



Treatment outcome and prognostic factors of Korean patients with chronic lymphocytic leukemia: a multicenter retrospective study

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Background/Aims: Compared with Western countries, chronic lymphocytic leukemia (CLL) rarely occurs in Asia and has different clinical characteristics. Thus, we aimed to evaluate the clinical characteristics, treatment outcomes, and prognostic significance of Korean patients with CLL.

Methods: We retrospectively analyzed 90 patients with CLL who had received chemotherapy at 6 centers in Korea between 2000 and 2012.

Results: Compared with Western patients with CLL, Korean patients with CLL express lambda (42.0%) and atypical markers such as CD22 and FMC7 (76.7% and 40.0%, respectively) more frequently. First-line chemotherapy regimens included chlorambucil (n = 43), fludarabine and cyclophosphamide (FC) (n = 20), fludarabine (n = 13), rituximab-FC (n = 4). The remaining patients were treated with other various regimens (n = 10). The 5-year overall survival (OS) and progression-free survival (PFS) rates were 79.3% and 28.1%, respectively. Multivariate analyses showed that hyperleukocytosis ($\geq 100 \times 10^3/\mu\text{L}$), extranodal involvement, and the Binet C stage were significant negative prognostic factors for OS (hazard ratio [HR] 4.75, $p = 0.039$; HR 21.6, $p = 0.002$; and HR 4.35, $p = 0.034$, respectively). Cytogenetic abnormalities including complex karyotypes (≥ 3), del(11q), and del(17) had a significantly adverse impact on both OS and PFS ($p < 0.001$ and $p = 0.010$, respectively).

Conclusions: Initial hyperleukocytosis, extranodal involvement, complex karyotype, del(17) and del(11q) need to be considered in the risk stratification system for CLL.

Keywords: Leukemia, lymphocytic, chronic, B-cell; Treatment outcome; Prognosis; Korean

INTRODUCTION

Chronic lymphocytic leukemia (CLL) is a mature B cell neoplasm, usually with coexpressed cluster of differentiation 5 (CD5) and CD23 [1]. The diagnostic criteria of CLL requires the presence of at least $5 \times 10^9/\text{L}$ monoclonal B lym-

phocytes with the CLL phenotype in peripheral blood [2]. To determine the disease extent and prognosis, Rai and Binet clinical staging systems have been used to predict the prognostic risks of CLL for > 3 decades [2-4]. With the advancement of genetic analysis techniques, genetic abnormalities were

found to have important clinical implications. Immunoglobulin heavy-chain variable (IGHV) gene somatic hypermutations are found in 50% to 60% of CLL cases and are associated with more favorable outcomes [5,6]. About 80% of the patients have cytogenetic abnormalities detected by fluorescence *in situ* hybridization (FISH) [7]. Del(11q), del(17p), and del(6q) were associated with chemotherapy resistance and worse outcome [6]. Del(13q), however, has been known to predict a more favorable clinical course [7].

CLL is the most common type of leukemia in Western countries; however, CLL rarely occurs in Asian populations [8-10]. In Korea, the annual incidence of CLL is only 0.11 per 100,000 people, while that in the Western countries is 2.8 to 3.9 per 100,000 people, which is 25 to 35 times higher than that in Korea [10,11]. Asian CLL has been reported to have different immunophenotypes and clinical characteristics [8,12]. Therefore, the clinical outcomes and prognostic factors after chemotherapy in Asian patients with CLL might be different from those of Western patients with CLL. However, only a few studies have been reported on CLL in Asia because of the limited patient population. Therefore, we aimed to investigate the clinical characteristics, treatment outcomes, and prognostic significances of Korean patients with CLL.

METHODS

We retrospectively analyzed 90 newly diagnosed patients with CLL who received chemotherapy at one of six centers in Korea between 2000 and 2012. CLL diagnosis and response evaluation were based on the International Workshop on CLL criteria, updated in 2008. Patients with small lymphocytic lymphoma were excluded in this study. Data on baseline clinicopathological features, treatment modalities, and outcomes were obtained from the medical records of each patient. Cytogenetics and FISH were performed using the bone marrow samples of 73 patients and 27 patients, respectively, at the time of CLL diagnosis. Immunophenotypic data at CLL diagnosis were available in 66 patients. Surface antigen expression on leukemic blasts was determined by 4-color flow cytometric analysis using either the FACScan system and Cell Quest software or the

FACSCanto system and FACS Diva software (all from Becton Dickinson, San Jose, CA, USA). A CD45 blast gating method was used to distinguish leukemic blasts from other cells. The following antibodies directed to membrane antigens were used for standard immunophenotypic analyses of CLL cells: anti-CD3, anti-CD5, anti-CD7, anti-CD10, anti-CD19, anti-CD20, anti-CD22, anti-CD23, FMC7, surface membrane immunoglobulin (SmIg; kappa and lambda), HLA-DR (Becton Dickinson, Franklin Lakes, NJ, USA), and terminal deoxynucleotidyl transferase (TdT; Dako, Glostrup, Denmark). Positivity for each surface antigen was defined as expression on $\geq 20\%$ leukemic cells, except for TdT positivity, which was defined as expression on $\geq 10\%$ leukemic cells.

