, A., Ikegami, T., Kawagishi, N., Ohdan, H., Kasahara, M., & Umeshita, K. (2014). Impact of Rituximab Desensitization on Blood-Type-Incompatible Adult Living Donor Liver Transplantation: A Japanese Multicenter Study. *American Journal of Transplantation*, 14(1), 102-114. <https://doi.org/https://doi.org/10.1111/ajt.12520>
9. Fine, J. P., & Gray, R. J. (1999). A Proportional Hazards Model for the Subdistribution of a Competing Risk. *Journal of the American Statistical Association*, 94(446), 496-509. <https://doi.org/10.1080/01621459.1999.10474144>
10. Fiorini, G., Paracchini, M. L., & Fornasieri, A. (1982). Modifications in peripheral blood lymphocyte subpopulations induced by plasmapheresis and immunosuppressive drugs [Article]. *Plasma Therapy and Transfusion Technology*, 3(4), 389-393. <https://www.scopus.com/inward/record.uri?eid=2-s2.0-0020365906&partnerID=40&md5=ec975b947b70b614de3c7d0582460c14>

11. Fridman, W. H., Pages, F., Sautes-Fridman, C., & Galon, J. (2012). The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer*, 12(4), 298-306. <https://doi.org/10.1038/nrc3245>
12. Gerhardt, R. E., Ntoso, K. A., Koethe, J. D., Lodge, S., & Wolf, C. J. (1992). Acute plasma separation with hemodialysis equipment. *J Am Soc Nephrol*, 2(9), 1455-1458. <https://doi.org/10.1681/asn.V291455>
13. Goldaracena, N., & Barbas, A. S. (2019). Living donor liver transplantation. *Current Opinion in Organ Transplantation*, 24(2). https://journals.lww.com/co-transplantation/fulltext/2019/04000/living_donor_liver_transplantation.6.aspx
14. Gordon, R. D., Iwatsuki, S., Esquivel, C. O., Tzakis, A., Todo, S., & Starzl, T. E. (1986). Liver transplantation across ABO blood groups. *Surgery*, 100(2), 342-348. <https://www.ncbi.nlm.nih.gov/pubmed/3526607>
15. Gugenheim, J., Samuel, D., Bismuth, H., & Reynes, M. (1990). Liver transplantation across ABO blood group barriers. *The Lancet*, 336(8714), 519-523. [https://doi.org/https://doi.org/10.1016/0140-6736\(90\)92082-S](https://doi.org/https://doi.org/10.1016/0140-6736(90)92082-S)
16. Hibi, T., Chieh, A. K. W., Chan, A. C.-Y., & Bhangu, P. (2020). Current status of liver transplantation in Asia. *International Journal of Surgery*, 82. https://journals.lww.com/international-journal-of-surgery/fulltext/2020/10001/current_status_of_liver_transplantation_in_asia.2.aspx
17. Jamshidian, A., & Gharagozloo, M. (2012). Can plasma exchange therapy induce regulatory T lymphocytes in multiple sclerosis patients? *Clin Exp Immunol*, 168(1), 75-77. <https://doi.org/10.1111/j.1365-2249.2011.04547.x>
18. Kalathil, S., Lugade, A. A., Miller, A., Iyer, R., & Thanavala, Y. (2013). Higher Frequencies of GARP+CTLA-4+Foxp3+ T Regulatory Cells and Myeloid-Derived Suppressor Cells in Hepatocellular Carcinoma Patients Are Associated with Impaired T-Cell Functionality. *Cancer Research*, 73(8), 2435-2444. <https://doi.org/10.1158/0008-5472.Can-12-3381>
19. Kang, S. H., Song, G.-W., & Yoon, Y.-I. (2019). Outcome of ABO-incompatible adult living-donor liver transplantation for patients with hepatocellular carcinoma. *HPB*, 21, S344-S345.
20. Kaplan, A. A. (2008). Therapeutic Plasma Exchange: Core Curriculum 2008. *American Journal of Kidney Diseases*, 52(6), 1180-1196. <https://doi.org/https://doi.org/10.1053/j.ajkd.2008.02.360>
21. Kim, J. M., Kwon, C. H. D., Joh, J.-W., Han, S., Yoo, J., Kim, K., Sinn, D. H., Choi, G.-S., Gerber, D. A., Egawa, H., & Lee, S.-K. (2018). ABO-incompatible Living Donor Liver Transplantation With Rituximab and Total Plasma Exchange Does Not Increase Hepatocellular Carcinoma Recurrence. *Transplantation*, 102(10). https://journals.lww.com/transplantjournal/fulltext/2018/10000/abo_incompatible_livin_g_donor_liver.24.aspx
22. Kim, J. M., Kwon, C. H. D., Joh, J.-W., Kang, E.-S., Park, J. B., Lee, J. H., Kim, S. J., Paik, S. W., Lee, S.-K., & Kim, D. W. (2013). ABO-incompatible living donor liver transplantation is suitable in patients without ABO-matched donor. *Journal of Hepatology*, 59(6), 1215-1222. <https://doi.org/https://doi.org/10.1016/j.jhep.2013.07.035>
23. Kim, S. H., Lee, E. C., Na, B. G., & Park, S. J. (2019). Impact of ABO-incompatibility on hepatocellular carcinoma recurrence after living donor liver transplantation. *European Journal of Surgical Oncology*, 45(2), 180-186.

