

Genome-wide association and replication studies for handedness in a Korean community-based cohort

Youhyun Song^{1,2} | Dasom Lee³ | Ja-Eun Choi³ | Ji Won Lee^{4,5}  | Kyung-Won Hong³

¹Department of Family Medicine, Gangnam Severance Hospital, Yonsei University College of Medicine, Seoul, South Korea

²Healthcare Research Team, Health Promotion Center, Gangnam Severance Hospital, Yonsei University College of Medicine, Seoul, South Korea

³Theragen Bio Co. Ltd., Gyeonggi-do, South Korea

⁴Department of Family Medicine, Severance Hospital, Yonsei University College of Medicine, Seoul, South Korea

⁵Institute for Innovation in Digital Healthcare, Yonsei University, Seoul, South Korea

Correspondence

Ji Won Lee, Department of Family Medicine, Severance Hospital, Yonsei University College of Medicine, 50-1 Yonsei-ro, Seodaemun-gu, Seoul 03722, South Korea.
Email: indi5645@yuhs.ac

Kyung-Won Hong, Healthcare R&D Division, Theragen Bio Co. Ltd., A-10F, Samhwan HIPEX, 240, Pangyoek-ro, Bundang-gu, Seongnam-si, Gyeonggi-do, South Korea.
Email: kyungwon.hong@theragenbio.com

Youhyun Song and Dasom Lee are co-first authors.

Funding information

Institute for Information & Communications Technology Promotion (IITP) grant funded by the Korea government (Ministry of Science and ICT), Grant/Award Number: 2019-31-1293; Ministry of Trade, Industry and Energy, Grant/Award Number: 20002781; Korea Disease Control and Prevention Agency, Grant/Award Number: 4800-4848-501

Abstract

Introduction: Handedness is a conspicuous characteristic in human behavior, with a worldwide proportion of approximately 90% of people preferring to use the right hand for many tasks. In the Korean population, the proportion of left-handedness is relatively low at approximately 7%–10%, similar to that in other East-Asian cultures in which the use of the left hand for writing and other public activities has historically been oppressed.

Methods: In this study, we conducted two genome-wide association studies (GWASs) between right-handedness and left-handedness, and between right-handedness and ambidexterity using logistic regression analyses using a Korean community-based cohort. We also performed association analyses with previously reported variants and our findings.

Results: A total of 8806 participants were included for analysis, and the results identified 28 left-handedness-associated and 15 ambidexterity-associated loci; of these, two left-handedness loci (*NEIL3* [rs11726465] and *SVOPL* [rs117495448]) and one ambidexterity locus (*PDE8B/WDR41* [rs118077080]) showed near genome-wide significance. Association analyses with previously reported variants replicated *ANKS1B* (rs7132513) in left-handedness and *ANKIB1* (rs2040498) in ambidexterity.

Conclusion: The variants and positional candidate genes identified and replicated in this study were largely associated with brain development, cerebral asymmetry, neurological processes, and neuropsychiatric diseases in line with previous findings. As the first East-Asian GWAS related to handedness, these results may provide an intriguing reference for further human neurologic research in the future.

KEY WORDS

ambidexterity, genome-wide association study, genetic, handedness, left-handedness

1 | INTRODUCTION

Handedness is a prominent variation in human behavior. There are various hypotheses based on extensive studies regarding the development of handedness, and these remain highly controversial. However, many studies regard neurological or brain asymmetry as the main cause (Kang et al., 2017; Papadatou-Pastou et al., 2020; Paracchini, 2021). A recent genome-wide association study (GWAS) meta-analysis regarding handedness based on Caucasians, the largest to date, reported 41 genes significant for left-handedness and seven genes significant for ambidexterity (Cuellar-Partida et al., 2021). According to this study, the list of genome-wide significant associations includes multiple variants associated with genes involved in microtubules, brain morphology, and neuropsychiatric traits.

Globally, the proportion of left-handed people is approximately 10%–12%; in Korean adults, the proportion of left-handedness is 7%–10% (Hardyck & Petrinovich, 1977). The probability of being left-handed may be affected by the time of birth or geographical region and may have a large cultural influence, along with genetic pathways. In the past, writing or performing public acts using one's left hand has been strictly forbidden in several East-Asian cultures, often accompanying forced "correction" to right-handedness (Zheng et al., 2020). Accordingly, based on a 2013 "Gallup Korea" survey, the proportion of left-handed Koreans in their 20s was the highest at 8%, and that among those in their 60s was the lowest at 2% (Gallup (Korea), 2013). Under such circumstances, genetic factors can be considered to play a strong role in growing up to be left-handed despite cultural restraints in the elderly Korean population.

To the best of our knowledge, there have been no GWAS analyses on handedness that are focused on East-Asians. In this study, we aimed to identify the genetic variants associated with handedness in an East-Asian population-based cohort. We also attempted to examine previously reported genetic variants regarding handedness using our results from Korean data.

2 | MATERIALS AND METHODS

2.1 | Study population

This study used data from the KoGES—Ansan and Ansung conducted by the Korea Centers for Disease Control and Prevention. The KoGES aims to investigate the genetic and environmental etiology of non-communicable chronic diseases such as type 2 diabetes, obesity, and cardiovascular diseases. In total, 8806 participants between the ages of 40 and 69 years were recruited from 2001 to 2002 through two population-based prospective cohort studies conducted in the Ansung ($n = 4178$) and Ansan ($n = 4628$) regions of South Korea. After excluding participants with missing values ($n = 34$), the number of participants included in this study totaled 8806. Detailed information about the KoGES has been described in a previous report (Kim & Han, 2017), including identity-by-state, multidimensional scaling,

and principal component analyses (Cho et al., 2009). This research project was approved by the Institutional Review Board of Theragen Etex (approval numbers: 700062-20190819-GP-006-02). This study complied with the ethical principles of the Declaration of Helsinki. Baseline and follow-up recruitment and assessment were conducted after obtaining written informed consent.

2.2 | Data collection

The study participants were interviewed by trained medical staff regarding their lifestyle, sociodemographic status, disease history, and other factors. Handedness was classified into three categories by questionnaire (right-handedness, left-handedness, and ambidexterity), and trained staff completed the questionnaire by a direct interview. Anthropometric and clinical measurements were also obtained through standard protocol to evaluate health status, as detailed previously (Kim & Han, 2017).

2.3 | Genotypes

Genotype data were provided by the Center for Genome Science, Korea National Institute of Health. The genotype data were generated by the Korea Biobank Array (Affymetrix, Santa Clara, CA, USA) (Moon et al., 2019). The experimental results were filtered using the following quality control criteria: call rate $>97\%$, minor allele frequency $>1\%$, and Hardy-Weinberg equilibrium test p -value $< 1 \times 10^{-5}$. After the quality control procedures, experimentally determined genotypes were used as the genotype imputation dataset against the 1000 Genome Phase 1 and 2 Asian panels. Finally, the GWAS included 7,975,321 single nucleotide polymorphisms (SNPs) from chromosomes 1 to 22.

2.4 | Statistical analyses

The proportions of handedness were compared using chi-squared tests for the three handedness categories (right-handedness, left-handedness, and ambidexterity). Two GWASs examined the likelihood of left-handedness or ambidexterity rather than right-handedness. The GWASs were performed via logistic regression, adjusting for age, sex, and residence area as covariates, using PLINK version 1.09. The genomic inflation factor lambda was 1.00 for both GWASs (see the quantile-quantile plot in Figure S1). Odds ratios (ORs) and 95% confidence intervals (95% CIs) were calculated. The threshold for significant associations was defined as p -value $< 5.00 \times 10^{-8}$. The top significant SNPs obtained through these analyses were classified into loci based on whether they satisfied the criteria of $D' > 0.8$ and $R^2 > 0.8$; the LD link tool (<https://ldlink.nci.nih.gov/?tab=home>) was used to confirm the linkage disequilibrium with the cluster SNPs at a range of ± 10 kb.

TABLE 1 Baseline demographic of the study population.

Variables	Total	Handedness groups			
		Right-handedness	Left-handedness	Ambidexterity	p-Value
N (%)	8806	7722 (87.69)	340 (3.86)	744 (8.45)	
Age (mean \pm SD)	52.2 \pm 8.9	52.3 \pm 9.0	51.9 \pm 8.8	51.3 \pm 8.5	9.15E-03
Sex (male %)	4167 (47.3)	3616 (46.8)	179 (52.6)	372 (50.0)	3.39E-02
Residence area (Ansung/Ansan %)	47.4/52.6	46.8/53.2	56.8/43.2	49.7/50.3	6.61E-04

2.5 | Replication test

To understand the similarity of the handedness-related genetic backgrounds between Korean and European populations, we also examined previously reported handedness-related SNPs in our dataset.