This study was approved by the Institutional Review Boards (Konkuk University Medical Center, KUH 1010527; and Ulsan University Hospital, 2015-04-008) of all the participating hospitals, Korea. Written informed consent by the patients was waived due to a retrospective nature of our study.

Statistical analysis

Continuous variables were compared using Student's *t* test, whereas categorical variables were analyzed using either the Pearson's chi-square test, or Fisher's exact test. Survival probabilities were estimated using the Kaplan-Meier method and differences in survival distributions were compared using the log-rank test. Overall survival (OS) was calculated from the date of CLL diagnosis to the date of death or last follow-up. Progression-free survival (PFS) was measured from the time of initial chemotherapy to the date of progression or death from any cause. Among patients attaining complete remission (CR) after induction chemotherapy, relapse-free survival (RFS) was defined from the time of CR to the date of relapse, or death from any cause. After chemotherapy, the overall response rate was defined as CR plus the partial remission (PR) rates. Factors affecting CR achievement were determined by either Pearson's chi-square test, or Fisher's exact test. Factors affecting OS, PFS, and RFS were analyzed using the log-rank test in univariate analyses, and the Cox proportional hazard model in multivariate analyses. The following variables were included in the analysis of prognostic factors: age, white blood cell (WBC) and platelet counts, hemoglobin level, presence of constitutional symptoms, lymphade-

nopathy or splenomegaly at CLL diagnosis, expression of each surface antigen on leukemic blasts, cytogenetics, Rai stage, Binet system, chemotherapeutic regimens, and responses following chemotherapy. All statistical analyses were performed using the SPSS version 21.0 software (IBM Corp., Armonk, NY, USA). Variables with $p < 0.1$ in univariate analyses were included in the multivariate analyses. For all analyses, p values were 2 sided, and $p < 0.05$ was considered statistically significant.

RESULTS

Patients

The baseline characteristics of the patients with CLL at diagnosis are listed in Table 1. The study population comprised 53 male and 37 female patients; the median age was 61.5 years (range, 25 to 93), and all included patients received systemic chemotherapy. The median time from diagnosis to treatment was 1.0 months (range, 0 to 128.6). The clinical characteristics and chemotherapeutic regimens at the time of chemotherapy are summarized in Supplementary Table 1. Approximately 43.3% of all treated patients were treated in Rai stages III or IV. Patients having either symptoms related to CLL including lymphadenopathy, splenomegaly, and constitutional symptoms or significantly high leukocyte counts were treated in Rai stage 0 (25.6%), I (14.7%), and II (15.6%). First-line chemotherapy comprised oral chlorambucil ($n = 43$, 47.8%), fludarabine and cyclophosphamide (FC) ($n = 20$, 22.2%); fludarabine alone ($n = 13$, 14.4%); fludarabine, cyclophosphamide, and rituximab (FCR) ($n = 4$, 4.4%); fludarabine and rituximab (FR) ($n = 1$ [1.1%]); and cyclophosphamide, vincristine, and prednisolone (CVP) ($n = 4$, 4.4%). The remaining patients ($n = 5$) received other various regimen including vincristine plus prednisolone (VP) or cyclophosphamide plus prednisolone. The second-line chemotherapies were administered in 52 patients, of whom 16 received chlorambucil, and 12 received fludarabine. In addition, FCR ($n = 5$), FC ($n = 3$), FR ($n = 3$), and other various regimens ($n = 13$) including CVP; VP plus cyclophosphamide, adriamycin, vincristine, and prednisolone (CHOP); rituximab plus CHOP (R-CHOP); and fludarabine, cyclophosphamide, and mitoxantrone (FCM) were administered as second-line chemotherapy. Third-line chemotherapy was performed in 26 patients.

Table 1. Baseline characteristics at diagnosis

Characteristic	Total (n = 90)
Sex, male/female (n = 90)	53 (58.9)/37 (41.1)
Age, yr (n = 950)	61.5 (25–93)
Hypertension (n = 89)	22 (24.4)
Diabetes mellitus (n = 89)	11 (12.2)
History of cancer (n = 90)	4 (4.4)
Comorbidity (n = 89) ^a	16 (17.8)
Constitutional symptom (n = 89)	14 (15.6)
Lymphadenopathy (n = 84)	45 (50.0)
Bulky lymph node (> 5 cm) (n = 90)	2 (2.2)
Hepatomegaly (n = 82)	4 (4.4)
Splenomegaly (n = 83)	28 (31.1)
Extranodal involvement (n = 90)	4 (4.4)
WBC count, $\times 10^3/\mu\text{L}$ (n = 90)	27.4 (3.83–447.4)
Hemoglobin, g/dL (n = 90)	12.6 (2.1–19.9)
Platelet, $\times 10^3/\mu\text{L}$ (n = 90)	175.5 (5–308)
Lymphocyte count, $\times 10^3/\mu\text{L}$ (n = 88)	22.8 (1.4–438.5)
Bone marrow cellularity, % (n = 68)	60 (10–100)
Bone marrow neoplastic cells, % (n = 63)	77.0 (9.0–96.1)
LDH (n = 73)	285 (112–1112)
Stage (Binet system) (n = 90)	
A	54 (60.0)
B	9 (10.0)
C	27 (30.0)
Cytogenetics (n = 73) ^b	
Normal karyotype	60 (66.7)
Inv(9)/dup(9)	3 (3.3)
Del(13q)	1 (1.1)
Del(11q)	2 (2.2)
Trisomy 12	2 (2.2)
Complex karyotype (≥ 3)	3 (3.3)
Others	2 (2.2)
FISH	
IgH gene rearrangement (n = 20)	6
Del(13q) (n = 18)	7
Trisomy 12 (n = 21)	6
Del(11q) (n = 20)	2
Del(17p) (n = 25)	2

