



저작자표시-비영리-동일조건변경허락 2.0 대한민국

이용자는 아래의 조건을 따르는 경우에 한하여 자유롭게

- 이 저작물을 복제, 배포, 전송, 전시, 공연 및 방송할 수 있습니다.
- 이차적 저작물을 작성할 수 있습니다.

다음과 같은 조건을 따라야 합니다:



저작자표시. 귀하는 원저작자를 표시하여야 합니다.



비영리. 귀하는 이 저작물을 영리 목적으로 이용할 수 없습니다.



동일조건변경허락. 귀하가 이 저작물을 개작, 변형 또는 가공했을 경우에는, 이 저작물과 동일한 이용허락조건하에서만 배포할 수 있습니다.

- 귀하는, 이 저작물의 재이용이나 배포의 경우, 이 저작물에 적용된 이용허락조건을 명확하게 나타내어야 합니다.
- 저작권자로부터 별도의 허가를 받으면 이러한 조건들은 적용되지 않습니다.

저작권법에 따른 이용자의 권리는 위의 내용에 의하여 영향을 받지 않습니다.

이것은 [이용허락규약\(Legal Code\)](#)을 이해하기 쉽게 요약한 것입니다.

[Disclaimer](#)

**Effectiveness of carbon dioxide per-conditioning
before recanalization in hyperacute ischemic stroke
rat model**

Jae Wook Jung

**The Graduate School
Yonsei University
Department of Medicine**

**Effectiveness of carbon dioxide per-conditioning
before recanalization in hyperacute ischemic stroke
rat model**

**A Dissertation submitted
to the Department of Medicine
and the Graduate School of Yonsei University
in partial fulfillment of the
requirements for the degree of
Doctor of Philosophy in Medical Science**

Jae Wook Jung

May 2024

**This certifies that the Dissertation
of Jae Wook Jung is approved.**

Thesis Supervisor [Hyo Suk Nam]

Thesis Committee Member [Young Dae Kim]

Thesis Committee Member [Kee Ook Lee]

Thesis Committee Member [Il Kwon]

Thesis Committee Member [Jinkwon Kim]

**The Graduate School
Yonsei University
June 2024**

ACKNOWLEDGEMENTS

I am fortunate to be able to spend difficult times with so many priceless people at once. I want to extend my sincere gratitude to every one of them.

First, I would especially appreciate Dr. Hyo Suk Nam of the Department of Neurology as my thesis supervisor. He offered encouragement and invaluable research guidance. His profound teachings will stay with me for a lifetime. I would also like to express my sincere appreciation to my four advisors; Professor Young Dae Kim, Kee Ook Lee, Il Kwon, and Jinkwon Kim, who took the time to guide my thesis while medical examination and/or researching. On top of that, I'm deeply grateful to my lab advisor, Chung Eun Yoon, for her genuine interest and guidance. I sincerely appreciate her assistance.

Second, I would like to express my gratitude to my wife, So Min Park, and our very adorable and cute baby, Do Yoon Jung. My wife is a very precious person who waited silently by my side, always gave me strength, and comforted me when I had a hard time. This work would not have been possible without her precious support. I sincerely appreciate her sacrifice.

Finally, I would like to extend my deepest gratitude to my cherished mother and sister for their unwavering belief, support, love, and kind gestures. My existence is grounded in my family's consistent love, trust, and support. My father deeply wished to attend my graduation while he was alive. I believe that even from heaven, he'll be with me in spirit. Though he's no longer here, I want to convey my heartfelt gratitude for his love and support through this letter.

I am currently positioned in front of a fresh beginning line before I graduate. The knowledge I gained while earning my doctorate would be an excellent starting point for me to become a humble scientist. I'll never forget their help and compassion, once more.

TABLE OF CONTENTS

LIST OF FIGURES	iii
LIST OF TABLES	iv
ABSTRACT IN ENGLISH	v
1. INTRODUCTION	1
2. MATERIALS AND METHODS	2
2.1. Ethics statement and animals	2
2.2. Experimental design and protocol	3
2.3. Sample size and experimental group	5
2.4. Assessment of neurologic function	7
2.5. Arterial blood gas analysis	7
2.6. Infarct size measurement	7
2.7. Immunofluorescence analysis	8
2.8. Western blot analysis	9
2.9. Statistical analysis	10
3. RESULTS	11
3.1. Physiologic variables before and after CO ₂ per-conditioning	11
3.2. CO ₂ per-conditioning attenuates infarct size after tMCAO	14
3.3. CO ₂ per-conditioning alleviates neurologic deficit after tMCAO	16
3.4. CO ₂ per-conditioning downregulates peri-lesional oxidative stress after tMCAO	17
3.5. CO ₂ per-conditioning alleviates BBB failure after tMCAO	19
3.6. CO ₂ per-conditioning upregulates tight junction protein expressions after tMCAO	21
3.7. CO ₂ per-conditioning upregulates PDGFR β expressions after tMCAO	23

4. DISCUSSION	25
5. CONCLUSION	29
REFERENCES	30
ABSTRACT IN KOREAN	35

LIST OF FIGURES

<Fig 1> Experimental design	4
<Fig 2> Study groups	6
<Fig 3> Infarct size measurement	14
<Fig 4> Behavior test score of Garcia, Longa, mNSS, and rotarod test	16
<Fig 5> CO ₂ per-conditioning decreased the number of 8-OHdG positive cells	18
<Fig 6> CO ₂ per-conditioning decreased the number of MMP-9 /PDGFR β double positive cells	20
<Fig 7> CO ₂ per-conditioning increased the number of ZO-1 positive cells	22
<Fig 8> CO ₂ per-conditioning increased the number of PDGFR β positive cells	24
<Fig 9> Schematic flow of effectiveness of CO ₂ per-conditioning in acute ischemia	27

LIST OF TABLES

<Table 1> ABGA results of experimental animals with pre and post CO ₂ inhalation	12
<Table 2> Infarct size measurement	15

ABSTRACT

Effectiveness of carbon dioxide per-conditioning before recanalization in hyperacute ischemic stroke rat model

Background: This study investigated whether carbon dioxide (CO₂) per-conditioning reduced oxidative stress, blood-brain barrier breakdown, and neurologic deficit in a rat model of middle cerebral artery occlusion (MCAO).

Methods: MCAO rat models received intermittent inhalation of mixed gas (20% CO₂, 20% O₂, 60% N₂) or room air. After the surgery, arterial blood gas analysis and behavior test were conducted, and animals were euthanized for calculating infarct size, western blot analysis, and immunohistochemistry.

Results: Our results showed that CO₂ per-conditioning reduced infarct size and neurological deficit. Number of 8-OHdG positive cell and MMP-9/PDGFR β double positive cell expressions were significantly decreased after CO₂ per-conditioning. The expression of tight junction protein and PDGFR β were significantly elevated after CO₂ per-conditioning.

Conclusion: CO₂ per-conditioning not only protects the ischemic penumbra from ischemia-reperfusion injury and reduces neurologic deficit, but also maintains the integrity of blood-brain barrier and neurovascular unit, and reduces oxidative stress in a hyperacute stroke rat model. CO₂ per-conditioning is promising and requires further assessment for clinical application.

Key words: carbon dioxide, per-conditioning, ischemic rat model, pericyte, oxidative stress, blood-brain barrier, ischemia-reperfusion injury

Effectiveness of carbon dioxide pre-conditioning before recanalization in hyperacute ischemic stroke rat model

1. Introduction

Acute stroke is the second leading cause of morbidity and mortality worldwide.¹ Ischemic stroke accounts for 85% of all strokes followed by hemorrhagic stroke.^{2,3} A significant proportion of patients with a history of acute ischemic stroke experience long-term disability, resulting in substantial healthcare costs. Nevertheless, clinicians have limited treatment options for hyperacute ischemic stroke, which mainly consist of intravenous thrombolytic therapy and endovascular treatment (EVT). Up until now EVT is considered the most effective treatment for hyperacute ischemic stroke with large vessel occlusion.⁴ Due to developments in mechanical devices and the technical skills associated with EVT, the recanalization rate has dramatically increased to approximately 80%.⁵ However, the rate of favorable outcomes remains at approximately 50%, despite proper recanalization with timely treatment.⁶

During the post-reperfusion state, the rapid restoration of blood flow leads to an elevation in tissue oxygenation levels, triggering a surge in the generation of reactive oxygen species (ROS).⁷ It makes ischemia-reperfusion injury that affects peri-lesional penumbra.⁸ Ischemia-reperfusion injury has multiple mechanisms to make cell dysfunction like overproduction of ROS, consumption of antioxidants, inflammation, blood-brain barrier (BBB) disruption, apoptosis, and increasing excitotoxicity.^{9,10} As a result, cerebral damage expands to encompass the penumbra and adjacent brain parenchyma.³

Therefore, how to avoid or reduce ischemia-reperfusion injury is the prominent point of treatment, which is called neuroprotection.^{3,11} Until recently, a number of neuroprotective agents were developed and subjected to clinical trials; however, despite their success in preclinical studies, none

of these agents have proven successful in clinical trials.¹²⁻¹⁴

Most of the neuroprotective trials are aimed for pre-conditioning mechanism. It is not relevant to clinical setting because acute ischemic stroke is unpredictable event, therefore, pre-conditioning trials may have limitations in the real life.¹⁵ Rather than pre-conditioning, per-conditioning may be more suitable because this intervention is carried out between onset of ischemia and recanalization. Clinically, per-conditioning corresponds to a treatment just before EVT. In addition, a recent clinical trial demonstrated the successful implementation of per-conditioning in conjunction with EVT.¹⁶

Carbon dioxide (CO₂) is harmless gas and has powerful vasodilative effects on the cerebral vessels.¹⁷ CO₂ is maintained within narrow range (35–45 mmHg) in physiologic state and has an intense diffusion capacity with an ability to cross the BBB. Several studies showed that arterial carbon dioxide tension (PaCO₂) is strong determinant of cerebral blood flow. Furthermore, CO₂ has anti-oxidant, anti-epileptic, and anti-inflammatory effects.^{15,18-23}

This study aims to show that intermittent CO₂ inhalation during the per-conditioning phase can improve outcomes in hyperacute ischemic stroke animal models by reducing oxidative stress and blood-brain barrier disruption, and saves the neurovascular unit, especially pericytes.