3 | RESULTS

3.1 | Population characteristics

Handedness data were obtained from self-reported questionnaires. Among 8806 individuals aged 40–69 years (mean age, 52.2 \pm 8.9 years), 4167 (47.3%) were male. Further, 7722, 340, and 744 identified themselves as right-handed, left-handed, and ambidextrous, respectively. Females showed a higher tendency for right-handedness than males (53.2% females vs. 46.8% males), whereas males showed more left-handedness than females (52.6% males vs. 47.4% females). No significant difference between sexes was found with ambidexterity (50.0%) (Table 1).

3.2 | Replication results

Cuellar-Partida et al. identified 41 left-handedness loci and seven ambidexterity loci in people of European ancestry. They also found a low genetic correlation between the two traits ($R_g = 0.26$). We investigated the associations of the 48 handedness loci identified by Cuellar-Partida et al. and the 43 loci identified in the Korean population in this study (p -value $< .05$). ANKS1B (rs7132513, OR = 0.67, 95% C.I. = 0.45–0.99, p -value = .04831) in left-handedness and ANKIB1 (rs2040498, OR = 0.77, 95% C.I. = 0.64–0.93, p -value = .006035) in ambidexterity showed associative significance. The reported association between ANKIB1 and ambidexterity was greater than 1, but Cuellar-Partida et al. expressed this based on the effective allele rather than the minor allele. In this study, our result was smaller than 1 based on the minor allele, which can be considered an equally reproducible result (Table 2, see Table S3 for the original GWAS replication results). In addition, we performed a meta-analysis with the results from the 23andMe dataset included in Cuellar-Partida et al.'s study.

The two replicated variants showed increased statistical significance in the meta-analysis (see Table S4).

3.3 | GWAS findings

We identified 28 and 15 loci related to left-handedness and ambidexterity, respectively, which showed genome-wide suggestive significance (p -value $< 10^{-5}$); the relevant GWAS results are presented in Table 3 and shown as Manhattan plots in Figure 1. There were no common loci between the two traits. From the 28 loci related to left-handedness, 20 candidate genes were identified by proximity (NEIL3 [rs11726465], SVOPHL [rs117495448], SGIP1 [rs78509271], KLB [rs28573932], CLYBL [rs77395534], NTRK3 [rs56069748], RIPK4 [rs3746895], CX3CR1 [rs938207], MYRFL [rs192721285], CACNA1E [rs3766982], ARHGEF7 [rs35337719], TRIB2 [rs142891714], CDH18 [rs770303], TAFA1 [rs907082], TMEM132D [rs1451902], RNFT2 [rs140649861], PELI2 [rs76895919], PLA2G4A [rs77239173], CIT [rs74436390], and INTS6 [rs142002811]), and from 15 loci related to ambidexterity, 13 candidate genes were identified (PDE8B/WDR41 [rs118077080], IGSF11 [rs138907675], SYK [rs182637278], ST8SIA4 [rs79041957], WDR41 [rs145242715], GATA3 [rs263422], GLMN [rs150150088], SNAI2 [rs80114285], CTNNND2 [rs4476693], ROBO1 [rs75578608], PTPRD [rs7862596], ST8SIA3 [rs12606549], and TRNT1 [rs186743]) (the original GWAS results in their entirety are shown in Table S1 [left-handedness] and Table S2 [ambidexterity].)

Among the above, rs11726465 (NEIL3, OR = 1.7, 95% C.I. = 1.4–2.0, p -value = 6×10^{-8}) and rs117495448 (SVOPHL, OR = 2.2, 95% C.I. = 1.7–3.0, p -value = 9×10^{-8}) for left-handedness, and rs118077080 (PDE8B/WDR41, OR = 2.1, 95% C.I. = 1.6–2.7, p -value = 6×10^{-8}) for ambidexterity nearly met the threshold for genome-wide significance (p -value $< 5 \times 10^{-8}$). The respective signal plots are shown in Figure 2.

4 | DISCUSSION

To the best of our knowledge, this study represents the first East-Asian GWAS for handedness to date. We used data from KoGES, a database representative of the Korean population to perform a GWAS analysis, and identified significant genetic loci associated with

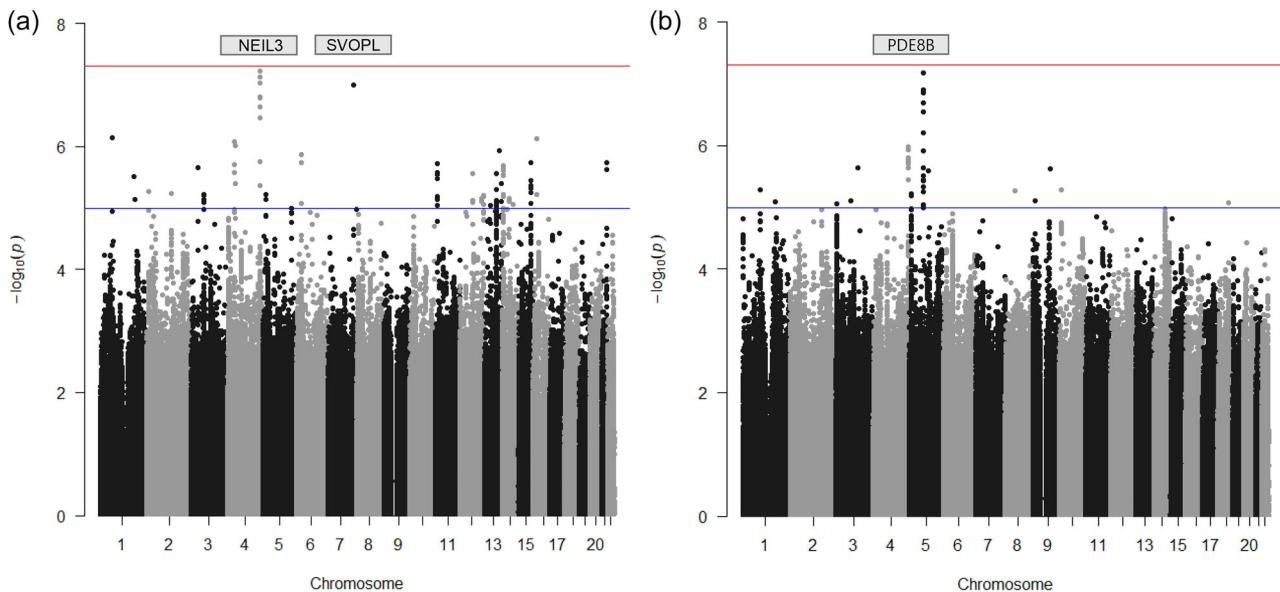


FIGURE 1 Manhattan plots showing the genome-wide association study (GWAS) results. (a) Left-handedness and (b) ambidexterity. Genome-wide significant p -value criteria (p -value $< 5 \times 10^{-8}$) are shown in red, and genome-wide suggestive p -value criteria ($5 \times 10^{-8} \leq p\text{-value} < 1 \times 10^{-5}$) are shown in blue. The y-axis is $-\log_{10}$ (p -value) of the single nucleotide polymorphisms (SNPs), and the x-axis is chromosomal position (hg19). Summary statistics for the lead variants at genome-wide significant loci are presented in Table 3 along with the gene nearest to the lead SNP. Candidate genes were identified as the gene nearest to the lead SNPs with nearby variant clusters showing similar significance.

left-handedness and ambidexterity; we further identified their candidate genes and hypothesized potential genetic correlations between handedness and neuropsychiatric development or traits. We also conducted association analyses with the results of the largest genetic study to date by Cuellar-Partida et al. to identify common loci associated with handedness between Caucasians and East-Asians.

Several studies performed worldwide have attempted to identify how handedness develops biologically and culturally; however, the underlying mechanism remains unclear (de Kovel et al., 2019; de Kovel & Francks, 2019; Michel, 2021; Papadatou-Pastou et al., 2020). Moreover, Asian populations have been relatively under-represented in these studies (Zheng et al., 2020). Twin and familial studies have shown that approximately 25% of the variance in handedness can be attributed to genetics (de Kovel et al., 2019; Zheng et al., 2020). However, only a fraction of this percentage can be attributed to specific genes, and relevant large-scale studies are ongoing to uncover more of these. Genetic correlations identified to date implicate high polygenicity and shared biology between handedness, cerebral asymmetries, neurodevelopment/plasticity, neurodegenerative processes, and neuropsychiatric diseases (Cuellar-Partida et al., 2021; Kong et al., 2021; Paracchini, 2021). In addition, left-handedness and ambidexterity have been suggested to be influenced by different genetic mechanisms (Cuellar-Partida et al., 2021).