- <https://doi.org/https://doi.org/10.1016/j.ejso.2018.07.066>
24. Koizumi, K., Hoshiai, M., Moriguchi, T., Katsumata, N., Toda, T., Kise, H., Hasebe, Y., Kono, Y., Sunaga, Y., Yoshizawa, M., Watanabe, A., Harii, N., Goto, J., Kagami, K., Abe, M., Matsuda, K., & Sugita, K. (2019). Plasma Exchange Downregulates Activated Monocytes and Restores Regulatory T Cells in Kawasaki Disease. *Ther Apher Dial*, 23(1), 92-98. <https://doi.org/10.1111/1744-9987.12754>
 25. Lai, Q., Sapisochin, G., Gorgen, A., Vitale, A., Halazun, K. J., Iesari, S., Schaefer, B., Bhangu, P., Mennini, G., Wong, T. C. L., Uemoto, S., Lin, C.-C., Mittler, J., Ikegami, T., Yang, Z., Frigo, A. C., Zheng, S.-S., Soejima, Y., Hoppe-Lotichius, M., . . . Lerut, J. P. (2021). Evaluation of the Intention-to-Treat Benefit of Living Donation in Patients With Hepatocellular Carcinoma Awaiting a Liver Transplant. *JAMA Surgery*, 156(9), e213112-e213112. <https://doi.org/10.1001/jamasurg.2021.3112>
 26. Lee, C.-F., Cheng, C.-H., Wang, Y.-C., Soong, R.-S., Wu, T.-H., Chou, H.-S., Wu, T.-J., Chan, K.-M., Lee, C.-S., & Lee, W.-C. (2015). Adult Living Donor Liver Transplantation Across ABO-Incompatibility. *Medicine*, 94(42). https://journals.lww.com/md-journal/fulltext/2015/10030/adult_living_donor_liver_transplantation_across.62.aspx
 27. Lee, J.-H., Cho, Y., Kim, H. Y., Cho, E. J., Lee, D. H., Yu, S. J., Lee, J. W., Yi, N.-J., Lee, K.-W., Kim, S. H., Kim, J. M., Joh, J.-W., Teperman, L. W., Park, J. S., Kim, Y. J., Suh, K.-S., & Yoon, J.-H. (2016). Serum Tumor Markers Provide Refined Prognostication in Selecting Liver Transplantation Candidate for Hepatocellular Carcinoma Patients Beyond the Milan Criteria. *Annals of Surgery*, 263(5). https://journals.lww.com/annalsofsurgery/fulltext/2016/05000/serum_tumor_markers_provide_refined.4.aspx
 28. Lee, J., Lee, J. G., Lee, J. J., Kim, M. S., Ju, M. K., Choi, G. H., Choi, J. S., Kim, S. I., & Joo, D. J. (2015). Results of ABO-Incompatible Liver Transplantation Using a Simplified Protocol at a Single Institution. *Transplantation Proceedings*, 47(3), 723-726. <https://doi.org/https://doi.org/10.1016/j.transproceed.2015.02.004>
 29. Lee, S. D., Kim, S. H., Kong, S.-Y., Kim, Y.-K., Lee, S.-A., & Park, S.-J. (2014). ABO-incompatible living donor liver transplantation without graft local infusion and splenectomy. *HPB*, 16(9), 807-813. <https://doi.org/https://doi.org/10.1111/hpb.12215>
 30. Lee, S. D., Kim, S. H., Kong, S.-Y., Kim, Y.-K., & Park, S.-J. (2015). Kinetics of B, T, NK lymphocytes and isoagglutinin titers in ABO incompatible living donor liver transplantation using rituximab and basiliximab. *Transplant Immunology*, 32(1), 29-34. <https://doi.org/https://doi.org/10.1016/j.trim.2014.11.216>
 31. Lee, S. G. (2015). A Complete Treatment of Adult Living Donor Liver Transplantation: A Review of Surgical Technique and Current Challenges to Expand Indication of Patients. *American Journal of Transplantation*, 15(1), 17-38. <https://doi.org/https://doi.org/10.1111/ajt.12907>
 32. Lee, S. G., Moon, D. B., Shin, H., Kim, K. H., Ahn, C. S., Ha, T. Y., Song, G. W., Jung, D. H., & Park, G. C. (2012). Living Donor Liver Transplantation for Hepatocellular Carcinoma: Current Status in Korea. *Transplantation Proceedings*, 44(2), 520-522. <https://doi.org/https://doi.org/10.1016/j.transproceed.2012.02.003>
 33. Lee, W.-C., Cheng, C.-H., Lee, C.-F., Hung, H.-C., Lee, J.-C., Wu, T.-H., Wang, Y.-C., Wu, T.-J., Chou, H.-S., & Chan, K.-M. (2022). Quick preparation of ABO-incompatible living donor liver transplantation for acute liver failure. *Clinical Transplantation*, 36(3), e14555. <https://doi.org/https://doi.org/10.1111/ctr.14555>
 34. Matsuno, N., Iwamoto, H., Nakamura, Y., Hama, K., Kihara, Y., Konno, O., Jojima, Y.,