Values are presented as number (%) or median (range). WBC, white blood cell; LDH, lactate dehydrogenase; FISH, fluorescence *in situ* hybridization; IgH, immunoglobulin heavy locus.

^aComorbidity includes cerebrovascular disease, heart disease, liver disease, and lung disease other than hypertension, diabetes and prior cancers.

^bAt the time of diagnosis.

Table 2. Immunophenotypic characteristics

Immunophenotype (patient no. analyzed for the immunophenotype)	No. of positive patient (%)
CD3 (+) (n = 40)	3 (7.5)
CD5 (+) (n = 64)	50 (78.1)
CD7 (+) (n = 55)	4 (7.3)
CD19 (+) (n = 66)	63 (95.5)
CD20 (+) (n = 57)	54 (94.7)
CD22 (+) (n = 43)	33 (76.7)
CD23 (+) (n = 41)	37 (90.2)
CD45 (+) (n = 22)	22 (100)
CD56 (+) (n = 16)	0
FMC7 (+) (n = 30)	12 (40.0)
HLA-DR (+) (n = 24)	24 (100)
TdT (+) (n = 29)	3 (10.3)
SmIg (n = 50)	
Kappa (+)	19 (38.0)
Lambda (+)	21 (42.0)
Negative	10 (20.0)

TdT, terminal deoxynucleotidyl transferase; SmIg, surface membrane immunoglobulin.

Only two patients who achieved PR, and had stable disease after chemotherapy underwent allogeneic hematopoietic stem cell transplantation (allo-HCT).

Immunophenotypic characteristics

Immunophenotypic findings of CLL patients are summarized in Table 2. CD19 expression was observed in all tested patients with B cell CLL, except three with T cell-type CLL. Approximately 78.1% of all tested patients showed CD5 positivity, while 14 patients presented with atypical CLL (CD5 negative). Interestingly, CD22 and FMC7 positivity was frequently found in 76.7% (33/43) and 40.0% (12/30) of the patients, respectively. Expression of kappa and lambda SmIg were observed in 38.0% and 42.0% of the patients, respectively.

Chemotherapy responses

The clinical characteristics of patients at the time of chemotherapy are summarized in Supplementary Table 1. Twelve patients (12.6%) presented with constitutional symptoms such as fever, night sweats, or significant weight loss ($\geq 10\%$ for 6 months). Among 45

Table 3. Treatment outcomes after chemotherapy

Variable	CR rate	Response rate (CR + PR)
1st line chemotherapy regimen (n = 90)		
Chlorambucil (n = 43)	7 (16.3)	29 (67.5)
Fludarabine (n = 13)	1 (7.7)	7 (63.6)
FCR (n = 4)	1 (25)	3 (75.0)
FC (n = 20)	1 (35.0)	18 (90)
CVP (n = 4)	0	3 (75.0)
RF (n = 1)	0	1 (100)
Others (hydroxyurea, VP, Cy + PD) (n = 5)	0	2 (40.0)
Cytogenetics (n = 73) ^a		p = 0.011
IgH rearrangement (n = 6)	4 (66.7)	
Normal karyotype/ others (n = 62)	11 (17.7)	
Del(11q)/del(17)/CK (n = 5)	0	

Values are presented as number (%).

CR, complete remission; PR, partial remission; FCR, fludarabine, cyclophosphamide, and rituximab; FC, fludarabine and cyclophosphamide; CVP, cyclophosphamide, vincristine and prednisolone; RF, rituximab and fludarabine; VP, vincristine and prednisolone; Cy + PD, cyclophosphamide and prednisolone; IgH, immunoglobulin heavy locus; CK, complex karyotype.

^aAt the initiation of first-line chemotherapy.

patients who received chemotherapy at Binet A, 40.0% had progressive higher WBC counts $> 40.0 \times 10^3/\mu\text{L}$, 52.5% had symptomatic lymphadenopathy or splenomegaly, and 11.4% had constitutional symptoms. After administering first-line chemotherapy, 18.9% of the patients achieved CR. The CR rate after the maximum third-line chemotherapy was 27.8%. Only two patients achieved a CR after third-line chemotherapy. The overall response rates were 70.0%, and 32.2% after first-, and second-line chemotherapy, respectively. Among the 25 patients who achieved CR, 12 (48.0%) experienced relapse. Two patients with refractory CLL who underwent allo-HCT survived after achieving CR status. The CR rates according to initial chemotherapeutic regimen are summarized in Table 3. The CR rates of the FCR, and FC regimens were 25%, and 35%, respectively (Table 3). There were, however, significant differences in the CR rate regarding cytogenetic abnormalities ($p = 0.011$) (Ta-