2. MATERIALS AND METHODS

2.1. Ethics statement and animals

All experiments were carried out in accordance with Institutional Animal Care and Use Committee of Yonsei University Health System, and the protocol was approved by the Institutional Animal Care and Use Committee of Yonsei University Health System (approval number: 2020-0331). All efforts were made to minimize suffer and the number of rats. In total, 42 male Wistar rat (8-week-old, 270–320g) were obtained (ORIENT BIO, Seongnam, South Korea). Experimental animals could freely access to food and water. The animal room was adjusted on a 12-h light/dark cycle with constant temperature of 22°C ± 2°C.

2.2. Experimental design and protocol

Before the operation, rotarod training had been performed for three days. A heating lamp were used to keep the rectal body temperature at 36.6–37.4°C. Wistar rats were deeply anesthetized with 4.5% isoflurane and maintained with 1.5% isoflurane using a mask during surgery. We made middle cerebral artery infarction models. In the process of induction of middle cerebral artery infarction, first, the animal was placed in a supine position, midline neck incision was made, and the soft tissues over the trachea were gently retracted. Second, the left common carotid artery (CCA) was separated from the vagus nerve and then ligated. Following that, the left external carotid artery (ECA) was ligated consecutively. Third, both the occipital artery and pterygopalatine artery were ligated together to obstruct the blood flow. Fourth, the microvascular clip was temporarily positioned in the left internal carotid artery (ICA) close to the CCA junction and the tied section of ECA was incised using microscissors to insert the monofilament (4035556PK10; Doccol Co., Albuquerque, NM, USA). Finally, the filament was cautiously advanced up to 20 mm into the ICA to occlude the origin of the middle cerebral artery (MCA). MCA was occluded for 90 min. After 30 min from induction of middle cerebral artery occlusion (MCAO), the rat began inhaling a mixed gas (20% CO₂, 20% O₂, 60% N₂; flow rate = 1L/min) with 3 cycles of inhalation for 5 min followed by 10 min recovery session (normoxic atmosphere). At the end of 90 min transient MCAO (tMCAO), the rats were anesthetized, and the microfilament was removed for recanalization of the MCA. Sham animals underwent the surgical procedure without any occlusion, while the animals with permanent MCAO (pMCAO) did not undergo the microfilament removal procedure (Figure 1).

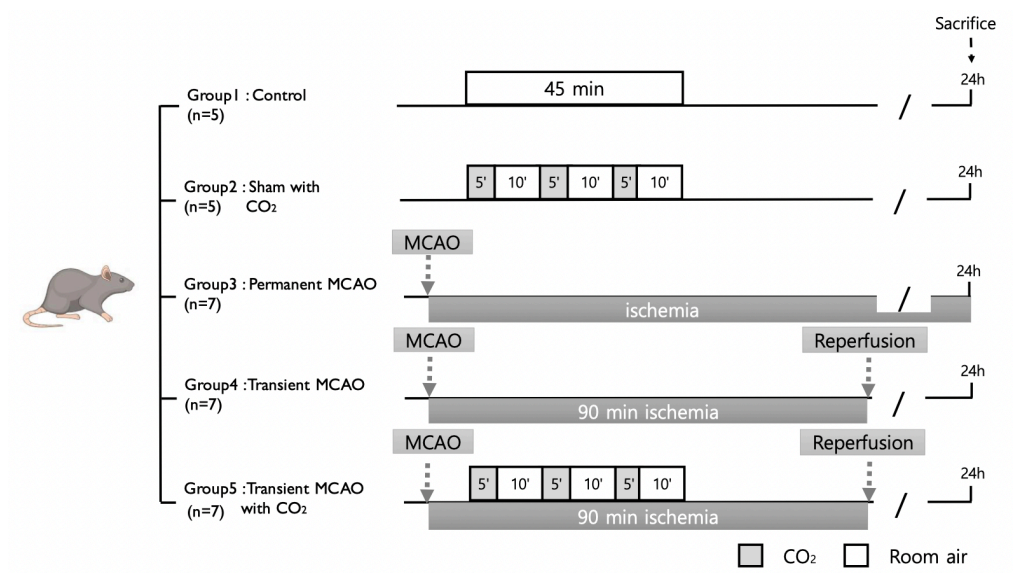


Figure 1. Experimental design.

After the operation, the rats were placed on a cage. The physiological variables including pH, pO₂, pCO₂, HCO₃ were recorded by arterial blood gas analysis before and after the CO₂ inhalation.

2.3. Sample size and experimental group

The total sample size for the study was determined using MedCalc statistical software version 22.017 (MedCalc Software by Ostend, Belgium), targeting an alpha level of 0.05 and a power of 0.8. The calculation was based on the mean difference in infarct size observed between the groups in a preliminary study. The comparison of two means' method indicated that a minimum of 25 animals were needed for the study. To account for an anticipated mortality rate and technical errors of 40%, the study included 42 rats in total. Forty-two rats practiced rotarod training, but 11 rats did not successfully complete the training. 31 rats were randomly allocated into five groups (Figure 2).

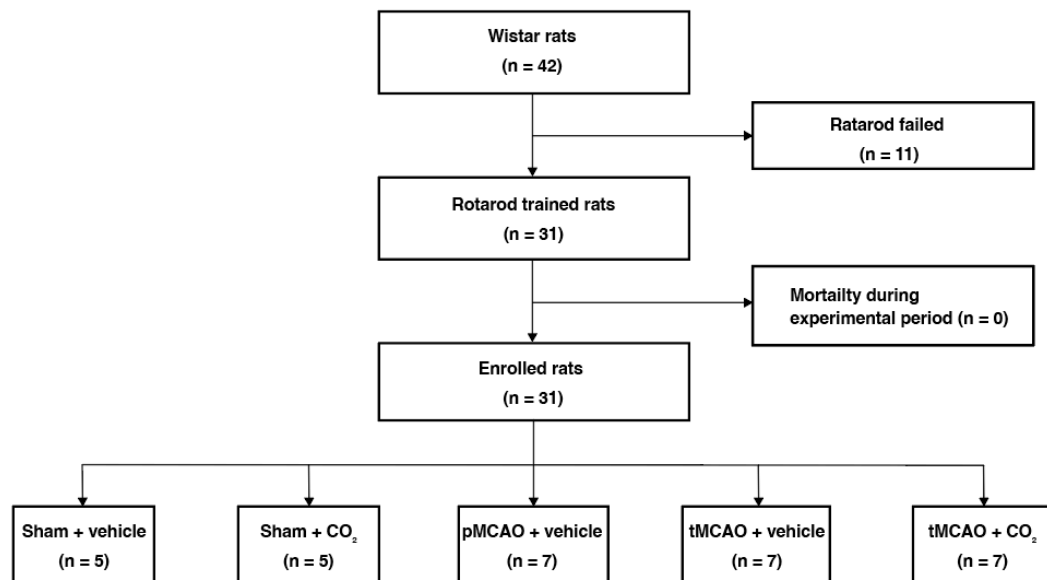


Figure 2. Study groups.

sham-operated rat with room air inhalation (Sham + vehicle group, n=5), sham-operated rat with intermittent CO₂ inhalation (sham + CO₂ group, n=5), pMCAO-operated rat with room air inhalation (pMCAO + vehicle group, n=7), tMCAO-operated rat with room air inhalation (tMCAO + vehicle group, n=7), and tMCAO-operated rat with intermittent CO₂ inhalation (tMCAO + CO₂ group, n=7).