- Akashi, I., Mijiti, A., Ashizawa, T., & Nagao, T. (2008). ABO-Incompatible Adult Living Donor Liver Transplantation for Hepatocellular Carcinoma. *Transplantation Proceedings*, 40(8), 2497-2500. <https://doi.org/https://doi.org/10.1016/j.transproceed.2008.07.054>
35. Mazzaferro, V., Llovet, J. M., Miceli, R., Bhoori, S., Schiavo, M., Mariani, L., Camerini, T., Roayaie, S., Schwartz, M. E., Grazi, G. L., Adam, R., Neuhaus, P., Salizzoni, M., Bruix, J., Forner, A., De Carlis, L., Cillo, U., Burroughs, A. K., Troisi, R., . . . Majno, P. (2009). Predicting survival after liver transplantation in patients with hepatocellular carcinoma beyond the Milan criteria: a retrospective, exploratory analysis. *The Lancet Oncology*, 10(1), 35-43. [https://doi.org/10.1016/S1470-2045\(08\)70284-5](https://doi.org/10.1016/S1470-2045(08)70284-5)
36. Mazzaferro, V., Regalia, E., Doci, R., Andreola, S., Pulvirenti, A., Bozzetti, F., Montalto, F., Ammatuna, M., Morabito, A., & Gennari, L. Liver Transplantation for the Treatment of Small Hepatocellular Carcinomas in Patients with Cirrhosis. *New England Journal of Medicine*, 334(11), 693-700. <https://doi.org/10.1056/NEJM199603143341104>
37. Mehdipour, M., Etienne, J., Liu, C., Mehdipour, T., Kato, C., Conboy, M., Conboy, I., & Kiproff, D. D. (2021). Attenuation of age-elevated blood factors by repositioning plasmapheresis: A novel perspective and approach. *Transfus Apher Sci*, 60(3), 103162. <https://doi.org/10.1016/j.transci.2021.103162>
38. Miyagi, S., Kawagishi, N., Sekiguchi, S., Akamatsu, Y., Sato, K., Takeda, I., Kobayashi, Y., Tokodai, K., Fujimori, K., & Satomi, S. (2012). The Relationship Between Recurrences and Immunosuppression on Living Donor Liver Transplantation for Hepatocellular Carcinoma. *Transplantation Proceedings*, 44(3), 797-801. <https://doi.org/https://doi.org/10.1016/j.transproceed.2012.01.012>
39. Miyara, M., & Sakaguchi, S. (2007). Natural regulatory T cells: mechanisms of suppression. *Trends in Molecular Medicine*, 13(3), 108-116. <https://doi.org/https://doi.org/10.1016/j.molmed.2007.01.003>
40. Motter, J. D., Massie, A. B., Garonzik-Wang, J. M., Pfeiffer, R. M., Yu, K. J., Segev, D. L., & Engels, E. A. (2023). Cancer Risk Following HLA-Incompatible Living Donor Kidney Transplantation. *Transplant Direct*, 9(8), e1505. <https://doi.org/10.1097/TXD.0000000000001505>
41. Ninomiya, M., Shirabe, K., Facciuto, M. E., Schwartz, M. E., Florman, S. S., Yoshizumi, T., Harimoto, N., Ikegami, T., Uchiyama, H., & Maehara, Y. (2015). Comparative Study of Living and Deceased Donor Liver Transplantation as a Treatment for Hepatocellular Carcinoma. *Journal of the American College of Surgeons*, 220(3), 297-304.e293. <https://doi.org/https://doi.org/10.1016/j.jamcollsurg.2014.12.009>
42. Ogawa, K., & Takada, Y. (2016). Living vs. deceased-donor liver transplantation for patients with hepatocellular carcinoma. *Transl Gastroenterol Hepatol*, 1, 35. <https://doi.org/10.21037/tgh.2016.04.03>
43. Ohue, Y., & Nishikawa, H. (2019). Regulatory T (Treg) cells in cancer: Can Treg cells be a new therapeutic target? *Cancer Sci*, 110(7), 2080-2089. <https://doi.org/10.1111/cas.14069>
44. Rela, M., & Rammohan, A. (2021). Why are there so many liver transplants from living donors in Asia and so few in Europe and the US? *Journal of Hepatology*, 75(4), 975-980. <https://doi.org/https://doi.org/10.1016/j.jhep.2021.05.036>
45. Sakaguchi, S., Yamaguchi, T., Nomura, T., & Ono, M. (2008). Regulatory T Cells and Immune Tolerance. *Cell*, 133(5), 775-787. <https://doi.org/https://doi.org/10.1016/j.cell.2008.05.009>