Among the 28 left-handedness loci, two (NEIL3 and SVOPL) showed near genome-wide significance. NEIL3, an important gene related to DNA repair, has been widely implicated in diverse brain functions (Hildrestrand et al., 2009; Kunath et al., 2021; Regnell et al., 2012). Recently, NEIL3 has been identified to enable stable neural representation of space by shaping CA1 transcription in mouse hippocampal

neurons, with $\text{NEIL3}^{-/-}$ mice showing impaired spatial performance (Kunath et al., 2021). Other studies have also shown an important role of NEIL3 in fetal neurogenesis (Hildrestrand et al., 2009), as well as maintenance of adult neurogenesis in the hippocampus, thus implicating associations with neurodegenerative disease (Regnell et al., 2012). Further, composite substance dependence phenotypes (Wetherill et al., 2014) and impulsivity (Ehlers et al., 2016) associations have been reported. These findings are interesting because handedness (as well as temporal lobe epilepsy) has been identified to affect right-to-left amygdalar and hippocampal volume ratios (Szabo et al., 2001).

SVOPL is a paralog of the SVOP gene that encodes synaptic vesicle 2 (SV2)-related proteins expressed in neurons, and is expressed in all brain regions while appearing earlier than SV2 in development (Yao et al., 2013). SVOPL has been found to be significant in Parkinson's disease by large-scale whole-exome sequencing (Jansen et al., 2017), potentially deleterious in autosomal-dominant lateral temporal epilepsy (Dazzo et al., 2015), and is associated with a super-variant associated with brain connectivity (Li et al., 2021). Further, it has been found to be associated as a non-word repetition marker in specific language impairment risk (Luciano et al., 2013). Although accumulating evidence suggests a genetic relationship in variants associated with handedness (especially left-handedness), cerebral asymmetry, and neuropsychiatric/developmental disorders such as dyslexia, Parkinson's disease, and schizophrenia, these remain to be clearly identified (Brandler & Paracchini, 2014; Wiberg et al., 2019).

Of the 15 ambidexterity loci, PDE8B showed the highest, near genome-wide significance. PDE8B plays a major role in the breakdown of cyclic AMP, a key messenger in dopamine signaling (Bollen

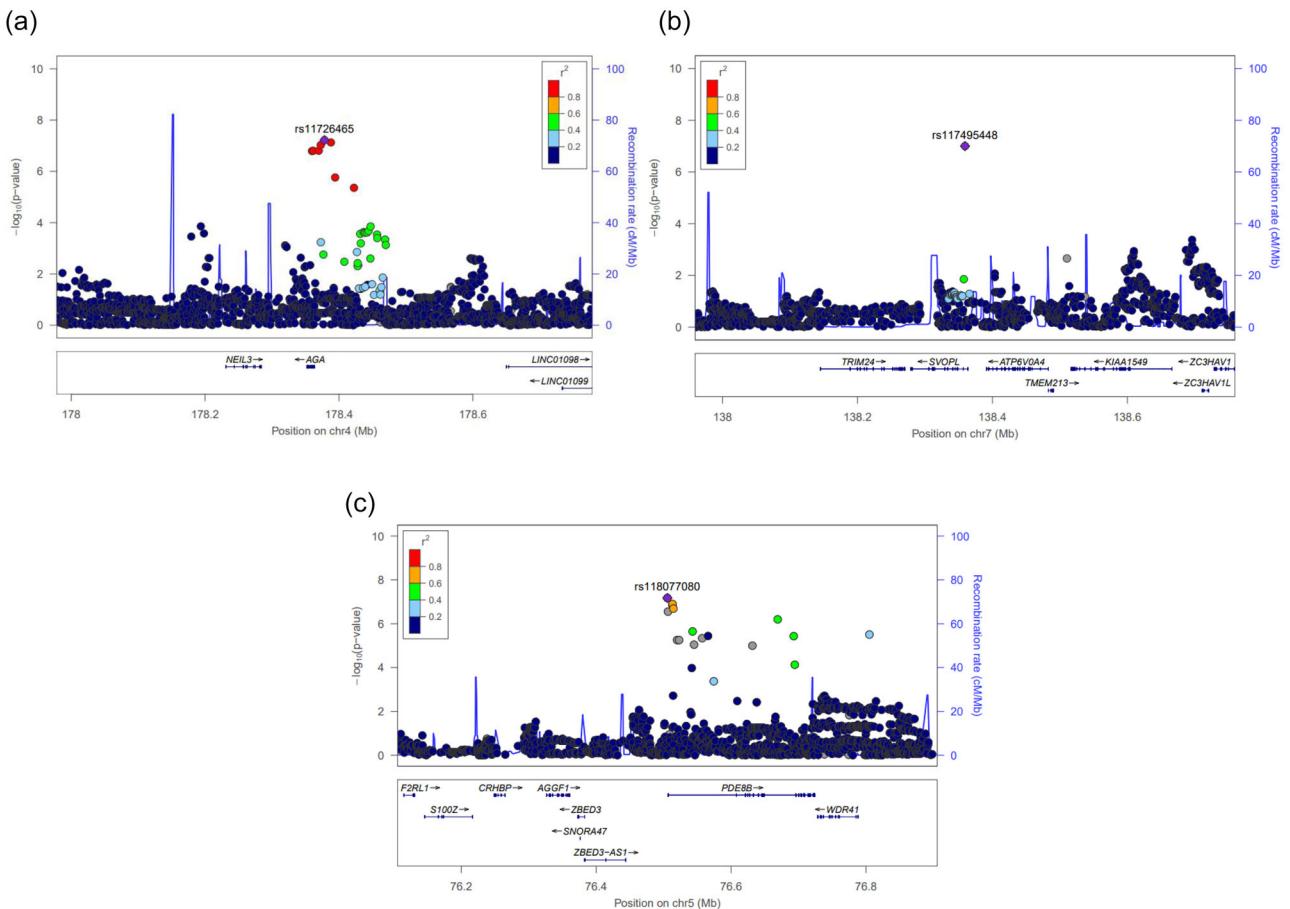


FIGURE 2 Regional association plots for lead single nucleotide polymorphisms (SNPs). (a/b) Left-handedness and (c) ambidexterity. SNP is plotted by chromosomal position (hg19; x-axis) and association with handedness from the current study ($-\log_{10}$ (p-value); y-axis).

& Prickaerts, 2012). *PDE8B* expression analysis in humans has shown the highest levels in the striatum, hippocampal formation, cerebellum, and cortex (Pérez-Torres et al., 2003); hippocampus-dependent brain functions such as spatial pattern recognition and contextual fear are known to be regulated by cAMP signaling; further, striatal regulation of cAMP downstream from dopamine pathways plays a major role in numerous aspects of motor function as well as motivated behavior and reward learning. Accordingly, *PDE8B* has been implicated in autosomal-dominant striatal degeneration (Appenzeller et al., 2010), with higher levels observed in Alzheimer's patients (Pérez-Torres et al., 2003); in agreement with these findings, *PDE8B* KO mice demonstrate enhanced motor performance as well as alleviated motor coordination decay (Tsai et al., 2012). Along with *PDE8B*, the adjacently located *WDR41* is associated with dopamine signaling and development as well as autophagy in neurons; it is also suspected to be involved in ALS/FTLD (Budini et al., 2017; Goodier et al., 2020; Sullivan et al., 2016). These two genes were identified to have peak associations in a study on caudate volume, which suggested relevance to common conditions affecting the caudate (Stein et al., 2011). We consider this finding as significant because the basal ganglia including the caudate is a brain structure involved in many common neurological and

psychiatric diseases as it regulates the dopaminergic system; cerebral asymmetries involving this region have been previously found to have associations with such disorders as well as handedness (Jang et al., 2017; Peterson et al., 1993). Additionally, functional nigrostriatal dopamine system asymmetry has been identified in motor lateralization using fluorodopa-PET (de la Fuente-Fernandez et al., 2000).

Other genes near the variant also showed involvement in brain development (*ZBED3*) (Ruan et al., 2021), in GABAergic interneurons in the prefrontal cortex (Ketcheson et al., 2017), hippocampal abnormalities (Ensink et al., 2021), depression, and other neuropsychiatric disorders such as substance abuse (*CRHBP*) (Curley et al., 2021; Kalin, 2018).