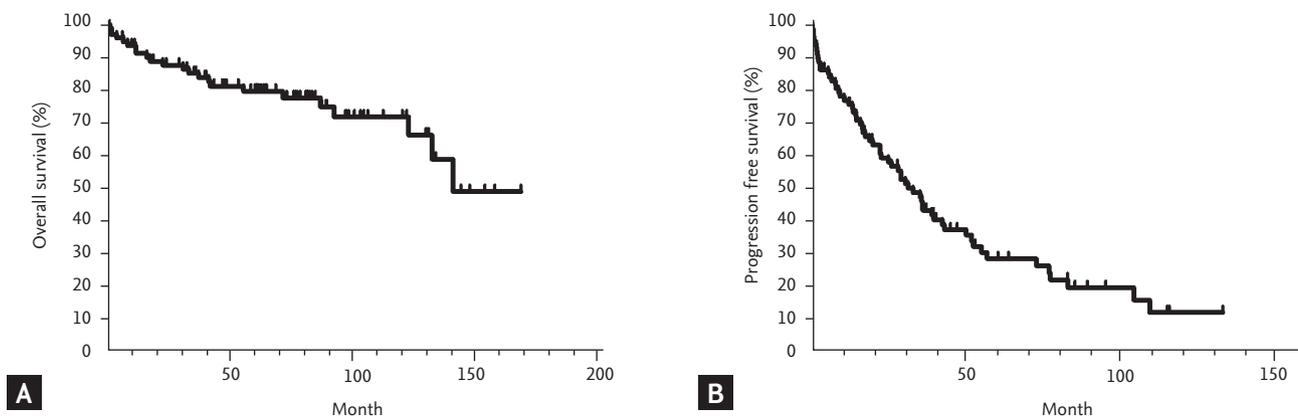


Figure 1. Survivals of Korean patients with chronic lymphocytic leukemia. (A) Overall survival and (B) progression-free survival.

ble 3). Patients with immunoglobulin heavy locus (IgH) rearrangement showed a significantly higher CR rate than those with no IgH rearrangement (66.7% vs. 17.2%, respectively; $p = 0.011$). However, patients with del(11q), del(17), or complex karyotype (≥ 3 chromosomal abnormalities) did not achieve CR.

Univariate analysis for survival outcomes

The median follow-up duration for surviving patients was 80.5 months (range, 2.0 to 167.8). The 5-year survival rate for all included patients was 79.3%, and the median OS was not reached (Fig. 1A). During the follow-up period, 23 patients with persistent disease died, either from infection ($n = 12$), or disease progression ($n = 11$). After the initial chemotherapy regimen, the median PFS was 31.0 months (95% confidence interval [CI], 23.1 to 39.1) and the 5-year PFS rate was 28.1% (Fig. 1B). The median RFS of the 25 patients who achieved CR after chemotherapy was 50.3 months (95% CI, 8.6 to 91.9), while 12 patients experienced relapse. The 5-year RFS rate for the patients who achieved CR was 47.3%. Univariate analyses for OS and PFS according to the clinical parameters are summarized in Table 4. The traditional staging criteria according to Rai system and Binet system had significant effects on the OS ($p = 0.001$, and $p = 0.006$, respectively). Lymphadenopathy, splenomegaly, and extranodal involvement were significantly associated with lower OS rates ($p = 0.029$, $p = 0.003$, and $p = 0.008$, respectively). Initial hyperleukocytosis ($WBC \geq 100 \times 10^3/\mu L$), anemia (hemoglobin < 10 g/dL), and thrombocytopenia (platelet count $< 80 \times 10^3/\mu L$) were significantly associated with

worse OS ($p < 0.001$, $p = 0.004$, and $p = 0.032$, respectively). In contrast to the risk classification of the Rai and Binet systems, in which platelet count less than $100 \times 10^3/\mu L$ was associated with worst prognosis, our data showed that patients with a platelet count between 80×10^3 and $110 \times 10^3/\mu L$ had survival rates comparable to those with higher platelet counts ($> 110 \times 10^3/\mu L$) (5-year OS rate: 92.9% vs. 82.0%, respectively), whereas those with a lower platelet count ($< 80 \times 10^3/\mu L$) had a significantly lower OS rate (46.7%, $p = 0.032$). In addition, the univariate analyses revealed that higher proportions of neoplastic cells in the bone marrow ($\geq 80\%$), and higher lactate dehydrogenase (LDH) (≥ 500 IU/L) levels were significantly associated with worse OS rates ($p = 0.021$, and $p = 0.003$, respectively). In particular, higher LDH levels (> 500 IU/L) were also an adverse factor for predicting PFS ($p = 0.016$). Lymphadenopathy tended to be associated with worse PFS ($p = 0.067$). Cytogenetic abnormalities had a significant impact on OS and PFS ($p = 0.002$ and $p = 0.006$, respectively). IgH gene rearrangement was a significant favorable factor for OS (5-year OS and PFS: 100% and 50%, respectively). Compared with normal karyotypes and other karyotypes, del(11q), del(17), and complex karyotypes (≥ 3 chromosomal abnormalities) were significant negative prognosticators (5-year OS and PFS rates: 82.9% vs. 26.7%, $p = 0.002$; and 29.7% vs. 0%, $p = 0.006$, respectively).