2.4. Assessment of neurologic function

Neurologic evaluation was performed at 24 h after the initiation of the surgical procedure. We carried out various scoring methods for appropriate assessment – Garcia test, Longa test, rotarod motor test, and Modified Neurological Severity Score (mNSS). The neurologic deficits of the rats were examined by a blinded observer. To evaluate neurologic function, blinded investigator measured Longa test, Garcia test, rotarod motor test and, mNSS for motor, sensory and reflex of experimental animals. Baseline assessment of rotarod motor test was conducted. The Garcia test consists of a 6-factor scale with each factor having a maximum score of 3, with total scores ranging between 3 and 18.²⁴ The Longa test is a 5-point scale with a minimum score of 0 for no neurological deficit and a maximum score of 4 for no spontaneous walking and depressed.²⁵ The rotarod motor test, animals are placed on the rotarod cylinder, and duration the animals endured on the rotarod is measured.^{26,27} The last is mNSS, which included motor, sensory, reflex and balance tests. Neurological function is graded on a scale of 0 to 18.^{26,28} The higher the score, the more severe is the injury.²⁶

2.5. Arterial blood gas analysis

The artery on the ventral aspect of the experimental animal's tail was used for the collection of arterial blood. For baseline blood gas results, we collected arterial blood of whole rats before the operation, and in the groups with CO₂ inhalation, additional arterial blood was obtained immediately after the 3 cycles of inhalation. Arterial blood gases were analyzed using OPTI CCA blood gas and electrolyte analyzer (OPTI Medical System, Roswell, GA, USA).

2.6. Infarct size measurement

At 24 h after surgery, experimental animals were euthanized with intraperitoneal injection of trichloroacetaldehyde hydrate (0.4 g/kg) and followed by cardiac perfusion with 4% paraformaldehyde and normal saline. The brains were removed, dissected at 2 mm intervals, totally generating 7 sections and 4 of total sections were stained with 2% 2-3-5-triphenyl tetrazolium chloride (TTC) solution (T8877; Sigma-Aldrich, Darmstadt, Germany) and fixed in formalin. The infarct size of each section was measured by computer image analyzing software (Scion Image ®, Frederick, MD, USA). Infarct size was normalized to the contralateral area and expressed as a percentage. Average values from the 4 sections were calculated.

2.7. Immunofluorescence analysis

Immunofluorescence analysis was performed between 8-hydroxy-2-deoxyguanosine (8-OHdG), platelet derived growth factor receptor beta (PDGFR β) and matrix metalloproteinase 9 (MMP-9), zonula occludens-1 (ZO-1), or PDGFR β to analyze the oxidative damage of deoxyribonucleic acid (DNA), MMP-9 positive pericytes, tight junction expression, and pericytes expression. Staining was carried out using 4- μ m thick paraffin sections. A two-day immunofluorescence staining for paraffin protocol was performed. After being deparaffinized and rehydrated, the sections were steamed with epitope retrieval solution (IW-1100; IHC-TEK, Austin, TX, USA) using steamer (IHC World, Woodstock, MD, USA) for 40 min. After 20 min of cooling, the sections were rinsed by phosphate-buffered saline (PBS) and permeabilization was performed with 1xPBS-T (PBS with 0.3% triton X-100) for 10 min at room temperature. Then the sections were blocked with blocking solution (1% bovine serum albumin + 10% horse serum + 1xPBS-T) for 1 hr at room temperature. Subsequently, the sections were incubated in primary antibodies against PDGFR β (anti-rabbit monoclonal antibodies, 1:200 dilution, ab32570; Abcam, Cambridge, UK), ZO-1 (anti-rabbit polyclonal antibodies, 1:200 dilution, 61-7300; Thermo Fisher Scientific, Waltham, MA, USA), MMP-9 (anti-goat polyclonal antibodies, 1:100 dilution, sc-6841; Santa Cruz biotechnology, Dallas, TX, USA), and 8-OHdG (anti-mouse monoclonal antibodies, 1:200 dilution, ab48508; Abcam, Cambridge, UK)

overnight at 4°C. Second day, the sections were incubated with enzyme-conjugated secondary Ab (Alexa fluor 488-labeled goat anti-rabbit IgG antibody (1:200 dilution, ab150077; Abcam, Cambridge, UK), Alexa fluor 647-labeled goat anti-mouse IgG antibody (1:200 dilution, ab150115; Abcam, Cambridge, UK), and Alexa fluor 647-labeled donkey anti-goat IgG antibody (1:200 dilution, ab150131; Abcam, Cambridge, UK) for 1 hr at room temperature in the dark to avoid photobleaching. After washing by PBS for 3 times, the sections were mounted with antifade mounting medium with 4',6-diamidino-2-phenylindole (DAPI) (Vector Laboratories, Newark, CA, USA). The sections were scanned with a confocal microscope (LSM 710, Zeiss Instruments, Oberkochen, Germany). The investigator chose cortical or striatal penumbra region of interest (ROI) per section and observed the expression of 8-OHdG, PDGFR β /MMP-9, ZO-1 and PDGFR β . Quantification of the positively stained cells in the ROI and fluorescence intensity was performed with image analysis software (ImageJ, National Institutes of Health, Bethesda, MD, USA).

2.8. Western blot analysis

Proteins from the rat brain samples were extracted in homogenization buffer (50 mM Tris, 120 mM NaCl, pH 7.4) containing protease inhibitors (complete Mini, GIBCO, UK), separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), and transferred to polyvinylidene fluoride (PVDF) membrane (cat. #1620177, Bio-Rad Laboratories, Hercules, CA, USA). The blocked membranes were incubated with the primary antibody overnight at 4°C. The primary antibodies were against PDGFR β (anti-rabbit monoclonal antibodies, 1:1000 dilution, ab32570; Abcam, Cambridge, UK) and ZO-1 (anti-rabbit polyclonal antibodies, 1:500 dilution, 61-7300; Thermo Fisher Scientific, Waltham, MA, USA). After the membranes were rinsed, the membranes were incubated with enzyme conjugated secondary antibodies for 1 hour at room temperature. Horseradish peroxidase-conjugated anti-rabbit immunoglobulin G (1:10,000 dilution, ab205718; Abcam, Cambridge, UK) were used as secondary antibodies. Electrochemiluminescent detection was performed by LAS4000 (GE healthcare life sciences, NY, USA). Blot bands were quantified

with ImageJ software (v1.4, NIH, Bethesda, MD, USA). β -actin (sc-47778; Santa Cruz biotechnology, Dallas, TX, USA) was blotted on the membrane as a loading control. The quantified values were presented as a percentage of sham group (100%).

2.9. Statistical analysis

All the data in this study were presented as the median and interquartile range (IQR). Kruskal-Wallis H test was performed to compare the baseline results of whole groups, and Wilcoxon signed rank test was performed to compare pairwise results of arterial blood gas analyses of groups with CO₂ inhalation. Kruskal-Wallis H test followed by the post-hoc Dunn's multiple comparison was performed in score of behavior test and cell count of immunohistostaining. The differences were considered statistically significant at a value of $p < 0.05$, and all p-values are two-tailed. Statistical analyses were performed using R version 4.2.1 (R Foundation for Statistical Computing, Vienna, Austria).

3. RESULTS

3.1. Physiologic variables before and after CO₂ per-conditioning

No rat experienced mortality throughout the duration of the experiment (0/31, 0%). Body weight and rectal temperature were not statistically different among the experimental groups (data not shown). pH, pCO₂, pO₂, and HCO₃ levels of baseline arterial blood gas analyses did not show significant difference among the groups (For pH, $p = 0.8$, for pCO₂, $p > 0.9$, for pO₂, $p = 0.8$, and for HCO₃, $p > 0.9$). After the CO₂ inhalation, pH of sham + CO₂ and tMCAO + CO₂ groups were reduced ($p = 0.062$, $p = 0.022$, respectively, Table 1). The opposite patterns were shown in pCO₂ level of both groups.

Table 1 ABGA results of experimental animals with pre and post CO₂ inhalation

Experimental group	Before CO ₂ inhalation ¹	After CO ₂ inhalation ¹	p-value ²
pH			
Sham (median [IQR])	7.48 [7.45, 7.50]		
Sham + CO ₂	7.46 [7.45, 7.48]	7.40 [7.39, 7.42]	0.062
pMCAO	7.47 [7.45, 7.48]		
tMCAO	7.46 [7.45, 7.48]		
tMCAO + CO ₂	7.47 [7.47, 7.51]	7.31 [7.29, 7.44]	0.022
pCO₂ (mmHg)			
Sham	34.0 [27.0, 35.0]		
Sham + CO ₂	30.0 [29.0, 31.0]	38.0 [31.0, 41.0]	0.062
pMCAO	31.0 [23.0, 39.0]		
tMCAO	31.0 [28.0, 35.5]		
tMCAO + CO ₂	27.0 [24.0, 33.5]	40.0 [34.0, 45.5]	0.016
pO₂ (mmHg)			
Sham	134.0 [113.0, 143.0]		
Sham + CO ₂	135.0 [124.0, 144.0]	106.0 [64.0, 109.0]	0.062
pMCAO	143.0 [116.0, 148.5]		
tMCAO	135.0 [127.0, 148.5]		
tMCAO + CO ₂	146.0 [122.0, 153.0]	97.0 [84.5, 109.5]	0.016
HCO₃ (mEq/L)			
Sham	24.2 [17.4, 25.0]		
Sham + CO ₂	21.7 [21.5, 22.1]	19.4 [18.6, 26.1]	0.81
pMCAO	22.2 [16.7, 26.4]		
tMCAO	21.0 [20.4, 24.0]		
tMCAO + CO ₂	19.2 [18.7, 23.8]	21.1 [20.4, 21.7]	0.94

¹Data were shown as median [IQR].