Among the 28 suggestively significant left-handedness variants, we could identify 20 genes proximal to the loci; of these, 13 are related to neurological pathways such as synapses, neurons, or neuronal signaling (*AGA* (Chen et al., 2021; Saarela et al., 2004), *SVOPL* (Yao et al., 2013), *SGIP1* (Hajkova et al., 2016), *KLB* (Jackson et al., 2019), *NTRK3* (Ito et al., 2020), *CX3CR1* (Pawelec et al., 2020), *CACNA1E* (Carvill, 2019), *ARHGEF7* (Lopez Tobon et al., 2018), *TRIB2* (Dobens et al., 2021), *CDH18* (Bai et al., 2018), *TAFA1* (Sarver et al., 2021), *PELI2*

TABLE 2 Significant single nucleotide polymorphisms (SNPs) in the replication genome-wide association study (GWAS).

rs ID	Chromosome	Base pair	Proximal gene	Minor allele frequency	Major allele	Logistic regression analysis			HWE	
						Odds ratio	95% Confidence interval	p-Values	Sample size ^a	p-Values
Left-handedness										
rs732513	12	100324975	ANKS1B	0.06	G	C	0.67	0.45–0.99	4.83E-02	1107
Ambidexterity										8.40E-01
rs2040498	7	91899117	ANKIB1	0.11	T	A	0.77	0.64–0.93	6.04E-03	1282
Abbreviations: HWE, Hardy-Weinberg equilibrium.										

^aSample size for 80% power at $\alpha = .05$ is based on the parameter of this population, including minor allele frequency and effect size of the markers.

(Dai et al., 2019), and *CIT* (Ahmed et al., 2011)). Further, four genes are correlated with neuropsychiatric traits such as behavioral disorders, panic disorders, or Alzheimer's disease (*MYRFL* (Anney et al., 2008), *TNEN132* (Naik et al., 2018), *RNFT2* (Kamboh et al., 2012), and *PLA2G4A* (Sarkar et al., 2020)). Similarly, among the 15 suggestively significant ambidexterity-associated variants, we located 13 proximal genes. Of these, eight genes are related to cell signaling or neurons (*PDE8B* (Pearse et al., 2004; Fan et al., 2018), *ST8SIA4* (Berger et al., 2016), *WDR41* (Goodier et al., 2020), *GATA3* (Andrzejczuk et al., 2018), *SNAI2* (Wei et al., 2020), *CTNND2* (Turner et al., 2015), *ROBO1* (G. Wang et al., 2013), and *PTPRD* (Tomita et al., 2020)), and three are associated with brain development or neurodevelopmental, degenerative disorders (*IGSF11* (Higashine et al., 2018), *SYK* (Angibaud et al., 2011; Nazarian et al., 2019), and *TRNT1* (Chakraborty et al., 2014)).

We also performed association analyses with the findings of Cuellar-Partida et al. that are based on Caucasians and could replicate two variants in the Korean population. In left-handedness and ambidexterity, rs7132513 and rs2040498, respectively, showed common significance between the two study populations (p -value $\leq .05$). *ANKS1B* (rs7132513) encodes a protein predominantly expressed in the brain and testis, which interacts with amyloid beta protein precursor, and may play a role in normal neurodevelopment as well as the pathogenesis of Alzheimer's disease (Carbonell et al., 2019). *ANKIB1* (rs2040498) is a protein-coding gene predicted to be involved in ubiquitin-associated activity and has been identified in angiokeratoma corporis diffusum with arteriovenous fistulas and autism spectrum disorder (L.-S. Wang et al., 2010). Further, an important gene is *RNF19A*, which encodes the protein E3 ubiquitin ligase localized to Lewy bodies, which may be involved in ALS and Parkinson's disease (Park et al., 2015).

This study has several merits. To the best of our knowledge, it is the first GWAS on East-Asians to date, and the cohort used for analysis is a well-known large database representative of the Korean population. Two common variants were replicated from the largest handedness GWAS to date in a different ethnicity; both variants were correlated with genes involved in neurological processes. Further, owing to the unique East-Asian culture of disregarding left-handedness and strongly enforcing right-handedness, the identified left-handedness associated variants may have stronger predictive power than others. Our database included a large middle-aged population between 40–60 years of age, which was raised in a society in which left-handedness was taboo (only 2% of Koreans in their 60s identified as left-handed). The participants who maintained left-handedness despite environmental coercion may have been largely affected by the genetic variants identified in this study. The variants and candidate genes identified in this study potentially implicate novel associations or reinforce known relations between handedness and neurologic processes and conditions. However, it is not clear whether these relations between handedness and neuropsychiatric traits are directly, independently associated based on our analysis; further studies are warranted to explore the exact nature of these associations based on our findings.

However, this study has limitations. Although some variants nearly met the threshold for genome-wide significance (p -value $< 5 \times 10^{-8}$),

TABLE 3 Single nucleotide polymorphisms (SNPs) associated with handedness by genome-wide association study (GWAS).

rs ID	Chr	Base pair	Proximal gene	Minor allele frequency	Major allele	Logistic regression analysis	HWE		
							Odds ratio	95% Confidence interval	p-Values
Left-handedness									
rs7509271	1	67194629	SGIP1	0.06	T	C	2.00	1.50-2.54	7.19E-07
rs366982	1	181646981	CACNA1E	0.02	G	A	2.70	1.78-4.08	3.05E-06
rs77239173	1	186736169	PLA2G4A	0.02	T	C	2.30	1.60-3.33	7.39E-06
rs142891714	2	12739030	TRIB2	0.01	T	C	2.80	1.81-4.44	5.32E-06
rs113690115	2	137242045	2q22.1	0.04	C	T	2.00	1.50-2.79	5.90E-06
rs98207	3	39296828	CX3CR1	0.08	A	G	1.70	1.38-2.19	2.21E-06
rs907082	3	68523156	TAF41	0.08	A	G	1.80	1.37-2.23	6.07E-06
rs28573932	4	39445596	KLB	0.01	T	G	3.30	2.05-5.27	8.49E-07
rs141918675	4	43169480	4p13	0.01	T	A	3.00	1.94-4.69	9.74E-07
rs1726465	4	178378317	NEIL3	0.14	A	C	1.70	1.39-2.03	6.03E-08
rs770303	5	19473917	CDH18	0.25	A	G	1.50	1.25-1.74	5.93E-06
rs111677893	6	31347069		0.35	T	C	0.70	0.55-0.78	1.37E-06
rs117495448	7	138359303	SVOPL	0.04	A	G	2.20	1.67-3.02	2.98E-08
rs113628813	11	11726915	11p15.4-15.3	0.01	T	G	3.00	1.92-4.77	1.88E-06
rs192721285	12	70339563	MYRF1	0.02	A	C	2.40	1.65-3.41	2.74E-06
rs140649861	12	117284214	RNFT2	0.04	A	G	2.00	1.50-2.78	6.94E-06
rs74436390	12	120184134	C/T	0.01	A	T	3.00	1.85-4.86	7.86E-06
rs1451902	12	130248986	TMEV132D	0.39	C	T	1.40	1.22-1.66	6.17E-06
rs142002811	13	51963277	INTS6	0.01	C	T	3.00	1.84-4.79	8.94E-06
rs61200745	13	85238840	13q31.1	0.13	C	A	1.60	1.32-1.97	2.72E-06
rs77395534	13	100458813	CLYBL	0.04	G	A	2.20	1.58-2.93	1.16E-06

(Continues)

TABLE 3 (Continued)