With regard to immunophenotypic markers, expression of lambda SmIg showed a significantly worse PFS compared to expression of kappa SmIg, or the absence of both (5-year PFS rate: 6.3% vs. 40.4%, and 21.1%, re-

Table 4. Univariate analysis for survivals

Variable	5-year OS, %	p value	5-year PFS, %	p value
Sex, male/female (n = 90)	76.4/84.3	0.544	24.3/32.8	0.701
Age, < 65/≥ 65 yr (n = 90)	85.0/68.8	0.224	30.6/21.0	0.235
Stage (Rai system) ^a (n = 90)		0.001		0.038
0	100		26.0	
I	75.5		17.8	
II	73.3		64.8	
III	66.1		21.4	
IV	68.2		25.0	
Stage (Binet) (n = 90), % ^a		0.006		0.239
A	88.0		27.3	
B	72.9		53.3	
C	62.2		20.0	
Constitutional symptom (+/-) (n = 89), %	76.4/84.3	0.266	25.4/26.9	0.869
Lymphadenopathy ^a (+/-) (n = 84)	68.5/86.9	0.029	24.2/36.3	0.127
Splenomegaly ^a (+/-) (n = 83), %	63.4/84.0	0.003	33.2/25.7	0.340
Extranodal involment ^a (+/-) (n = 90), %	37.5/81.0	0.008	50.0/27.9	0.639
WBC count ^a , < 100/≥ 100 × 10 ³ /μL (n = 90)	83.9/28.6	< 0.001	29.3/14.3	0.049
Hemoglobin ^b , < 10/≥ 10 g/dL (n = 89)	55.9/85.7	0.004	11.8/30.6	0.056
Platelet ^b , < 80/80 to < 110/≥ 110 × 10 ³ /μL (n = 90)	46.7/92.3/81.7	0.032	13.3/0/33.3	0.174
Bone marrow neoplastic cells, < 80%/≥ 80% (n = 63)	88.0/73.1	0.021	28.7/41.4	0.296
LDH, < 500/≥ 500 IU/L (n = 70)	84.3/34.3	0.003	28.8/0	0.016
Cytogenetics (n = 75) ^a		0.002		0.006
IgH mutation	100		50.0	
Normal karyotype/other chromosomes	82.3		29.7	
Del(11q) or del(17) or CK	26.7		0	
Immunophenotyping, %				
SmIg (n = 50)		0.623		0.572
Kappa (+)	83.1		35.6	
Lambda (+)	76.0		13.6	
Negative	87.5		21.1	
CD5 (n = 37)		0.856		0.018
Bright	100		0	
Intermediate	88.4		21.6	
Dim/Negative	82.5		29.3	
CD20 (n = 30)		0.138		0.058
Bright	90.0		53.5	
Intermediate/dim/negative	76.9		12.7	
Initial chemotherapy regimen (n = 90)		0.255		0.007
FCR or RF	75.0		53.3	
FC	75.6		46.6	
Chlorambucil	85.1		23.0	
Fludarabine	59.8		15.4	

Table 4. Continued

Variable	5-year OS, %	<i>p</i> value	5-year PFS, %	<i>p</i> value
Others	88.9		12.5	
1st line chemotherapy response (n = 87)		< 0.001		< 0.001
CR	100		66.2	
PR	83.3		26.9	
SD	66.9		0	
PD	37.5		0	

OS, overall survival; PFS, progression-free survival; WBC, white blood cell; LDH, lactate dehydrogenase; IgH, immunoglobulin heavy locus; CK, complex karyotype; CD, cluster of differentiation; FCR, fludarabine, cyclophosphamide, and rituximab; RF, rituximab and fludarabine; FC, fludarabine and cyclophosphamide; CR, complete remission; PR, partial remission; SD, stable disease; PD, progressive disease.

^aAt diagnosis.

^bAt the initiation of first-line chemotherapy.

spectively, *p* = 0.040). The bright expression of CD5 was significantly associated with a worse PFS, compared to intermediate or dim/negative expression of CD5 (0% vs. 21.6, and 29.3%, respectively, *p* = 0.018). On the contrary, patients with bright expression of CD20 tended to show higher PFS rates than those with intermediate or dim expression of CD20 (53.5% vs. 12.7%, *p* = 0.058). Apart from light chain, CD5, and CD20 expression, there were no significant differences in the survival outcomes among patients in terms of other immunophenotypic markers.

The initial chemotherapy regimens including FCR or RF, FC, chlorambucil, fludarabine, and other various regimens, significantly affected the 5-year PFS (53.3%, 46.6%, 23.0%, 15.4%, and 12.5%, respectively, *p* = 0.007) and RFS rates (100%, 87.5%, 18.2%, 0%, and 0%, respectively, *p* = 0.004), although there was no significant difference in OS (*p* = 0.255). The response after initial chemotherapy was a significant predictor of OS (*p* < 0.001) and PFS (*p* < 0.001) (Table 4). All patients who achieved a CR even after second-, or third-line chemotherapy, or allogeneic stem cell transplantation survived more than 5 years.