²Wilcox signed rank test was performed for paired data.

Abbreviations: CO₂ = carbon dioxide; HCO₃ = bicarbonate; IQR = interquartile range; PCO₂ = partial pressure of carbon dioxide; pH = potential of hydrogen; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

3.2. CO₂ pre-conditioning attenuates infarct size after tMCAO

To evaluate the potential protective effects of intermittent CO₂ inhalation on the outcomes of rat models with MCAO, we assessed infarct size and conducted four different behavior tests to assess functional outcomes. In the TTC staining analyses, the tMCAO + CO₂ group showed a significantly smaller infarct size compared to both the tMCAO + vehicle and the pMCAO + vehicle group. (For tMCAO + vehicle, $p < 0.0001$; for pMCAO + vehicle, $p < 0.0001$, respectively, Figure 3). Between the tMCAO + vehicle and the pMCAO + vehicle groups, significant infarct size difference was found ($p < 0.0001$, Figure 3, Table 2).

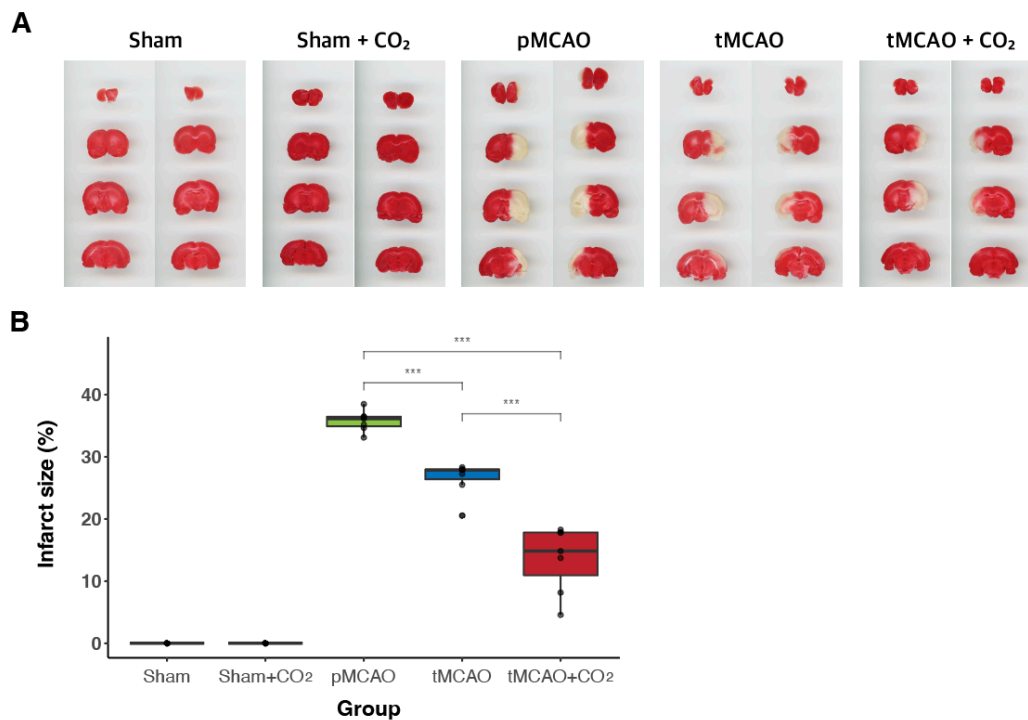


Figure 3. Infarct size measurement.

(A) Representative Images of TTC stain. (B) Quantitative analysis of infarct size according to the groups (** $p < 0.001$).

Abbreviations: CO₂ = carbon dioxide; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion; TTC = 2-3-5-triphenyl tetrazolium chloride.

Table 2 Infarct size measurement

Experimental group	Infarct size (%) ¹
Sham, median [IQR]	0
Sham + CO ₂	0
pMCAO	36.1 [34.9, 36.5]
tMCAO	27.9 [26.4, 28.0]
tMCAO + CO ₂	14.8 [11.0, 17.8]

¹Data were shown as median [IQR].

Abbreviations: CO₂ = carbon dioxide; IQR = interquartile range; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

3.3. CO₂ per-conditioning alleviates neurologic deficit after tMCAO

Regarding behavior tests, the tMCAO + CO₂ group exhibit lower mNSS scores compared to both the tMCAO + vehicle group and the pMCAO + vehicle group (Figure 4C). In addition, Garcia and rotarod test score of the tMCAO + CO₂ group were significantly higher than those of the other MCAO groups. (Figure 4A, 4D) In the Longa test, the tMCAO + CO₂ groups showed a lower median score compared to the other MCAO groups; however, this difference was not statistically significant (Figure 4B).

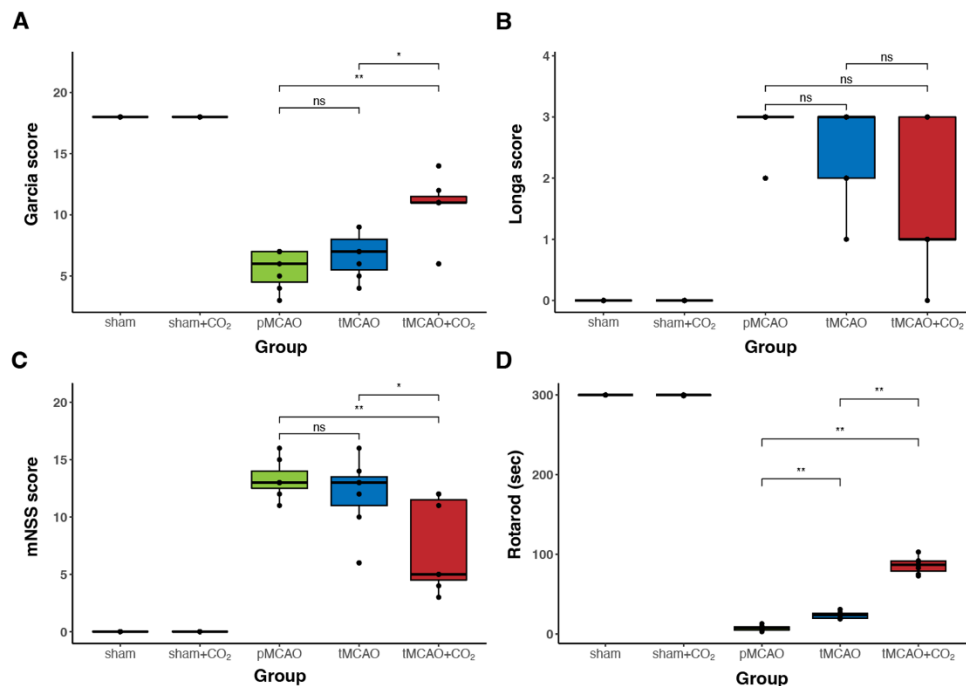


Figure 4. Behavior test score of Garcia, Longa, mNSS, and rotarod test.

Abbreviations: CO₂ = carbon dioxide; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

3.4. CO₂ per-conditioning downregulates peri-lesional oxidative stress after tMCAO

As shown in Figure 5, the number of 8-OHdG positive cells was significantly reduced in the tMCAO + CO₂ group compared to both the tMCAO + vehicle and pMCAO + vehicle groups ($p = 0.0026$, $p = 0.00058$, respectively). Furthermore, the pMCAO + vehicle group exhibited a significant increase in the number of 8-OHdG positive cells compared to the tMCAO + vehicle group ($p = 0.00058$). The sham + vehicle and sham + CO₂ groups showed a minimal presence of 8-OHdG positive cells.