rs ID	Chr	Base pair	Proximal gene	Minor allele frequency	Major allele	Logistic regression analysis			Sample size ^a	<i>p</i> -Values	
						Odds ratio	95% Confidence interval	<i>p</i> -Values			
rs35337719	13	111772219	ARHGEF7	0.06	T	C	1.80	1.42-2.38	4.01E-06	330	6.43E-01
rs7160724	14	28219361	14p12	0.48	C	T	0.70	0.59-0.81	6.00E-06	252	6.40E-01
rs73895919	14	56800208	PEL12	0.01	T	C	2.90	1.81-4.52	6.95E-06	383	4.08E-01
rs148414906	14	82337126	14q31.1	0.02	A	T	2.30	1.60-3.34	8.64E-06	407	6.92E-02
rs56069748	15	88373203	NTRK3	0.02	A	T	2.60	1.74-3.75	1.81E-06	333	7.75E-01
rs140084235	16	18024168	16p12.3	0.01	T	A	3.10	1.99-4.90	7.38E-07	356	6.43E-01
rs3746895	21	43162200	RIPK4	0.33	T	C	0.60	0.54-0.77	1.85E-06	151	3.12E-02
Ambidexterity											
rs150150088	1	92758007	GLMN	0.01	A	G	2.30	1.59-3.21	5.21E-06	632	4.11E-01
1:173302463:GA_G	1	173302463		0.06	R	D	1.60	1.28-1.88	7.98E-06	526	1.00E+00
rs186743	3	3178322	TRNT1	0.28	A	G	1.30	1.15-1.45	8.76E-06	540	2.70E-01
rs75578608	3	79347280	ROBO1	0.03	C	G	1.80	1.38-2.28	7.85E-06	608	1.00E+00
rs138907675	3	118725079	IGSF11	0.07	G	A	0.50	0.41-0.69	2.31E-06	317	1.16E-02
4:188281625:G_GGG	4	188281625		0.34	I	R	1.30	1.18-1.47	1.04E-06	496	5.21E-01
rs4476693	5	10882977	CTNND2	0.28	T	C	1.30	1.16-1.46	6.10E-06	542	8.54E-01
rs1180707080	5	<u>76505576</u>	<u>PDE8B_WDR41</u>	<u>0.02</u>	G	T	<u>2.10</u>	<u>1.60-2.72</u>	<u>6.69E-08</u>	<u>453</u>	<u>8.23E-01</u>
rs145242715	5	76805030	WDR41	0.01	A	C	2.30	1.64-3.35	3.11E-06	677	1.00E+00
rs79041957	5	100069852	ST8SIA4	0.06	C	A	1.60	1.33-2.00	2.60E-06	578	4.12E-02
rs80114285	8	49793013	SNAI2	0.02	G	A	2.10	1.51-2.82	5.32E-06	600	3.75E-01
rs7362596	9	10407141	PTPRD	0.23	C	T	1.30	1.17-1.49	7.89E-06	605	7.86E-04
rs182637278	9	93491736	SYK	0.01	C	G	2.30	1.65-3.35	2.35E-06	683	4.48E-02
rs233422	10	8068860	GATA3	0.08	C	T	1.50	1.26-1.79	5.15E-06	575	2.13E-01
rs12606549	18	55022163	ST8SIA3	0.29	T	C	1.30	1.16-1.46	8.43E-06	534	3.91E-04

Abbreviations: HWE, Hardy-Weinberg equilibrium.

^aSample size for 80% power at $\alpha = .05$ is based on the parameter of this population, including minor allele frequency and effect size of the markers.

most showed suggestive significance ($5 \times 10^{-8} \leq p\text{-value} < 1 \times 10^{-5}$). Multiple correction may have affected the results; although the yield of replication findings and a significant meta-analysis support our findings, further validation and international as well as domestic replication studies are warranted. Additionally, the inclusion of association loci based on European ancestry (and replication on such dataset) is another limitation, which is due to the current lack of Asian studies. Lastly, in this study, we observed associations with $p\text{-value} < .05$ based on general GWAS and replication reports (Gonzalez et al., 2016), but from a statistical aspect, said $p\text{-value}$ would not be significant by Bonferroni correction.

In conclusion, we present the first study to identify genetic variants associated with handedness in Koreans. We identified 28 and 15 variants associated with left-handedness and ambidexterity, respectively ($p\text{-value} \leq 10^{-5}$); of these, two and one variants associated with left-handedness and ambidexterity showed genome-wide significance, respectively. Further, one variant each associated with left-handedness and ambidexterity was replicated from the results of a large Western study ($p\text{-value} < .05$). Our results showed limited genetic similarity between left-handedness and ambidexterity in line with previous studies, again suggesting an independent genetic architecture. Further, both handedness-associated variants showed potential relations with genes involved in neurodevelopment or neuropsychiatric traits, suggesting laterality as polygenic and the result of complex neurological genetic pathways as implicated in previous studies. Further studies with larger databases and different ethnicities are warranted; overall, this study hopes to provide a stepping stone for future research into human behavior and brain development.

ACKNOWLEDGMENTS

This study was performed with bioresources from the National Biobank of Korea, the Centers for Disease Control and Prevention, Republic of Korea (2019-059). This study was supported by the Technology Innovation Program (20002781, A Platform for Prediction and Management of Health Risk Based on Personal Big Data and Lifelogging) funded by the Ministry of Trade, Industry and Energy (MOTIE), Korea; an Institute for Information & communications Technology Promotion (IITP) grant funded by the Korea government (Ministry of Science and ICT, MSIT) (2019-31-1293, Autonomous digital companion framework and application); and the Healthcare Bigdata Showcase Project by the Korea Disease Control and Prevention Agency, Republic of Korea (No. 4800-4848-501).

CONFLICT OF INTEREST STATEMENT

The authors declare no conflicts of interest.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding authors upon reasonable request.

ORCID

Ji Won Lee  <https://orcid.org/0000-0002-2666-4249>

PEER REVIEW

The peer review history for this article is available at <https://publons.com/publon/10.1002/brb3.3121>.

REFERENCES

- Ahmed, Z., Douglas, M. R., Read, M. L., Berry, M., & Logan, A. (2011). Citron kinase regulates axon growth through a pathway that converges on cofilin downstream of RhoA. *Neurobiology of Disease*, 41(2), 421–429. <https://doi.org/10.1016/j.nbd.2010.10.012>
- Andrzejczuk, L. A., Banerjee, S., England, S. J., Voufo, C., Kamara, K., & Lewis, K. E. (2018). Tal1, Gata2a, and Gata3 have distinct functions in the development of V2b and cerebrospinal fluid-contacting KA spinal neurons. *Frontiers in Neuroscience*, 12, 170. <https://doi.org/10.3389/fnins.2018.00170>
- Angibaud, J., Louveau, A., Baudouin, S. J., Nerrière-Daguin, V., Evain, S., Bonnemain, V., Hulin, P., Csaba, Z., Dournaud, P., Thinard, R., Naveilhan, P., Noraz, N., Pellier-Monnin, V., & Boudin, H. (2011). The immune molecule CD3zeta and its downstream effectors ZAP-70/Syk mediate ephrin signaling in neurons to regulate early neuritogenesis. *Journal of Neurochemistry*, 119(4), 708–722. <https://doi.org/10.1111/j.1471-4159.2011.07469.x>
- Anney, R. J. L., Lasky-Su, J., Ódúshláíne, C., Kenny, E., Neale, B. M., Mulligan, A., Franke, B., Zhou, K., Chen, W., Christiansen, H., Arias-Vásquez, A., Banaschewski, T., Buitelaar, J., Ebstein, R., Miranda, A., Mulas, F., Oades, R. D., Roeyers, H., Rothenberger, A., ... Sonuga-Barke, E. (2008). Conduct disorder and ADHD: Evaluation of conduct problems as a categorical and quantitative trait in the international multicentre ADHD genetics study. *American Journal of Medical Genetics. Part B, Neuropsychiatric Genetics*, 147B(8), 1369–1378. <https://doi.org/10.1002/ajmg.b.30871>
- Appenzeller, S., Schirmacher, A., Halfter, H., Bäumer, S., Pendziwiat, M., Timmerman, V., De Jonghe, P., Fekete, K., Stögbauer, F., Lüdemann, P., Hund, M., Quabius, E. S., Ringelstein, E. B., & Kuhlenbäumer, G. (2010). Autosomal-dominant striatal degeneration is caused by a mutation in the phosphodiesterase 8B gene. *American Journal of Human Genetics*, 86, 83–87. <https://doi.org/10.1016/j.ajhg.2009.12.003>
- Bai, Y.-H., Zhan, Y.-B., Yu, B., Wang, W.-W., Wang, L., Zhou, J.-Q., Chen, R.-K., Zhang, F.-J., Zhao, X.-W., Duan, W.-C., Wang, Y.-M., Liu, J., Bao, J.-J., Zhang, Z.-Y., & Liu, X.-Z. (2018). A novel tumor-suppressor, CDH18, inhibits glioma cell invasiveness via UQCRC2 and correlates with the prognosis of glioma patients. *Cellular Physiology and Biochemistry*, 48(4), 1755–1770. <https://doi.org/10.1159/000492317>
- Berger, R. P., Sun, Y. H., Kulik, M., Lee, J. K., Nairn, A. V., Moremen, K. W., Pierce, M., & Dalton, S. (2016). ST8SIA4-dependent polysialylation is part of a developmental program required for germ layer formation from human pluripotent stem cells. *Stem Cells*, 34(7), 1742–1752. <https://doi.org/10.1002/stem.2379>
- Bollen, E., & Prickaerts, J. (2012). Phosphodiesterases in neurodegenerative disorders. *IUBMB Life*, 64(12), 965–970. <https://doi.org/10.1002/iub.1104>
- Bandler, W. M., & Paracchini, S. (2014). The genetic relationship between handedness and neurodevelopmental disorders. *Trends in Molecular Medicine*, 20(2), 83–90. <https://doi.org/10.1016/j.molmed.2013.10.008>
- Budini, M., Buratti, E., Morselli, E., & Criollo, A. (2017). Autophagy and its impact on neurodegenerative diseases: New roles for TDP-43 and C9orf72. *Frontiers in Molecular Neuroscience*, 10, 170. <https://doi.org/10.3389/fnmol.2017.00170>
- Carbonell, A. U., Cho, C. H., Tindi, J. O., Counts, P. A., Bates, J. C., Erdjument-Bromage, H., Cvejic, S., Iaboni, A., Kvint, I., Rosensaft, J., Banne, E., Anagnostou, E., Neubert, T. A., Scherer, S. W., Molholm, S., & Jordan, B. A. (2019). Haploinsufficiency in the ANKS1B gene encoding AIDA-1 leads to a neurodevelopmental syndrome. *Nature Communications*, 10(1), 3529. <https://doi.org/10.1038/s41467-019-11437-w>