46. Schwartz, J., Padmanabhan, A., Aqui, N., Balogun, R. A., Connelly-Smith, L., Delaney, M., Dunbar, N. M., Witt, V., Wu, Y., & Shaz, B. H. (2016). Guidelines on the Use of Therapeutic Apheresis in Clinical Practice-Evidence-Based Approach from the Writing Committee of the American Society for Apheresis: The Seventh Special Issue. *J Clin Apher*, 31(3), 149-162. <https://doi.org/10.1002/jca.21470>
47. Siami, G. A., & Siami, F. S. (2001). Membrane plasmapheresis in the United States: a review over the last 20 years. *Ther Apher*, 5(4), 315-320. <https://doi.org/10.1046/j.1526-0968.2001.00316.x>
48. Sun, W., Li, W.-J., Wu, C.-Y., Zhong, H., & Wen, W.-P. (2014). CD45RA-Foxp3^{high} but not CD45RA-Foxp3^{low} suppressive T regulatory cells increased in the peripheral circulation of patients with head and neck squamous cell carcinoma and correlated with tumor progression. *Journal of Experimental & Clinical Cancer Research*, 33(1), 35. <https://doi.org/10.1186/1756-9966-33-35>
49. Tanabe, M., Shimazu, M., Wakabayashi, G., Hoshino, K., Kawachi, S., Kadomura, T., Seki, H., Morikawa, Y., & Kitajima, M. (2002). Intraportal infusion therapy as a novel approach to adult ABO-incompatible liver transplantation1. *Transplantation*, 73(12). https://journals.lww.com/transplantjournal/fulltext/2002/06270/intraportal_infusion_the_rapy_as_a_novel_approach.21.aspx
50. Thonhoff, J., Beers, D., Zhao, W., Wen, S., Wang, J., Lay, L., Thompson, L., Leveque, C., & Appel, S. (2016). Plasmapheresis Improves the Suppressive Function of Regulatory T Cells in Patients with Fast-Progressing Amyotrophic Lateral Sclerosis (P3.183). *Neurology*, 86(16 supplement), P3.183. https://doi.org/doi:10.1212/WNL.86.16_supplement.P3.183
51. Yang, C. Y., Lee, C. Y., Yeh, C. C., & Tsai, M. K. (2016). Renal transplantation across the donor-specific antibody barrier: Graft outcome and cancer risk after desensitization therapy. *J Formos Med Assoc*, 115(6), 426-433. <https://doi.org/10.1016/j.jfma.2015.11.006>
52. Yim, S. H., Kim, D.-G., Kang, M., Koh, H.-h., Choi, M. C., Min, E.-K., Lee, J. G., Kim, M. S., & Joo, D. J. (2023). Intention-to-treat analysis for survival benefit of ABO-incompatible living-donor liver transplantation in patients with a high Model for End-stage Liver Disease score. *Korean Journal of Transplantation*, 37(1), 77-77.
53. Yoon, Y.-I., Song, G.-W., Lee, S.-G., Hwang, S., Kim, K.-H., Kim, S.-H., Kang, W.-H., Cho, H.-D., Jwa, E.-K., Kwon, J.-H., Tak, E.-Y., & Kirchner, V. A. (2018). Outcome of ABO-incompatible adult living-donor liver transplantation for patients with hepatocellular carcinoma. *Journal of Hepatology*, 68(6), 1153-1162. <https://doi.org/https://doi.org/10.1016/j.jhep.2018.02.002>
54. Zhang, L., Liu, J., Wang, H., Zhao, C., Lu, J., Xue, J., Gu, Y., Hao, C., Lin, S., & Lv, C. (2014). Double filtration plasmapheresis benefits myasthenia gravis patients through an immunomodulatory action. *Journal of Clinical Neuroscience*, 21(9), 1570-1574. <https://doi.org/https://doi.org/10.1016/j.jocn.2013.11.046>