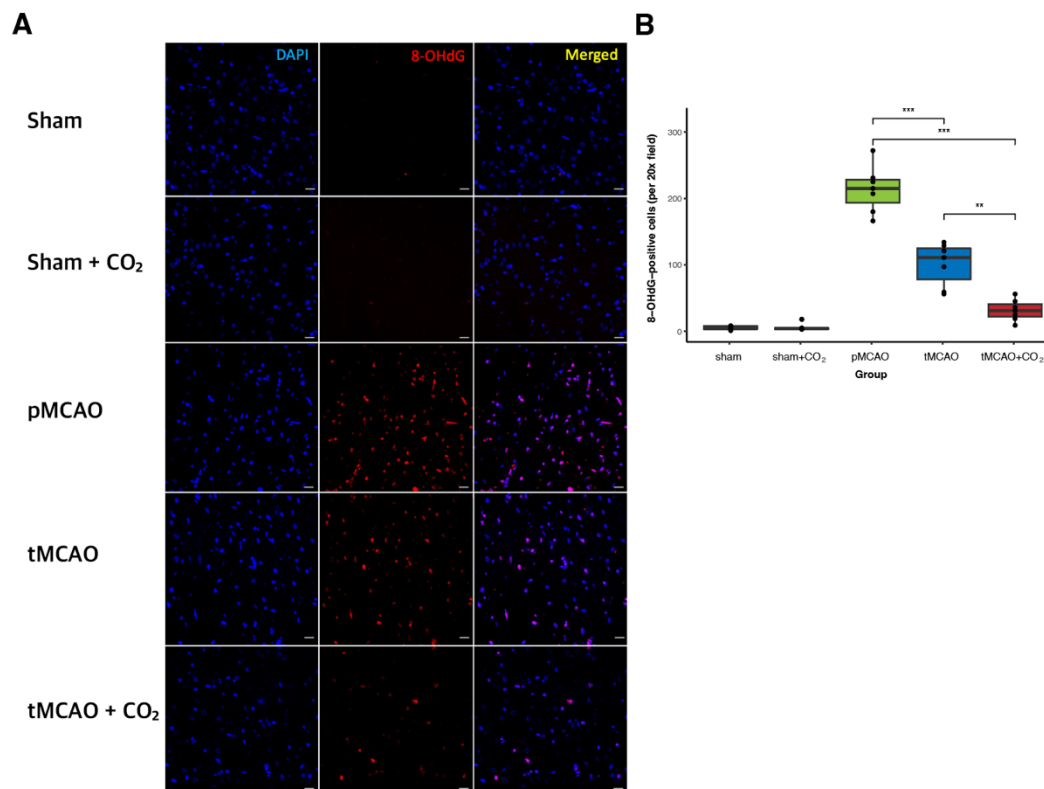


Figure 5. CO₂ per-conditioning decreased the number of 8-OHdG positive cells.

(A) Representative Images of immunofluorescence staining illustrated the 8-OHdG positive cells. scale bar = 20 μ m. (B) Quantitative analysis of 8-OHdG expression in immunofluorescence staining (** p < 0.01, *** p < 0.001). 8-OHdG expressions in all rat models presented similar pattern.

Abbreviations: 8-OHdG = 8-hydroxy-2-deoxyguanosine; CO₂ = carbon dioxide; DAPI = 4',6-diamidino-2-phenylindole; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

3.5. CO₂ per-conditioning alleviates BBB failure after tMCAO

In the peri-infarct areas, the tMCAO + CO₂ group showed a significant reduction in the number of MMP-9/PDGFR β double positive cells compared to the other MCAO groups ($p = 0.0014$, $p = 0.011$, respectively, Figure 6). Compared to the tMCAO + vehicle group, the pMCAO + vehicle group revealed a higher expression of MMP-9/PDGFR β double positive cells ($p = 0.016$, Figure 6).

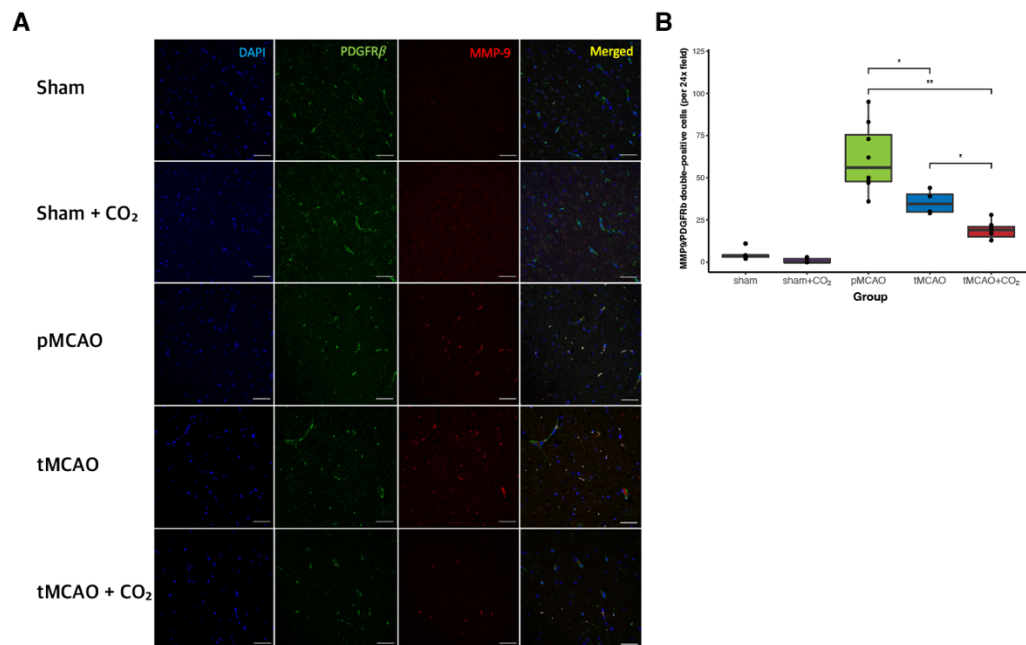


Figure 6. CO₂ per-conditioning decreased the number of MMP-9/PDGFR β double positive cells.

(A) Representative Images of immunofluorescence staining illustrated the PDGFR β , MMP-9, and MMP-9/PDGFR β double positive cells. scale bar = 20 μ m. (B) Quantitative analysis of MMP-9/PDGFR β expression in immunofluorescence staining (* $p < 0.05$, ** $p < 0.01$). MMP-9 and PDGFR β expressions in all rat models presented similar pattern.

Abbreviations: CO₂ = carbon dioxide; DAPI = 4',6-diamidino-2-phenylindole; MMP-9 = matrix metalloproteinase 9; PDGFR β = platelet derived growth factor receptor beta; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

3.6. CO₂ per-conditioning upregulates tight junction protein expressions after tMCAO

The tMCAO + CO₂ group showed reserved number of ZO-1 positive cells compared with the other MCAO groups ($p = 0.0019$, $p = 0.047$, respectively, Figure 7). Compared to the tMCAO + vehicle group, the pMCAO + vehicle group revealed a lower expression of ZO-1 positive cells ($p = 0.0019$, Figure 7). Western blot analysis showed that the tMCAO + CO₂ group had higher protein levels of ZO-1 than the other MCAO groups (Figure 7).

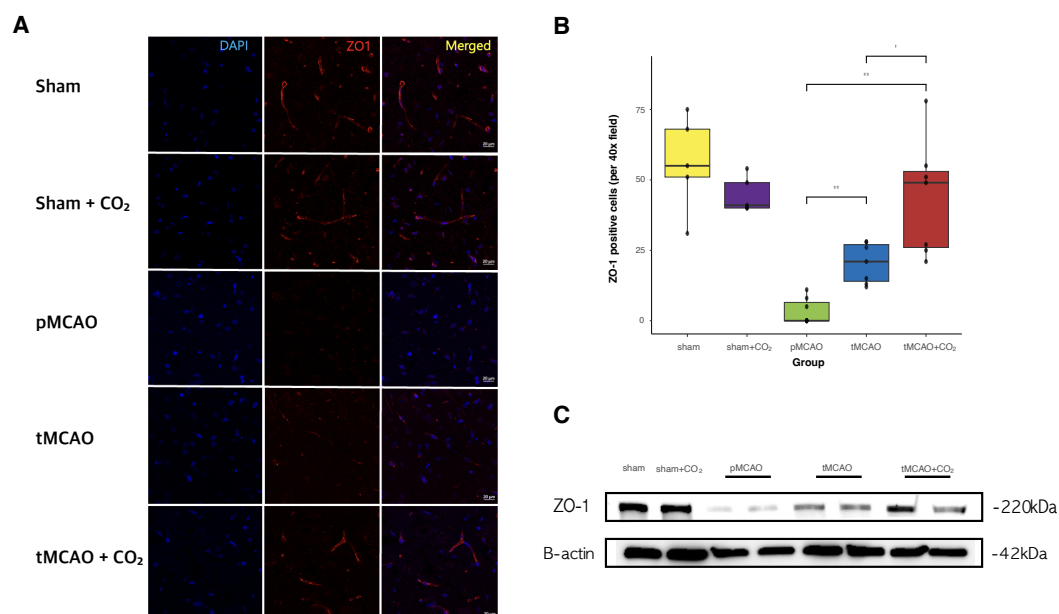


Figure 7. CO₂ per-conditioning increased the number of ZO-1 positive cells.

(A) Representative Images of immunofluorescence staining illustrated the ZO-1 positive cells. scale bar = 20 μ m. (B) Quantitative analysis of ZO-1 expression in immunofluorescence staining (* p < 0.05, ** p < 0.01). (C) Western blot analysis of ZO-1 expression. ZO-1 expressions in all rat models presented similar pattern.

Abbreviations: CO₂ = carbon dioxide; DAPI = 4',6-diamidino-2-phenylindole; ZO-1 = zonula occludens-1; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

3.7. CO₂ per-conditioning upregulates PDGFR β expressions after tMCAO

The tMCAO + CO₂ group showed reserved number of PDGFR β positive cells compared with the other MCAO groups ($p = 0.0021$, $p = 0.0025$, respectively, Figure 8). Western blot analysis showed that the tMCAO + CO₂ group had higher protein levels of PDGFR β than the other MCAO groups (Figure 8).