- Carvill, G. L. (2019). Calcium channel dysfunction in epilepsy: Gain of CACNA1E. *Epilepsy Currents*, 19(3), 199–201. <https://doi.org/10.1177/1535759719845324>
- Chakraborty, P. K., Schmitz-Abe, K., Kennedy, E. K., Mamady, H., Naas, T., Durie, D., Campagna, D. R., Lau, A., Sendamarai, A. K., Wiseman, D. H., May, A., Jolles, S., Connor, P., Powell, C., Heeney, M. M., Giardina, P.-J., Klaassen, R. J., Kannengiesser, C., Thuret, I., & Thompson, A. A. (2014). Mutations in TRNT1 cause congenital sideroblastic anemia with immunodeficiency, fevers, and developmental delay (SIFD). *Blood*, 124(18), 2867–2871. <https://doi.org/10.1182/blood-2014-08-591370>
- Chen, X., Shanoudj-Verber, S., Pollard, L., Hu, Y., Cathey, S. S., Tikkanen, R., & Gray, S. J. (2021). Pre-clinical gene therapy with AAV9/AGA in aspartylglucosaminuria mice provides evidence for clinical translation. *Molecular Therapy*, 29(3), 989–1000. <https://doi.org/10.1016/j.ymthe.2020.11.012>
- Cho, Y. S., Go, M. J., Kim, Y. J., Heo, J. Y., Oh, J. H., Ban, H.-J., Yoon, D., Lee, M. H., Kim, D.-J., Park, M., Cha, S.-H., Kim, J.-W., Han, B.-G., Min, H., Ahn, Y., Park, M. S., Han, H. R., Jang, H.-Y., Cho, E. Y., ... Shin, C. (2009). A large-scale genome-wide association study of Asian populations uncovers genetic factors influencing eight quantitative traits. *Nature Genetics*, 41(5), 527–534. <https://doi.org/10.1038/ng.357>
- Cuellar-Partida, G., Tung, J. Y., Eriksson, N., Albrecht, E., Aliev, F., Andreassen, O. A., Barroso, I., Beckmann, J. S., Boks, M. P., Boomsma, D. I., Boyd, H. A., Breteler, M. M. B., Campbell, H., Chasman, D. I., Cherkas, L. F., Davies, G., De Geus, E. J. C., Deary, I. J., Deloukas, P., ... Duffy, D. L. (2021). Genome-wide association study identifies 48 common genetic variants associated with handedness. *Nature Human Behaviour*, 5(1), 59–70. <https://doi.org/10.1038/s41562-020-00956-y>
- Curley, D. E., Webb, A. E., Sheffler, D. J., & Haass-Koffler, C. L. (2021). Corticotropin releasing factor binding protein as a novel target to restore brain homeostasis: Lessons learned from alcohol use disorder research. *Frontiers in Behaviour Neuroscience*, 15, 786855. <https://doi.org/10.3389/fnbeh.2021.786855>
- Dai, D., Yuan, J., Wang, Y., Xu, J., Mao, C., & Xiao, Y. (2019). Peli1 controls the survival of dopaminergic neurons through modulating microglia-mediated neuroinflammation. *Scientific Reports*, 9(1), 8034. <https://doi.org/10.1038/s41598-019-44573-w>
- Dazzo, E., Fanciulli, M., Serioli, E., Minervini, G., Pulitano, P., Binelli, S., Di Bonaventura, C., Luisi, C., Pasini, E., Striano, S., Striano, P., Coppola, G., Chiavegato, A., Radovic, S., Spadotto, A., Uzzau, S., La Neve, A., Giallonardo, A. T., Mecarelli, O., ... Ottman, R. (2015). Heterozygous reelin mutations cause autosomal-dominant lateral temporal epilepsy. *American Journal of Human Genetics*, 96(6), 992–1000. <https://doi.org/10.1016/j.ajhg.2015.04.020>
- De Kovel, C. G. F., Carrión-Castillo, A., & Francks, C. (2019). A large-scale population study of early life factors influencing left-handedness. *Scientific Reports*, 9(1), 584. <https://doi.org/10.1038/s41598-018-37423-8>
- De Kovel, C. G. F., & Francks, C. (2019). The molecular genetics of hand preference revisited. *Scientific Reports*, 9(1), 5986. <https://doi.org/10.1038/s41598-019-42515-0>
- De La Fuente-Fernández, R., Kishore, A., Calne, D. B., Ruth, T. J., & Stoessl, A. J. (2000). Nigrostriatal dopamine system and motor lateralization. *Behavioural Brain Research*, 112(1–2), 63–68. [https://doi.org/10.1016/S0166-4328\(00\)00165-0](https://doi.org/10.1016/S0166-4328(00)00165-0)
- Dobens, L. L., Nauman, C., Fischer, Z., & Yao, X. (2021). Control of cell growth and proliferation by the tribbles pseudokinase: Lessons from Drosophila. *Cancers (Basel)*, 13(4), 883. <https://doi.org/10.3390/cancers13040883>
- Ehlers, C. L., Gizer, I. R., Bizon, C., Slutske, W., Peng, Q., Schork, N. J., & Wilhelmsen, K. C. (2016). Single nucleotide polymorphisms in the REG-CTNNA2 region of chromosome 2 and NEIL3 associated with impulsivity in a Native American sample. *Genes, Brain, and Behavior*, 15(6), 568–577. <https://doi.org/10.1111/gbb.12297>
- Ensink, J. B. M., Keding, T. J., Henneman, P., Venema, A., Papale, L. A., Alisch, R. S., Westerman, Y., van Wingen, G., Zantvoord, J., Middeldorp, C. M., Mannens, M. M. A. M., Herringa, R. J., & Lindauer, R. J. L. (2021). Differential DNA methylation is associated with hippocampal abnormalities in pediatric posttraumatic stress disorder. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging*, 6(11), 1063–1070.
- Fan, T.-S., Wu, R.-M., Lin, H.-I., Cheng, C., & Lin, C.-H. (2018). PDE8B mutation is not associated with Parkinson's disease in a Taiwanese population. *Neurobiology of Aging*, 71, 265.e15–265.e16. <https://doi.org/10.1016/j.neurobiolaging.2018.05.024>
- Gallup(Korea). (2013). *Gallup report*. <https://www.gallup.co.kr/gallupdb/reportContent.asp?seqNo=491>
- Gonzalez, S., Gupta, J., Villa, E., Mallawaarachchi, I., Rodriguez, M., Ramirez, M., Zavala, J., Armas, R., Dassori, A., Contreras, J., Flores, D., Jerez, A., Ontiveros, A., Nicolini, H., & Escamilla, M. (2016). Replication of genome-wide association study (GWAS) susceptibility loci in a Latino bipolar disorder cohort. *Bipolar Disorders*, 18(6), 520–527. <https://doi.org/10.1111/bdi.12438>
- Goodier, J. L., Soares, A. O., Pereira, G. C., Devine, L. R., Sanchez, L., Cole, R. N., & García-Pérez, J. L. (2020). C9orf72-associated SMCR8 protein binds in the ubiquitin pathway and with proteins linked with neurological disease. *Acta Neuropathologica Communications*, 8(1), 110. <https://doi.org/10.1186/s40478-020-00982-x>
- Hájková, A., Techlovská, Š., Dvořáková, M., Chambers, J. N., Kumpošt, J., Hubálková, P., Prezeau, L., & Blahos, J. (2016). SGIP1 alters internalization and modulates signaling of activated cannabinoid receptor 1 in a biased manner. *Neuropharmacology*, 107, 201–214. <https://doi.org/10.1016/j.neuropharm.2016.03.008>
- Hardyck, C., & Petriniovich, L. F. (1977). Left-handedness. *Psychological Bulletin*, 84(3), 385–404. <https://doi.org/10.1037/0033-2909.84.3.385>
- Higashine, K., Hashimoto, K., Tsujimoto, E., Oishi, Y., Hayashi, Y., & Miyamoto, Y. (2018). Promotion of differentiation in developing mouse cerebellar granule cells by a cell adhesion molecule BT-IgSF. *Neuroscience Letters*, 686, 87–93. <https://doi.org/10.1016/j.neulet.2018.08.049>
- Hildrestrand, G. A., Neurauter, C. G., Diep, D. B., Castellanos, C. G., Krauss, S., Bjørås, M., & Luna, L. (2009). Expression patterns of Neil3 during embryonic brain development and neoplasia. *BMC Neuroscience*, 10, 45. <https://doi.org/10.1186/1471-2202-10-45>
- Ito, J., Nakano, Y., Shima, H., Miwa, T., Kogure, Y., Isshiki, K., Yamazaki, F., Oishi, Y., Morimoto, Y., Kataoka, K., Okita, H., Hirato, J., Ichimura, K., & Shimada, H. (2020). Central nervous system ganglionuroblastoma harboring MYO5A-NTRK3 fusion. *Brain Tumor Pathology*, 37(3), 105–110. <https://doi.org/10.1007/s10014-020-00371-1>
- Jackson, T. C., Janesko-Feldman, K., Carlson, S. W., Kotermanski, S. E., & Kochanek, P. M. (2019). Robust RBM3 and beta-klotho expression in developing neurons in the human brain. *Journal of Cerebral Blood Flow and Metabolism*, 39(12), 2355–2367. <https://doi.org/10.1177/0271678X19878889>
- Jang, H., Lee, J. Y., Lee, K. I., & Park, K. M. (2017). Are there differences in brain morphology according to handedness? *Brain and Behavior*, 7(7), e00730. <https://doi.org/10.1002/brb3.730>
- Jansen, I. E., Ye, H., Heetveld, S., Lechler, M. C., Michels, H., Steinstra, R. I., Lubbe, S. J., Drouet, V., Lesage, S., Majounie, E., Gibbs, J. R., Nalls, M. A., Ryten, M., Botia, J. A., Vandervoort, J., Simon-Sánchez, J., Castillo-Lizardo, M., Rizzu, P., Blauwendraat, C., ... Amin, N. (2017). Discovery and functional prioritization of Parkinson's disease candidate genes from large-scale whole exome sequencing. *Genome biology*, 18(1), 22. <https://doi.org/10.1186/s13059-017-1147-9>
- Kalin, N. H. (2018). Corticotropin-releasing hormone binding protein: Stress, psychopathology, and antidepressant treatment response. *American Journal of Psychiatry*, 175(3), 204–206. <https://doi.org/10.1176/appi.ajp.2018.18010059>
- Kamboh, M. I., Barmada, M. M., Demirci, F. Y., Minster, R. L., Carrasquillo, M. M., Pankratz, V. S., Younkin, S. G., Saykin, A. J., Sweet, R. A., Feingold, E., Dekosky, S. T., & Lopez, O. L. (2012). Genome-wide association analysis of age-at-onset in Alzheimer's disease. *Molecular Psychiatry*, 17(12), 1340–1346. <https://doi.org/10.1038/mp.2011.135>