Multivariate analysis

Multivariate analysis showed that initial hyperleukocytosis (WBC count ≥ 100 × 10³/μL) (HR, 4.748; *p* = 0.039) and extranodal involvement (HR, 21.639; *p* = 0.002), Binet stage C (HR, 4.354; *p* = 0.034), and chromosomal abnormalities including del(11q), del(17), or complex karyotype (≥ 3 chromosomal abnormalities) (HR, 17.022; *p* < 0.001)

were powerful adverse prognostic factors for OS and PFS (HR 17.022; *p* < 0.001; and HR 9.620; *p* = 0.010; respectively) (Table 5). Moreover, the light chain types of SmIg had a significant impact on PFS.

DISCUSSION

Among hematologic malignancies, CLL is a disease that demonstrates the greatest genetic differences between Western and Asian people [9]. CLL rarely occurs in Korean people, and only 56 patients are newly diagnosed with CLL annually [8]. The number of Korean patients with CLL has been slowly increasing [13]. The crude incidence rate of CLL or small lymphocytic lymphoma has increased from 0.19, to 0.31 between 1999 and 2012 according to the national cancer statistics [13]. Korean patients with CLL in our study were relatively young, with a median age at diagnosis of 61 years. In contrast, the median age at diagnosis in the United States is 70 years [9,14]. In general, younger age may favorably affect the treatment outcomes of hematologic malignancies. However, CLL occurring in Asian people showed a more aggressive progression than that occurring in Caucasian people [14]. Gunawardana et al. [14] reported that Asian patients required treatment almost twice as often as Caucasian patients (HR, 1.94) and had a shorter time to first treatment (*p* = 0.0063). A previous study from India suggested that a higher proportion of patients with CLL presented with an advanced clinical stage [15].

Table 5. Multivariate analysis

Variable	Overall survival		Progression-free survival	
	HR (95% CI)	p value	HR (95% CI)	p value
WBC count, $\geq 100, \times 10^3/\mu\text{L}$ ^a	4.748 (1.084–20.792)	0.039	-	-
Extranodal involvement ^a	21.639 (2.972–157.553)	0.002	-	-
Binet system ^a				
A	Reference		-	
B	0.455 (0.040–5.108)	0.523	-	
C	4.354 (1.117–16.975)	0.034	-	
Cytogenetics ^a				
Normal karyotype/other chromosomes	Reference		Reference	
Del(11q) or del(17) or CK	17.022 (3.674–78.864)	< 0.001	9.620 (1.732–53.444)	0.010
IgH mutation	-	0.988	0.503 (0.114–2.215)	0.364

HR, hazard ratio; CI, confidence interval; WBC, white blood cell; CK, complex karyotype (≥ 3); IgH, immunoglobulin heavy locus.

^aAt the time of diagnosis.

In the present study, Korean patients with CLL expressed a unique immunophenotypic pattern on the CLL cells. Compared with Western data, expression of atypical immunophenotypic markers such as CD22 and FMC7 were frequently observed among Korean CLL patients. Similarly, these findings were previously reported in a small retrospective study conducted by Jang et al. [12]. In addition, expression of lambda SmIg was more frequent among Asian patients with CLL than among Western patients. Even though there were no significant differences in univariate and multivariate analysis, lambda SmIg expression tended to be associated with a worse PFS compared with kappa SmIg expression. Therefore, compared to CLL observed among Western patients, a relatively greater proportion of patients with lambda SmIg expression in Korean patients with CLL might influence the treatment outcome, which was worse than that of Western patients with CLL. In terms of the clinical implication of immunophenotypic markers, we found that the intensity of the CD5 and CD20 expression was more important than the positivity thereof. Bright CD5 expressions was a significant predictor for worse PFS, compared to both intermediate-, and dim/negative expression.

Among the first-line chemotherapeutic regimens, the FCR and FC regimens achieved a CR rate of 25%, and response rate of 75%. The FCR regimen has been recommended as first-line chemotherapy in patients younger than 65 years without significant comorbidities accord-

ing to the 2018 National Comprehensive Cancer Network guidelines [16]. The CR and response rates of Korean patients with CLL treated with FCR seems to be lower than that reported in the CLL10 study [16]. Relatively smaller numbers of patients received rituximab-containing chemotherapy as first-, or second-line chemotherapy in our data. Nowadays, several novel agents including the Bruton's tyrosine kinase inhibitor (ibrutinib), anti-CD20 monoclonal antibody (obinutuzumab), bendamustine, and ofatumumab have been introduced as first-line therapy for CLL [8,17-19]. Unfortunately, the national medical insurance reimbursement system in Korea does not cover the expenses for the procurement of those novel agents as first-line chemotherapy for CLL. Therefore, our data have some limitations, as we were unable to include the therapeutic experiences of patients who were treated using newer agents.