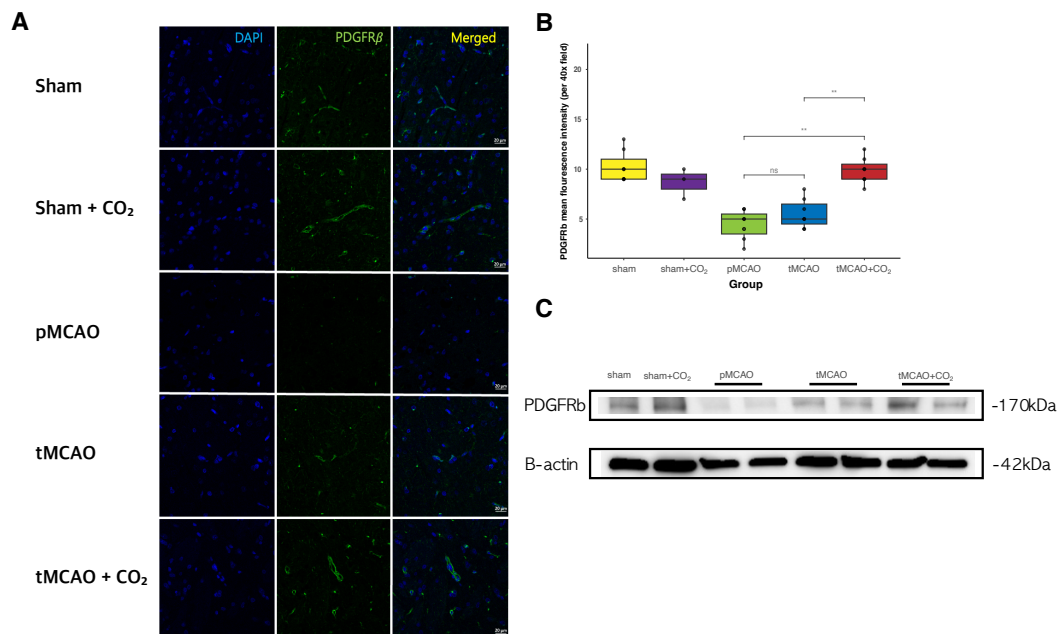


Figure 8. CO₂ per-conditioning increased the number of PDGFR β positive cells.

(A) Representative Images of immunofluorescence staining illustrated the PDGFR β positive cells. scale bar = 20 μ m. (B) Quantitative analysis of PDGFR β expression in immunofluorescence staining (** $p < 0.01$). (C) Western blot analysis of PDGFR β expression. PDGFR β expressions in all rat models presented similar pattern.

Abbreviations: CO₂ = carbon dioxide; DAPI = 4',6-diamidino-2-phenylindole; PDGFR β = platelet derived growth factor receptor beta; pMCAO = permanent middle cerebral artery occlusion; tMCAO = transient middle cerebral artery occlusion.

4. DISCUSSION

In the present study, we evaluated neuroprotective effect of intermittent CO₂ per-conditioning in tMCAO rat model. Taking advantage of experimental models, our data illustrated that CO₂ per-conditioning significantly decreased infarct size and neurologic deficit by reducing the oxidative stress, BBB breakdown, and death of pericytes after tMCAO. In addition, there were a link between structural lesion and functional deficit in our models.

Several studies have shown that mild acidosis via hypercapnia decreases infarct size and has a neuroprotective effect on cerebral ischemia in mouse or rat models.^{22,29-31} Previous studies have used pre-or post-conditioning methods, however, none of them performed hypercapnia intervention during ischemic period, which is per-conditioning. In this study, we focused on per-conditioning, that can be used in conjunction with the treatment of patients with hyperacute ischemic stroke. Additionally, CO₂ is fast acting vasodilatory gas and can cross BBB owing to its fat-soluble characteristics.^{32,33} Due to these specific features, CO₂ may exert a neuroprotective effect on ischemic state during per-conditioning. We suggest per-conditioning and induced mild hypercapnia will work synergistically with each other. Our study showed that intermittent CO₂ inhalation as a per-conditioning method is effective for neuroprotection. Future studies that compare this method with pre-conditioning or post-conditioning will further clarify the efficacy of per-conditioning.

The vasodilatory effect of hypercapnia is known to be modulated by vascular smooth muscle relaxation, which is caused by decreased endothelial intracellular Ca²⁺ level.^{34,35} Relaxation of cerebral artery smooth muscles are induced as PaCO₂ increases, which mediated through accompanying changes in pH and/or directly by CO₂.^{22,30,36} Cerebral blood flow (CBF) decreases by 2-3% for each mmHg decrease of PaCO₂ from its normal resting value owing to the vasoconstrictive effects of hypocapnia, conversely, CBF increases by 3-4% for each unit increase in PaCO₂, reaching when PaCO₂ is elevated by 10-20 mmHg above normal resting state.³⁷ In this study, we intermittently inhaled 20% CO₂, which is the most efficient CO₂ concentration identified in pilot and other studies.^{22,38} Based on ABGA results of CO₂ inhalation groups, the PaCO₂ values in our

study did not rise more than 20 mmHg from the baseline PaCO₂ value, indicating the absence of excessive hypercapnia. Since PaCO₂ is a major determinant of CBF regulation, especially in acute hypoxia, mild hypercapnia in cerebral ischemia models may have a significant impact on reversing the penumbra by alleviating the decrease in CBF.³⁷ In contrast, excessive increase in CO₂ concentration resulted in poor outcomes through increased intracranial pressure rather than neuroprotective effects.²⁹ In clinical field, recent multi-national, multi-center, randomized trial showed that mild hypercapnia is unlikely to cause clinically relevant elevations in intracranial pressure.³⁹ In this regard, it is critical to maintain adequate mild hypercapnia during ischemia for safety aspect. In clinical field, one randomized controlled trial demonstrated beneficial effect of targeted therapeutic mild hypercapnia after cardiac arrest.⁴⁰ In this clinical trial, the median difference of PaCO₂ between the intervention and control groups was 8 mmHg, and this result was similar to our ABGA results. Our findings could help to design future studies on the clinical efficacy of mild hypercapnia for patients with ischemic stroke.

In our study, we revealed that oxidative stress and BBB breakdown after cerebral ischemia can be attenuated by CO₂ pre-conditioning. Oxidative stress which is caused by excessive production of ROS due to imbalance between pro-oxidants and anti-oxidants is known as a key deleterious factor in brain ischemia. During the reperfusion phase, while thrombolysis therapy removed the blood clot, blood reperfusion can have detrimental effects on brain cells due to the excessive production of ROS.⁴¹ Initiation of reperfusion produces large amounts of superoxide by reducing NAD⁺ to NADH in mitochondrial respiratory chain, also ROS occurs in the cytoplasm through the reaction of NADPH-oxidase.⁴² One study showed that CO₂ can interact with reactive oxygen species, scavenge the peroxynitrite which is produced from superoxide, and prevent oxidative stress.⁴³ This anti-oxidative effect of CO₂ may have contributed to the reduction of oxidative stress induced by the ischemia-reperfusion state, as shown by the attenuation of 8-OHdG in our experimental results.

MMP-9 is well known mediator of BBB breakdown caused by degradation of basement membrane proteins in ischemia-reperfusion injury.^{44,45} In addition, high MMP-9 levels were associated with size of infarction and unsuccessful recanalization.^{46,47} Our result showed that mild hypercapnia attenuated MMP-9 expression and upregulated ZO-1. These findings suggested that mild

hypercapnia can decrease BBB breakdown by attenuating MMP-9 expression and increasing ZO-1 expression.