- Kang, S. J., Kang, K. A., Jang, H., Lee, J. Y., Lee, K. I., Kwoen, M. S., Kim, J. S., & Park, K. M. (2017). Brain morphology according to age, sex, and handedness. *Annals of Clinical Neurophysiology*, 19(2), 93. <https://doi.org/10.14253/acn.2017.19.2.93>
- Ketchesin, K. D., Huang, N. S., & Seasholtz, A. F. (2017). Cell type-specific expression of corticotropin-releasing hormone-binding protein in GABAergic interneurons in the prefrontal cortex. *Frontiers in Neuroanatomy*, 11, 90. <https://doi.org/10.3389/fnana.2017.00090>
- Kim, Y., & Han, B.-G. (2017). Cohort profile: The Korean genome and epidemiology study (KoGES) consortium. *International Journal of Epidemiology*, 46(2), e20. <https://doi.org/10.1093/ije/dyw316>
- Kong, X.-Z., Postema, M., Schijven, D., Castillo, A. C., Pepe, A., Crivello, F., Joliot, M., Mazoyer, B., Fisher, S. E., & Francks, C. (2021). Large-scale phenomic and genomic analysis of brain asymmetrical skew. *Cerebral Cortex*, 31(9), 4151–4168. <https://doi.org/10.1093/cercor/bhab075>
- Kunath, N., Bugaj, A. M., Bigonah, P., Fernandez-Berrocal, M. S., Bjørås, M., & Ye, J. (2021). DNA repair enzyme NEIL3 enables a stable neural representation of space by shaping transcription in hippocampal neurons. *iScience*, 24(12), 103470. <https://doi.org/10.1016/j.isci.2021.103470>
- Li, T., Hu, J., Wang, S., & Zhang, H. (2021). Super-variants identification for brain connectivity. *Human Brain Mapping*, 42(5), 1304–1312. <https://doi.org/10.1002/hbm.25294>
- López Tobón, A., Suresh, M., Jin, J., Vitriolo, A., Pietralla, T., Tedford, K., Bossenz, M., Mahnken, K., Kiefer, F., Testa, G., Fischer, K.-D., & Püschel, A. W. (2018). The guanine nucleotide exchange factor Arhgef7/betaPix promotes axon formation upstream of TC10. *Scientific Reports*, 8(1), 8811. <https://doi.org/10.1038/s41598-018-27081-1>
- Luciano, M., Evans, D. M., Hansell, N. K., Medland, S. E., Montgomery, G. W., Martin, N. G., Wright, M. J., & Bates, T. C. (2013). A genome-wide association study for reading and language abilities in two population cohorts. *Genes, Brain, and Behavior*, 12(6), 645–652. <https://doi.org/10.1111/gbb.12053>
- Michel, G. F. (2021). Handedness development: A model for investigating the development of hemispheric specialization and interhemispheric coordination. *Symmetry*, 13(6), 992. <https://doi.org/10.3390/sym13060992>
- Moon, S., Kim, Y. J., Han, S., Hwang, M. Y., Shin, D. M., Park, M. Y., Lu, Y., Yoon, K., Jang, H.-M., Kim, Y. K., Park, T.-J., Song, D. S., Park, J. K., Lee, J.-E., & Kim, B.-J. (2019). The Korea biobank array: Design and identification of coding variants associated with blood biochemical traits. *Scientific Reports*, 9(1), 1382. <https://doi.org/10.1038/s41598-018-37832-9>
- Naik, R. R., Sotnikov, S. V., Diepold, R. P., Iurato, S., Markt, P. O., Bultmann, A., Brehm, N., Mattheus, T., Lutz, B., Erhardt, A., Binder, E. B., Schmidt, U., Holsboer, F., Landgraf, R., & Czibere, L. (2018). Polymorphism in Tmem132d regulates expression and anxiety-related behavior through binding of RNA polymerase II complex. *Translational Psychiatry*, 8(1), 1. <https://doi.org/10.1038/s41398-017-0025-2>
- Nazarian, A., Yashin, A. I., & Kulminski, A. M. (2019). Genome-wide analysis of genetic predisposition to Alzheimer's disease and related sex disparities. *Alzheimers Research & Therapy*, 11(1), 5. <https://doi.org/10.1186/s13195-018-0458-8>
- Papadatou-Pastou, M., Ntolka, E., Schmitz, J., Martin, M., Munafò, M. R., Ocklenburg, S., & Paracchini, S. (2020). Human handedness: A meta-analysis. *Psychological Bulletin*, 146(6), 481–524. <https://doi.org/10.1037/bul0000229>
- Paracchini, S. (2021). Recent advances in handedness genetics. *Symmetry*, 13(10), 1792. <https://doi.org/10.3390/sym13101792>
- Park, H., Yang, J., Kim, R., Li, Y., Lee, Y., Lee, C., Park, J., Lee, D., Kim, H., & Kim, E. (2015). Mice lacking the PSD-95-interacting E3 ligase, Dorfin/Rnf19a, display reduced adult neurogenesis, enhanced long-term potentiation and impaired contextual fear conditioning. *Scientific Reports*, 5(1), 16410. <https://doi.org/10.1038/srep16410>
- Pawelec, P., Ziemka-Nalecz, M., Sypecka, J., & Zalewska, T. (2020). The impact of the CX3CL1/CX3CR1 axis in neurological disorders. *Cells*, 9(10), 2277. <https://doi.org/10.3390/cells9102277>
- Pearse, D. D., Pereira, F. C., Marcillo, A. E., Bates, M. L., Berrocal, Y. A., Filbin, M. T., & Bunge, M. B. (2004). cAMP and Schwann cells promote axonal growth and functional recovery after spinal cord injury. *Nature Medicine*, 10(6), 610–616. <https://doi.org/10.1038/nm1056>
- Pérez-Torres, S., Cortés, R., Tolnay, M., Probst, A., Palacios, J. M., & Mengod, G. (2003). Alterations on phosphodiesterase type 7 and 8 isozyme mRNA expression in Alzheimer's disease brains examined by *in situ* hybridization. *Experimental Neurology*, 182(2), 322–334. [https://doi.org/10.1016/S0014-4886\(03\)00042-6](https://doi.org/10.1016/S0014-4886(03)00042-6)
- Peterson, B. S., Riddle, M. A., Cohen, D. J., Katz, L. D., Smith, J. C., & Leckman, J. F. (1993). Human basal ganglia volume asymmetries on magnetic resonance images. *Magnetic Resonance Imaging*, 11(4), 493–498. [https://doi.org/10.1016/0730-725X\(93\)90468-S](https://doi.org/10.1016/0730-725X(93)90468-S)
- Regnell, C. E., Hildrestrand, G. A., Sejersted, Y., Medin, T., Moldestad, O., Rolseth, V., Krokeide, S. Z., Suganthan, R., Luna, L., Bjørås, M., & Bergersen, L. H. (2012). Hippocampal adult neurogenesis is maintained by Neil3-dependent repair of oxidative DNA lesions in neural progenitor cells. *Cell Reports*, 2(3), 503–510. <https://doi.org/10.1016/j.celrep.2012.08.008>
- Ruan, X., Liu, G., Zhou, J., Chen, P., Sun, C., Liu, W., Wu, C., Hou, L., Yin, B., Qiang, B., Shu, P., & Peng, X. (2021). Zbed3 is indispensable for Wnt signaling regulation of cortical layers formation in developing brain. *Cerebral Cortex*, 31(9), 4078–4091. <https://doi.org/10.1093/cercor/bhab070>
- Saarela, J., Oinonen, C., Jalanko, A., Rouvinen, J., & Peltonen, L. (2004). Auto-proteolytic activation of human aspartylglucosaminidase. *Biochemical Journal*, 378(2), 363–371. <https://doi.org/10.1042/bj20031496>
- Sarkar, C., Jones, J. W., Hegdekar, N., Thayer, J. A., Kumar, A., Faden, A. I., Kane, M. A., & Lipinski, M. M. (2020). PLA2G4A/cPLA2-mediated lysosomal membrane damage leads to inhibition of autophagy and neurodegeneration after brain trauma. *Autophagy*, 16(3), 466–485. <https://doi.org/10.1080/15548627.2019.1628538>
- Sarver, D. C., Lei, X., & Wong, G. W. (2021). FAM19A (TAFA): An emerging family of neurokines with diverse functions in the central and peripheral nervous system. *Acs Chemical Neuroscience*, 12(6), 945–958. <https://doi.org/10.1021/acscchemneuro.0c00757>
- Stein, J. L., Hibar, D. P., Madsen, S. K., Khamis, M., Mcmahon, K. L., De Zubicaray, G. I., Hansell, N. K., Montgomery, G. W., Martin, N. G., Wright, M. J., Saykin, A. J., Jack, C. R., Weiner, M. W., Toga, A. W., & Thompson, P. M. (2011). Discovery and replication of dopamine-related gene effects on caudate volume in young and elderly populations (N = 1198) using genome-wide search. *Molecular Psychiatry*, 16(9), 927–937. <https://doi.org/10.1038/mp.2011.32>
- Sullivan, P. M., Zhou, X., Robins, A. M., Paushter, D. H., Kim, D., Smolka, M. B., & Hu, F. (2016). The ALS/FTLD associated protein C9orf72 associates with SMCR8 and WDR41 to regulate the autophagy-lysosome pathway. *Acta Neuropathologica Communications*, 4(1), 51. <https://doi.org/10.1186/s40478-016-0324-5>
- Szabo, C. A., Xiong, J., Lancaster, J. L., Rainey, L., & Fox, P. (2001). Amygdalar and hippocampal volumetry in control participants: Differences regarding handedness. *American Journal of Neuroradiology*, 22(7), 1342–1345.
- Tomita, H., Cornejo, F., Aranda-Pino, B., Woodard, C. L., Rioseco, C. C., Neel, B. G., Alvarez, A. R., Kaplan, D. R., Miller, F. D., & Cancino, G. I. (2020). The protein tyrosine phosphatase receptor delta regulates developmental neurogenesis. *Cell Reports*, 30(1), 215–228.e5. <https://doi.org/10.1016/j.celrep.2019.11.033>
- Tsai, L.-C. L., Chan, G. C.-K., Nangle, S. N., Shimizu-Albergue, M., Jones, G. L., Storm, D. R., Beavo, J. A., & Zweifel, L. S. (2012). Inactivation of Pde8b enhances memory, motor performance, and protects against age-induced motor coordination decay. *Genes, Brain, and Behavior*, 11(7), 837–847. <https://doi.org/10.1111/j.1601-183X.2012.00836.x>
- Turner, T. N., Sharma, K., Oh, E. C., Liu, Y. P., Collins, R. L., Sosa, M. X., Auer, D. R., Brand, H., Sanders, S. J., Moreno-De-Luca, D., Pihur, V., Plona, T., Pike, K., Soppet, D. R., Smith, M. W., Cheung, S. W., Martin, C. L., State, M. W., Talkowski, M. E., ... Huganir, R. (2015). Loss of delta-catenin function