OS of Korean patients with CLL who received chemotherapy was not reached, and the 5-year PFS rate was only 28.1%. Although the Rai, and Binet staging systems were introduced more than 37 years ago, these two prognostic systems have been widely used to stratify the prognosis of patients with CLL [3,4]. With regard to the therapeutic outcome prediction in CLL, genetic abnormalities have important clinical implications. There might be ethnic differences in the clinical implication of genetic aberrations, including IgH mutations. In our study, IgH gene rearrangement was associated with a higher CR rate. In the CLL10 study, however, IgH mutations had no influ-

ence on either CR rate, or overall response rate in both treatment arms (FCR vs. bendamustine and rituximab) [16]. Del(11q), del(17p), and del(6q) have been reported to be associated with a worse outcome [6,7]. Previous large-scale studies reported that patients with del(17p) and TP53 mutations had poorer chemotherapy responses and survival rates [20-22]. We also demonstrated that patients with a complex karyotype (≥ 3 cytogenetic abnormalities) as well as del(11q) and del(17p) were important adverse factors in terms of CR achievement, OS, and PFS in patients with CLL. None of the patients with a complex karyotype (≥ 3) as well as del(11q) and del(17p) attained a CR after chemotherapy. In CLL, the prognostic significance of a complex karyotype had also been reported in other recent studies [23-25].

For the prediction of CLL prognosis, Wierda et al. [26] suggested that age, absolute lymphocyte count, beta 2-microglobulin, sex, and nodal involvement were significant prognostic factors in addition to Rai staging. Similarly, we also demonstrated that hyperleukocytosis ($\geq 100 \times 10^3/\mu\text{L}$) was an independently significant poor prognostic factor for Korean patients with CLL. Whereas traditional prognostic stratification systems such as the Rai and Binet systems cannot reflect tumor burdens, hyperleukocytosis ($\geq 100 \times 10^3/\mu\text{L}$) as well as the proportion of neoplastic cells in the bone marrow ($\geq 80\%$), and higher LDH (≥ 500 IU/L) levels, which might reflect tumor burden, were also important factors affecting treatment outcomes in Korean patients with CLL, as shown in our univariate analysis of the survival rates. In addition, extranodal involvement was a poor prognostic factor, although extranodal involvement was only found in 4.2% of the patients in our study. The extranodal sites involved were pleura, cerebrospinal fluid (CSF), brain, parotid gland, and nasopharynx.

We also found that mild thrombocytopenia (platelet count between $80 \times 10^3/\mu\text{L}$ and $110 \times 10^3/\mu\text{L}$) did not have a poor prognostic significance in our data. The survival of patients with platelet counts between $80 \times 10^3/\mu\text{L}$ and $110 \times 10^3/\mu\text{L}$ was not different from that of patients with a platelet count $> 110 \times 10^3/\mu\text{L}$. Traditionally, according to the Rai and Binet staging systems, patients with thrombocytopenia (platelet count: $< 100 \times 10^3/\mu\text{L}$, irrespective of organomegaly) were categorized into the worst prognostic group, stage IV and stage C, respectively [3]. In contrast to the results of previous studies using

clinical staging systems such as Rai and Binet systems, the German CLL Study Group (GCLLSG) data, or the CLL international prognostic index (CLL-IPI) [3,4,6,22], our data showed that a platelet count $< 80 \times 10^3/\mu\text{L}$ had poor prognostic significance. Therefore, we suggest that thrombocytopenia needs to be more stratified in Korean patients with CLL. If patients have a platelet count $> 80 \times 10^3/\mu\text{L}$ without any other therapeutic indications for CLL such as progressive organomegaly, severe anemia, and constitutional symptoms, a more conservative approach might be acceptable, and chemotherapy might be deferred for these patients.

Our data have several limitations. This study is retrospective in nature, included a small number of patients, and had limited FISH data. Our study population had limited exposure to newer therapeutic agents such as rituximab. However, our results showed the unique clinical characteristics and implications of CLL among Korean patients, including its clinical factors and immunophenotypic features and more stratified additional prognostic factors.

Although a diagnosis of CLL requires the presence of $\geq 5 \times 10^9/\text{L}$ B lymphocytes in the peripheral blood, sustained for at least 3 months [27], some patients with CLL in the present study have less than $5 \times 10^9/\text{L}$ B lymphocytes. According to the 2008 International Workshop on Chronic Lymphocytic Leukemia, the presence of a cytopenia caused by a typical marrow infiltrate establishes the diagnosis of CLL regardless of the number of peripheral blood B lymphocytes, or the presence of lymph node involvement [27].

To the best of our knowledge, this is the first multicenter retrospective study to analyze the treatment outcomes and prognostic factors of Korean patients with CLL who received first-line chemotherapy. Considering the rarity of CLL in Asia, our sample size is not really small.

In conclusion, Korean patients with CLL have different characteristics in terms of the frequency of lambda SmIg expression and atypical CD22 and FMC7 marker positivity. Hyperleukocytosis, extranodal involvement, Binet stage C, a complex karyotype, del(17) and del(11q) were independent significant negative predictors. We suggest that our results might be incorporated in a better risk stratification system of CLL in the future.