In the present study, we found that mild hypercapnia attenuated death of pericytes in ischemic state. Pericytes, one of neurovascular unit components, are perivascular mural cells on blood microvessels. Pericytes can stabilize the capillary wall, maintain the BBB, prevent neuroinflammation, and regulate the capillary diameter and blood flow.⁴⁸⁻⁵⁰ In the ischemic state, contraction of pericytes triggers capillary constriction, and then erythrocytes are trapped in microvessels.^{51,52} After recanalization of occluded vessels, pericytes remain contracted despite successful recanalization.⁵¹ It is a main cause of no-reflow phenomenon. As such, pericytes play an important role in the microcirculation environment. One study showed that increasing PaCO₂ induces relaxation of pericytes, whereas decreasing PaCO₂ causes contraction of pericytes.²¹ This study implies that CO₂ can affect alteration of pericytes, in this regard, the mechanism of beneficial roles of CO₂ may be mediated by pericytes. In addition, because pericytes contraction also can be induced by ROS, the anti-oxidant effect of hypercapnia may relieve pericytes contraction, which triggers capillary constriction.⁵¹

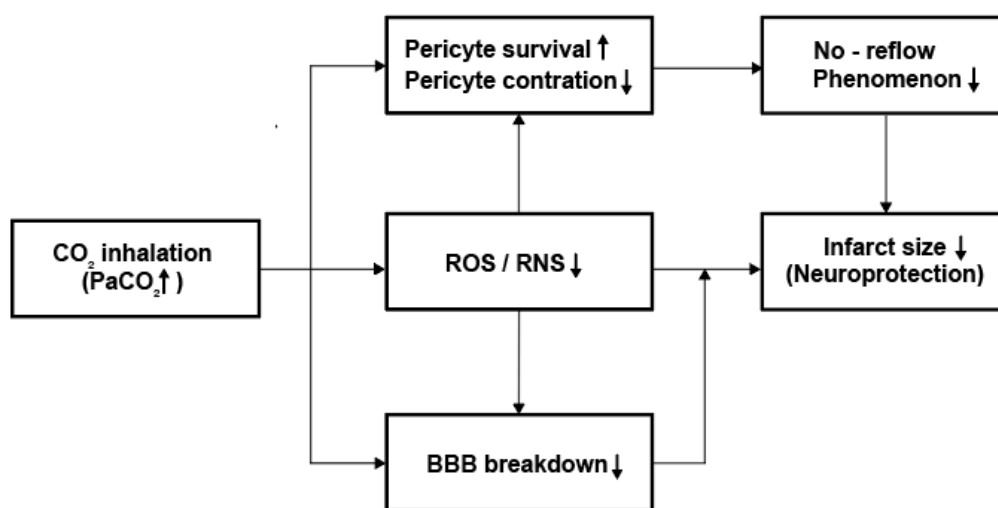


Figure 9. Schematic flow of effectiveness of CO₂ per-conditioning in acute ischemia.

These various effects of CO₂ inhalation during transient cerebral ischemia may alleviate size of ischemic stroke and neurologic deterioration (Figure 11). Due to its minimal reactivity and low risk in daily life, CO₂ can be used outside of hospital. As there is no significant clinical risk, the early initiation of mild hypercapnia can be considered even before patients with acute neurologic deficits arrive at the hospital. If the stability of mild hypercapnia is confirmed for conditions such as cerebral hemorrhage and infectious brain disease in the future, conducting clinical trials may lead to promising outcomes.

There are several limitations in this study. First, the relative limited observational period was followed (24 hr). In clinical fields, the modified Rankin Scale, which measures degree of disability or dependence after a stroke is evaluated at some weeks or even months after the stroke insult. If the rats equivalent of a human year is nearly 2 weeks, then the evaluations need to be prolonged to a week, or longer. Future studies need to clarify structural and functional changes of insulted experimental groups until weeks and even months. Second, stroke has sexually dimorphism in clinical field. Because women own a protective female sex hormone, they have a lower risk of stroke incidence relative to men.⁵³ The results from experimental stroke model may have difference in sex. From all these considerations, sex difference should be considered in further study to determine the effect of intermittent CO₂ inhalation in ischemic stroke models. Third, we did not measure cerebral blood flow in the present study. However, another study showed hypercapnia preserved cerebral blood flow in the ischemic state.⁵⁴ In addition, one study indicated hypercapnia has been reported to cause vasodilation and increase cerebral blood flow.⁵⁵ Finally, we did not measure concurrent PaCO₂ level during intervention and surgery. The follow-up ABGA results after CO₂ inhalation showed a lower PaCO₂ level than anticipated. This occurred because, despite conducting the ABGA procedure immediately after three cycle of CO₂ inhalation, there was at least 10-min gap from the last CO₂ inhalation due to the recovery phase.

5. CONCLUSION

Mild hypercapnia not only protects the ischemic penumbra from ischemia-reperfusion injury and reduces neurologic deficit, but also maintains the integrity of BBB and neurovascular unit, and reduces oxidative stress in a hyperacute stroke rat model. Therapeutic hypercapnia is promising and requires further assessment for clinical application.

References

1. Diseases GBD, Injuries C. Global burden of 369 diseases and injuries in 204 countries and territories, 1990-2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet* 2020;396:1204-22.
2. Hankey GJ. Stroke. *Lancet* 2017;389:641-54.
3. Gonzalez-Nieto D, Fernandez-Serra R, Perez-Rigueiro J, Panetsos F, Martinez-Murillo R, Guinea GV. Biomaterials to Neuroprotect the Stroke Brain: A Large Opportunity for Narrow Time Windows. *Cells* 2020;9.
4. Chen CJ, Ding D, Starke RM, Mehndiratta P, Crowley RW, Liu KC, et al. Endovascular vs medical management of acute ischemic stroke. *Neurology* 2015;85:1980-90.
5. Goyal M, Menon BK, van Zwam WH, Dippel DW, Mitchell PJ, Demchuk AM, et al. Endovascular thrombectomy after large-vessel ischaemic stroke: a meta-analysis of individual patient data from five randomised trials. *Lancet* 2016;387:1723-31.
6. Goyal M, Menon BK, van Zwam WH, Dippel DW, Mitchell PJ, Demchuk AM, et al. Endovascular thrombectomy after large-vessel ischaemic stroke: a meta-analysis of individual patient data from five randomised trials. *Lancet* 2016;387:1723-31.
7. Rodrigo R, Fernández-Gajardo R, Gutiérrez R, Matamala JM, Carrasco R, Miranda-Merchak A, et al. Oxidative stress and pathophysiology of ischemic stroke: novel therapeutic opportunities. *CNS Neurol Disord Drug Targets* 2013;12:698-714.
8. Shi K, Tian DC, Li ZG, Ducruet AF, Lawton MT, Shi FD. Global brain inflammation in stroke. *Lancet Neurol* 2019;18:1058-66.
9. Puig B, Brenna S, Magnus T. Molecular Communication of a Dying Neuron in Stroke. *Int J Mol Sci* 2018;19.
10. Orellana-Urzua S, Rojas I, Libano L, Rodrigo R. Pathophysiology of Ischemic Stroke: Role of Oxidative Stress. *Curr Pharm Des* 2020;26:4246-60.
11. Fisher M. The ischemic penumbra: identification, evolution and treatment concepts.

- Cerebrovasc Dis 2004;17 Suppl 1:1-6.
12. O'Collins VE, Macleod MR, Donnan GA, Horky LL, van der Worp BH, Howells DW. 1,026 experimental treatments in acute stroke. *Ann Neurol* 2006;59:467-77.
 13. Neuroprotection: the end of an era? *Lancet* 2006;368:1548.
 14. Ghozy S, Reda A, Varney J, Elhawary AS, Shah J, Murry K, et al. Neuroprotection in Acute Ischemic Stroke: A Battle Against the Biology of Nature. *Front Neurol* 2022;13:870141.
 15. Sun MS, Jin H, Sun X, Huang S, Zhang FL, Guo ZN, et al. Free Radical Damage in Ischemia-Reperfusion Injury: An Obstacle in Acute Ischemic Stroke after Revascularization Therapy. *Oxid Med Cell Longev* 2018;2018:3804979.
 16. Renu A, Millan M, San Roman L, Blasco J, Marti-Fabregas J, Terceno M, et al. Effect of Intra-arterial Alteplase vs Placebo Following Successful Thrombectomy on Functional Outcomes in Patients With Large Vessel Occlusion Acute Ischemic Stroke: The CHOICE Randomized Clinical Trial. *JAMA* 2022;327:826-35.
 17. Asami K, Nakashima T, Morisaki H, Akanabe K, Kuno K, Yanagita N. Effects of hypercapnia on cochlear and cerebral blood flow in rabbits. *ORL J Otorhinolaryngol Relat Spec* 1995;57:239-44.
 18. Curley G, Laffey JG, Kavanagh BP. Bench-to-bedside review: carbon dioxide. *Crit Care* 2010;14:220.
 19. Tolner EA, Hochman DW, Hassinen P, Otahal J, Gaily E, Haglund MM, et al. Five percent CO(2) is a potent, fast-acting inhalation anticonvulsant. *Epilepsia* 2011;52:104-14.
 20. Kavanagh BP, Laffey JG. Hypercapnia: permissive and therapeutic. *Minerva Anesthesiol* 2006;72:567-76.
 21. Chen Q, Anderson DR. Effect of CO₂ on intracellular pH and contraction of retinal capillary pericytes. *Invest Ophthalmol Vis Sci* 1997;38:643-51.
 22. Fan YY, Shen Z, He P, Jiang L, Hou WW, Shen Y, et al. A novel neuroprotective strategy for ischemic stroke: transient mild acidosis treatment by CO₂ inhalation at reperfusion. *J Cereb Blood Flow Metab* 2014;34:275-83.
 23. Vannucci RC, Towfighi J, Heitjan DF, Brucklacher RM. Carbon dioxide protects the