- in severe autism. *Nature*, 520(7545), 51–56. <https://doi.org/10.1038/nature14186>
- Wang, G., Li, Y., Wang, X.-Y., Han, Z., Chuai, M., Wang, L.-J., Ho Lee, K. K., Geng, J.-G., & Yang, X. (2013). Slit/Robo1 signaling regulates neural tube development by balancing neuroepithelial cell proliferation and differentiation. *Experimental Cell Research*, 319(8), 1083–1093. <https://doi.org/10.1016/j.yexcr.2013.02.011>
- Wang, L.-S., Hranilovic, D., Wang, K., Lindquist, I. E., Yurcaba, L., Petkovic, Z.-B., Gidaya, N., Jernej, B., Hakonarson, H., & Bucan, M. (2010). Population-based study of genetic variation in individuals with autism spectrum disorders from Croatia. *BMC Medical Genetics*, 11, 134. <https://doi.org/10.1186/1471-2350-11-134>
- Wei, Q., Nakahara, F., Asada, N., Zhang, D., Gao, X., Xu, C., Alfieri, A., Brodin, N. P., Zimmerman, S. E., Mar, J. C., Guha, C., Guo, W., & Frenette, P. S. (2020). Snai2 maintains bone marrow niche cells by repressing osteopontin expression. *Developmental Cell*, 53(5), 503–513.e5. <https://doi.org/10.1016/j.devcel.2020.04.012>
- Wetherill, L., Kapoor, M., Agrawal, A., Bucholz, K., Koller, D., Bertelsen, S. E., Le, N., Wang, J.-C., Almasy, L., Hesselbrock, V., Kramer, J., Nurnberger, J. I., Schuckit, M., Tischfield, J. A., Xuei, X., Porjesz, B., Edenberg, H. J., Goate, A. M., & Foroud, T. (2014). Family-based association analysis of alcohol dependence criteria and severity. *Alcoholism, Clinical and Experimental Research*, 38(2), 354–366. <https://doi.org/10.1111/acer.12251>
- Wiberg, A., Ng, M., Al Omran, Y., Alfaro-Almagro, F., McCarthy, P., Marchini, J., Bennett, D. L., Smith, S., Douaud, G., & Furniss, D. (2019). Handedness, language areas and neuropsychiatric diseases: Insights from brain imaging and genetics. *Brain*, 142(10), 2938–2947. <https://doi.org/10.1093/brain/awz257>
- Yao, J., De La Iglesia, H. O., & Bajjalieh, S. M. (2013). Loss of the SV2-like protein SVOP produces no apparent deficits in laboratory mice. *PLoS One*, 8(7), e68215. <https://doi.org/10.1371/journal.pone.0068215>
- Zheng, M., McBride, C., Ho, C. S.-H., Chan, J. K.-C., Choy, K. W., & Paracchini, S. (2020). Prevalence and heritability of handedness in a Hong Kong Chinese twin and singleton sample. *BMC Psychology*, 8(1), 37. <https://doi.org/10.1186/s40359-020-00401-9>

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Song, Y., Lee, D., Choi, J.-E., Lee, J. W., & Hong, K.-W. (2023). Genome-wide association and replication studies for handedness in a Korean community-based cohort. *Brain and Behavior*, e3121. <https://doi.org/10.1002/brb3.3121>