APPENDICES

Appendix Table 1. Univariate and multivariate analysis of risk factors for recurrence free survival, according to ABOc/ABOi and TPE numbers.

Variables		Univariable HR (95% CI)	<i>P</i>	Multivariable HR (95% CI)	<i>P</i>
ABOi group	ABOc	Reference		Reference	
	ABOi/TPE<6	1.13 (0.68-1.90)	0.633	1.08 (0.63-1.85)	0.777
	ABOi/TPE≥6	2.51 (1.42-4.46)	0.002	1.99 (1.02-3.86)	0.042
Age	Years	0.99 (0.96-1.01)	0.331	-	
Sex	Female	0.81 (0.48-1.36)	0.427	-	
BMI		0.93 (0.88-1.00)	0.037	0.95 (0.89-1.01)	0.102
LT year	2011-2015	Reference		Reference	
	2016-2019	1.25 (0.78-2.00)	0.363		
	2020-2022	1.53 (0.93-2.54)	0.095		
Underlying for HCC	Hepatitis B	Reference			
	Hepatitis C	0.84 (0.39-1.82)	0.659	-	
	Non-B, Non-C	0.54 (0.28-1.04)	0.067	-	
Hypertension	Yes	0.88 (0.55-1.41)	0.587	-	
Diabetes mellitus	Yes	0.94 (0.61-1.42)	0.756	-	
Pretransplant MELD		1.07 (1.04-1.10)	<0.001	1.06 (1.03-1.09)	<0.001
Donor age	Years	1.00 (0.99-1.02)	0.650	-	
Donor sex	Female	0.83 (0.56-1.25)	0.374	-	
GRWR†	< 0.8	1.59 (0.85-2.98)	0.145	-	
Macrovesicular steatosis	≥ 10%	0.58 (0.29-1.16)	0.122	-	
Cold ischemic time	Minutes	1.00 (1.00-1.01)	0.019	1.00 (1.00-1.01)	0.622
Transfusion RBC	Packs	1.03 (1.02-1.04)	<0.001	1.02 (1.01-1.04)	0.002
Log_AFP	at LT	1.24 (1.12-1.38)	<0.001	1.11 (0.98-1.25)	0.093
Log_PIVKA	at LT	1.35 (1.22-1.49)	<0.001	1.03 (0.90-1.18)	0.659
Hepatectomy history	Yes	1.37 (0.87-2.14)	0.173	-	
Pretransplant LRT	Yes	2.96 (1.58-5.54)	0.001	2.91 (1.45-5.84)	0.003
Systemic treatment	Yes	3.66 (2.40-5.59)	<0.001	2.20 (1.37-3.53)	0.001
Total necrosis	Yes	1.26 (0.78-2.03)	0.352	-	
Viable tumor number		1.04 (1.02-1.05)	<0.001	1.02 (1.01-1.04)	0.007
Maximum tumor size	cm	1.10 (1.05-1.17)	<0.001	0.90 (0.82-0.98)	0.019
Microvascular invasion	Yes	2.64 (1.79-3.90)	<0.001	1.77 (0.97-3.22)	0.062
Poor differentiation	Yes	2.25 (1.53-3.31)	<0.001	1.30 (0.82-2.05)	0.268
Satellite nodule	Yes	3.62 (2.34-5.60)	<0.001	1.67 (0.90-3.10)	0.101