- perinatal brain from hypoxic-ischemic damage: an experimental study in the immature rat. *Pediatrics* 1995;95:868-74.
24. Garcia JH, Wagner S, Liu KF, Hu XJ. Neurological deficit and extent of neuronal necrosis attributable to middle cerebral artery occlusion in rats. Statistical validation. *Stroke* 1995;26:627-34; discussion 35.
 25. Longa EZ, Weinstein PR, Carlson S, Cummins R. Reversible middle cerebral artery occlusion without craniectomy in rats. *Stroke* 1989;20:84-91.
 26. Chen J, Sanberg PR, Li Y, Wang L, Lu M, Willing AE, et al. Intravenous administration of human umbilical cord blood reduces behavioral deficits after stroke in rats. *Stroke* 2001;32:2682-8.
 27. Hamm RJ, Pike BR, O'Dell DM, Lyeth BG, Jenkins LW. The rotarod test: an evaluation of its effectiveness in assessing motor deficits following traumatic brain injury. *J Neurotrauma* 1994;11:187-96.
 28. An Experimental Model of Closed Head Injury in Mice: Pathophysiology, Histopathology, and Cognitive Deficits. *Journal of Neurotrauma* 1996;13:557-68.
 29. Zhou Q, Cao B, Niu L, Cui X, Yu H, Liu J, et al. Effects of permissive hypercapnia on transient global cerebral ischemia-reperfusion injury in rats. *Anesthesiology* 2010;112:288-97.
 30. Zhang CH, Fan YY, Wang XF, Xiong JY, Tang YY, Gao JQ, et al. Acidic preconditioning protects against ischemia-induced brain injury. *Neurosci Lett* 2012;523:3-8.
 31. Shen Z, Zheng Y, Wu J, Chen Y, Wu X, Zhou Y, et al. PARK2-dependent mitophagy induced by acidic postconditioning protects against focal cerebral ischemia and extends the reperfusion window. *Autophagy* 2017;13:473-85.
 32. Ainslie PN, Duffin J. Integration of cerebrovascular CO₂ reactivity and chemoreflex control of breathing: mechanisms of regulation, measurement, and interpretation. *Am J Physiol Regul Integr Comp Physiol* 2009;296:R1473-95.
 33. Friis ML, Paulson OB, Hertz MM. Carbon dioxide permeability of the blood-brain barrier in man. The effect of acetazolamide. *Microvasc Res* 1980;20:71-80.

34. Edwards G, Félétou M, Weston AH. Endothelium-derived hyperpolarising factors and associated pathways: a synopsis. *Pflugers Arch* 2010;459:863-79.
35. Sandow SL, Haddock RE, Hill CE, Chadha PS, Kerr PM, Welsh DG, et al. What's where and why at a vascular myoendothelial microdomain signalling complex. *Clin Exp Pharmacol Physiol* 2009;36:67-76.
36. Yoon S, Zuccarello M, Rapoport RM. pCO(2) and pH regulation of cerebral blood flow. *Front Physiol* 2012;3:365.
37. Brugniaux JV, Hodges AN, Hanly PJ, Poulin MJ. Cerebrovascular responses to altitude. *Respir Physiol Neurobiol* 2007;158:212-23.
38. Cohen MV, Yang XM, Downey JM. The pH hypothesis of postconditioning: staccato reperfusion reintroduces oxygen and perpetuates myocardial acidosis. *Circulation* 2007;115:1895-903.
39. Eastwood G, Nichol AD, Hodgson C, Parke RL, McGuinness S, Nielsen N, et al. Mild Hypercapnia or Normocapnia after Out-of-Hospital Cardiac Arrest. *N Engl J Med* 2023;389:45-57.
40. Eastwood GM, Schneider AG, Suzuki S, Peck L, Young H, Tanaka A, et al. Targeted therapeutic mild hypercapnia after cardiac arrest: A phase II multi-centre randomised controlled trial (the CCC trial). *Resuscitation* 2016;104:83-90.
41. Yamato M, Egashira T, Utsumi H. Application of in vivo ESR spectroscopy to measurement of cerebrovascular ROS generation in stroke. *Free Radic Biol Med* 2003;35:1619-31.
42. Manzanero S, Santro T, Arumugam TV. Neuronal oxidative stress in acute ischemic stroke: sources and contribution to cell injury. *Neurochem Int* 2013;62:712-8.
43. Vesela A, Wilhelm J. The role of carbon dioxide in free radical reactions of the organism. *Physiol Res* 2002;51:335-9.
44. Yang Y, Estrada EY, Thompson JF, Liu W, Rosenberg GA. Matrix metalloproteinase-mediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *J Cereb Blood Flow Metab* 2007;27:697-709.

45. Bauvois B. New facets of matrix metalloproteinases MMP-2 and MMP-9 as cell surface transducers: outside-in signaling and relationship to tumor progression. *Biochim Biophys Acta* 2012;1825:29-36.
46. Heo JH, Kim SH, Lee KY, Kim EH, Chu CK, Nam JM. Increase in plasma matrix metalloproteinase-9 in acute stroke patients with thrombolysis failure. *Stroke* 2003;34:e48-50.
47. Ramos-Fernandez M, Bellolio MF, Stead LG. Matrix metalloproteinase-9 as a marker for acute ischemic stroke: a systematic review. *J Stroke Cerebrovasc Dis* 2011;20:47-54.
48. Armulik A, Genove G, Betsholtz C. Pericytes: developmental, physiological, and pathological perspectives, problems, and promises. *Dev Cell* 2011;21:193-215.
49. Armulik A, Genove G, Mae M, Nisancioglu MH, Wallgard E, Niaudet C, et al. Pericytes regulate the blood-brain barrier. *Nature* 2010;468:557-61.
50. Sweeney MD, Ayyadurai S, Zlokovic BV. Pericytes of the neurovascular unit: key functions and signaling pathways. *Nat Neurosci* 2016;19:771-83.
51. Yemisci M, Gursay-Ozdemir Y, Vural A, Can A, Topalkara K, Dalkara T. Pericyte contraction induced by oxidative-nitrative stress impairs capillary reflow despite successful opening of an occluded cerebral artery. *Nat Med* 2009;15:1031-7.
52. Hall CN, Reynell C, Gesslein B, Hamilton NB, Mishra A, Sutherland BA, et al. Capillary pericytes regulate cerebral blood flow in health and disease. *Nature* 2014;508:55-60.
53. Wenger NK, Speroff L, Packard B. Cardiovascular health and disease in women. *N Engl J Med* 1993;329:247-56.
54. Yang W, Zhang X, Wang N, Tan J, Fang X, Wang Q, et al. Effects of Acute Systemic Hypoxia and Hypercapnia on Brain Damage in a Rat Model of Hypoxia-Ischemia. *PLoS One* 2016;11:e0167359.
55. Duong TQ, Iadecola C, Kim SG. Effect of hyperoxia, hypercapnia, and hypoxia on cerebral interstitial oxygen tension and cerebral blood flow. *Magn Reson Med* 2001;45:61-70.

Abstract in Korean

급성 뇌경색 동물 모델에서 재관류 치료에 동반한 이산화탄소 흡입 치료의 효과

연구 배경: 중대뇌동맥 폐쇄 (MCAO) 설치류 모델에서 재관류 전 이산화탄소 흡입이 산화 스트레스, 뇌혈관장벽의 손상, 신경학적 손상을 감소시키는지를 조사하였다.

방법: Sham, Sham + CO₂, 일시적 중대뇌동맥 폐쇄(tMCAO), 영구적 중대뇌동맥 폐쇄(pMCAO), 그리고 tMCAO + CO₂군으로 나뉘서 실험을 진행하였다. , Sham + CO₂, tMCAO + CO₂군은 혼합 가스(20% CO₂, 20% O₂, 60% N₂)를 흡입하였다. MCAO 수술 후 각 실험군마다 동맥혈가스분석 및 행동 검사를 시행하였으며 24 시간 이후 뇌경색 크기 확인을 위한 조직 염색, 웨스턴 블롯, 면역형광염색을 시행하였다.

결과: 이산화탄소 흡입 치료를 한 tMCAO + CO₂군은 tMCAO군에 비해 뇌경색 크기가 작았고(tMCAO+CO₂군, 14.8 [11.0, 17.8] vs. tMCAO군, 27.9 [26.4, 28.0], $p < 0.0001$) 신경학적 점수도 tMCAO + CO₂군이 tMCAO군 또는 pMCAO군 보다 더 우수하였다. tMCAO + CO₂군에서 tMCAO군 또는 pMCAO군에 비해 8-OHdG와 MMP-9 의 발현이 유의미하게 감소하였다. tMCAO + CO₂군에서 tMCAO군 또는 pMCAO군에 비해 ZO-1과 PDGFR β 의 발현은 유의미하게 증가하였다.

결론: 경도의 고탄산혈증은 허혈 재관류 손상으로부터 허혈성 반응을 보호하며,

신경학적 결손을 줄일 뿐 아니라, 뇌혈관장벽과 신경혈관단위의 보전 및 산화 스트레스 줄인다. 치료적 고탄산혈증은 유망하며 임상 적용을 위한 추가 평가가 필요하다.

핵심되는 말: 이산화탄소, 허혈성 뇌졸중, 뇌경색 동물 모델, 산화 스트레스, 허혈 재관류 손상