PVTT	Yes	6.14 (2.67-14.12)	<0.001	2.83 (0.98-8.16)	0.054
------	-----	-------------------	--------	------------------	-------

† Graft weight was directly measured during operation.

ABOc, ABO compatible; ABOi, ABO incompatible; TPE, therapeutic plasma exchange; HR, hazard ratio; CI, confidence interval; BMI, body mass index; LT, liver transplantation; HCC, hepatocellular carcinoma; MELD, model for end-stage liver disease; GRWR, graft recipient weight ratio; AFP, alpha-feto protein; PIVKA, protein induced by vitamin K antagonist-II; LRT, locoregional treatment; PVTT, portal vein tumor thrombosis.

Appendix Table 2. Univariate and multivariate analysis of risk factors for HCC recurrence, according to ABOc/ABOi and TPE numbers.

Variables		Univariable HR (95% CI)	P	Multivariable HR (95% CI)	P
ABOi group	ABOc	Reference		Reference	
	ABOi/TPE<6	0.90 (0.44-1.84)	0.770	0.97 (0.46-2.01)	0.928
	ABOi/TPE≥6	2.67 (1.31-5.46)	0.007	2.42 (1.05-5.57)	0.037
Age	Years	0.95 (0.92-0.98)	0.004	0.96 (0.92-1.00)	0.048
Sex	Female	0.58 (0.28-1.23)	0.155	-	
BMI		0.95 (0.88-1.03)	0.208	-	
LT year	2011-2015	Reference		Reference	
	2016-2019	1.01 (0.57-1.78)	0.972	-	
	2020-2022	0.99 (0.52-1.89)	0.972	-	
Underlying for HCC	Hepatitis B	Reference			
	Hepatitis C	0.56 (0.17-1.78)	0.326	-	
	Non-B, Non-C	0.42 (0.17-1.06)	0.066	-	
Hypertension	Yes	0.61 (0.31-1.20)	0.153	-	
Diabetes mellitus	Yes	0.62 (0.34-1.12)	0.116	-	
Pretransplant MELD		0.96 (0.90-1.02)	0.157	-	
Donor age	Years	0.98 (0.96-1.01)	0.196	-	
Donor sex	Female	0.84 (0.50-1.40)	0.507	-	
GRWR†	< 0.8	1.17 (0.47-2.92)	0.736	-	
Macrovesicular steatosis	≥ 10%	0.52 (0.21-1.29)	0.156	-	
Cold ischemic time	Minutes	1.00 (1.00-1.01)	0.390	-	
Transfusion RBC	Packs	1.02 (1.00-1.04)	0.039	-	
Log_AFP	at LT	1.38 (1.22-1.57)	<0.001	1.09 (0.94-1.25)	0.260
Log_PIVKA	at LT	1.43 (1.26-1.61)	<0.001	1.14 (0.98-1.34)	0.091
Hepatectomy history	Yes	1.57 (0.90-2.73)	0.113	-	
Pretransplant LRT	Yes	5.44 (1.98-14.98)	0.001	7.00 (2.02-24.26)	0.002
Systemic treatment	Yes	3.99 (2.33-6.84)	<0.001	2.10 (1.16-3.82)	0.015
Total necrosis	Yes	0.71 (0.34-1.49)	0.362	-	
Viable tumor number		1.05 (1.03-1.07)	<0.001	1.04 (1.01-1.07)	0.004
Maximum tumor size		1.14 (1.06-1.22)	<0.001	0.92 (0.82-1.03)	0.150
Microvascular invasion	Yes	4.85 (2.96-7.95)	<0.001	2.07 (1.01-4.24)	0.046
Poor differentiation	Yes	3.39 (2.06-5.58)	<0.001	1.69 (0.95-3.02)	0.076
Satellite nodule	Yes	6.55 (3.94-10.91)	<0.001	1.83 (0.92-3.63)	0.085
PVTT	Yes	6.31 (1.96-20.31)	0.002	2.49 (0.57-10.86)	0.226

† Graft weight was directly measured during operation.

HCC, hepatocellular carcinoma; ABOc, ABO compatible; ABOi, ABO incompatible; TPE, therapeutic plasma

exchange; HR, hazard ratio; CI, confidence interval; BMI, body mass index; LT, liver transplantation; MELD, model for end-stage liver disease; GRWR, graft recipient weight ratio; AFP, alpha-feto protein; PIVKA, protein induced by vitamin K antagonist-II; LRT, locoregional treatment; PVTT, portal vein tumor thrombosis.

Abstract (In Korean)

ABO 혈액형 부적합 생체기증자 간이식에서 이식 전 6회 이상의 치료적 혈장 교환술과 간세포암 재발 증가 간의 연관성

배경

이전 연구들에서는 간세포암(HCC) 환자에서 ABO 부적합 생체기증자 간이식과 ABO 적합 생체 간이식의 중양학적 결과가 유사하다고 보고하였습니다. 우리는 ABO 부적합 생체기증자 간이식에서 치료적 혈장교환의 횟수와 간세포암의 결과 간의 관계를 분석하고자 하였습니다.

방법

본 연구는 단일 기관의 후향적 연구로, 간세포암을 가진 428명의 생체기증자 간이식 수혜자를 ABO 적합성 여부와 이식 전 시행한 치료적 혈장교환술의 횟수에 따라 세 그룹으로 분류하였습니다. 치료적 혈장교환술 횟수의 기준은 무재발 생존기간에 대한 Cubic-Spline 모델을 기반으로 6회 이상으로 설정되었으며, 그룹은 1. ABO 적합군 (ABOc, n=323), 2. ABO 부적합군 & 치료적 혈장교환술 6회 미만 시행 (ABOi/TPE<6, n=75), 3. ABO 부적합군 & 치료적 혈장교환술 6회 이상 시행 (ABOi/TPE≥6, n=30)으로 나뉘었습니다. 무재발 생존기간과 간세포암 재발률은 간세포암 결과에 영향을 미치는 다른 위험 요인을 조정한 후 비교하였습니다.

결과

세 그룹은 대부분의 인구통계학적 특성, 중앙표지자 및 병리적 특성에서 유사한 특성을 보였습니다. ABOi/TPE<6 그룹의 초기 동종응집소(IA) 역가는 1:64 (범위: 음성-1:512) 였으며, ABOi/TPE≥6 그룹에서는 1:512 (범위: 1:128-1:4096) 였습니다. 5년 무재발 생존기간은 ABOi/TPE≥6 그룹에서 유의하게 낮았으며 (ABOc 75.7% vs. ABOi/TPE<6 72.7% vs. ABOi/TPE≥6 50.0%, P=0.005), 간세포암 재발률은 ABOi/TPE≥6 그룹에서 유의하게 높았습니다 (16.4% vs. 17.0% vs. 39.4%, P=0.014). 다변량 Cox 회귀 분석에서 ABOi/TPE≥6은 무재발 생존기간 (aHR 1.99 CI 95% 1.02-3.86, P=0.042) 및 간세포암 재발 (aHR 2.42 CI 95% 1.05-5.57, P=0.037)에 대한 독립적인 위험 인자로 나타났습니다.

결론

간세포암 환자에게서 생체기증자 간이식 수술 전 치료적 혈장교환술을 6회 이상 시행하는 것은 ABO 부적합 생체기증자 간이식 후 간세포암 재발 위험을 높일 수

있습니다. 간세포암 환자를 대상으로 ABO 부적합 생체기증자 간이식을 계획할 때, 면역학적 위험을 유사하게 유지하면서 치료적 혈장교환술 횟수를 6회 미만으로 줄이는 전략이 필요할 것입니다.

핵심되는 말 : ABO 부적합 생체기증자 간이식, 간세포암, 치료적 혈장교환술, 종양학적 결과