KEY MESSAGE

1. Korean patients with, chronic lymphocytic leukemia (CLL) had higher lambda surface membrane immunoglobulin (SmIg) expression and atypical CD22 and FMC7 marker positivity than Western patients.
2. Hyperleukocytosis, extranodal involvement, and complex karyotype as well as del(17) and del(11q) and traditional Rai/Binet system are important adverse prognostic factors for CLL among Korean patients.
3. Mild thrombocytopenia ($80,000/\text{mm}^3 \leq$ platelet count $< 110,000/\text{mm}^3$) had no negative effects on the treatment outcomes.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

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REFERENCES

1. Matutes E, Owusu-Ankomah K, Morilla R, et al. The immunological profile of B-cell disorders and proposal of a scoring system for the diagnosis of CLL. *Leukemia* 1994;8:1640-1645.
2. Hallek M, Cheson BD, Catovsky D, et al. Guidelines for the diagnosis and treatment of chronic lymphocytic leukemia: a report from the International Workshop on Chronic Lymphocytic Leukemia updating the National Cancer Institute-Working Group 1996 guidelines. *Blood* 2008;111:5446-5456.
3. Binet JL, Auquier A, Dighiero G, et al. A new prognostic classification of chronic lymphocytic leukemia derived from a multivariate survival analysis. *Cancer* 1981;48:198-206.
4. Rai KR, Sawitsky A, Cronkite EP, Chanana AD, Levy RN, Pasternack BS. Clinical staging of chronic lymphocytic leukemia. *Blood* 1975;46:219-234.
5. Hamblin TJ, Davis Z, Gardiner A, Oscier DG, Stevenson FK. Unmutated Ig V(H) genes are associated with a more aggressive form of chronic lymphocytic leukemia. *Blood* 1999;94:1848-1854.
6. Pflug N, Bahlo J, Shanafelt TD, et al. Development of a comprehensive prognostic index for patients with chronic lymphocytic leukemia. *Blood* 2014;124:49-62.
7. Dohner H, Stilgenbauer S, Benner A, et al. Genomic aberrations and survival in chronic lymphocytic leukemia. *N Engl J Med* 2000;343:1910-1916.
8. Jeon YW, Cho SG. Chronic lymphocytic leukemia: a clinical review including Korean cohorts. *Korean J Intern Med* 2016;31:433-443.
9. Yang SM, Li JY, Gale RP, Huang XJ. The mystery of chronic lymphocytic leukemia (CLL): why is it absent in Asians and what does this tell us about etiology, pathogenesis and biology? *Blood Rev* 2015;29:205-213.
10. Yoon SO, Suh C, Lee DH, et al. Distribution of lymphoid neoplasms in the Republic of Korea: analysis of 5318 cases according to the World Health Organization classification. *Am J Hematol* 2010;85:760-764.
11. Chihara D, Ito H, Matsuda T, et al. Differences in incidence and trends of haematological malignancies in Japan and the United States. *Br J Haematol* 2014;164:536-545.
12. Jang MA, Yoo EH, Kim K, et al. Chronic lymphocytic leukemia in Korean patients: frequent atypical immunophenotype and relatively aggressive clinical behavior. *Int J Hematol* 2013;97:403-408.
13. Lee H, Park HJ, Park EH, et al. Nationwide statistical analysis of lymphoid malignancies in Korea. *Cancer Res Treat* 2018;50:222-238.
14. Gunawardana C, Austen B, Powell JE, et al. South Asian chronic lymphocytic leukaemia patients have more rapid disease progression in comparison to White patients. *Br J Haematol* 2008;142:606-609.
15. Agrawal N, Naithani R, Mahapatra M, et al. Chronic lymphocytic leukemia in India: a clinico-hematological profile. *Hematology* 2007;12:229-233.
16. Eichhorst B, Fink AM, Bahlo J, et al. First-line chemotherapy with bendamustine and rituximab versus fludarabine, cyclophosphamide, and rituximab in patients with advanced chronic lymphocytic leukaemia (CLL10): an international, open-label, randomised, phase

- 3, non-inferiority trial. *Lancet Oncol* 2016;17:928-942.
17. Woyach JA, Johnson AJ, Byrd JC. Chronic lymphocytic leukemia: recent progress and current challenges. *Semin Oncol* 2016;43:199-200.
18. Walter HS, Salles GA, Dyer MJ. New agents to treat chronic lymphocytic leukemia. *N Engl J Med* 2016;374:2185-2186.
19. Vela CM, McBride A, Jaglowski SM, Andritsos LA. Ibrutinib for treatment of chronic lymphocytic leukemia. *Am J Health Syst Pharm* 2016;73:367-375.
20. Zenz T, Eichhorst B, Busch R, et al. TP53 mutation and survival in chronic lymphocytic leukemia. *J Clin Oncol* 2010;28:4473-4479.
21. Rossi D, Cerri M, Deambrogi C, et al. The prognostic value of TP53 mutations in chronic lymphocytic leukemia is independent of Del17p13: implications for overall survival and chemorefractoriness. *Clin Cancer Res* 2009;15:995-1004.
22. International CLL-IPI working group. An international prognostic index for patients with chronic lymphocytic leukaemia (CLL-IPI): a meta-analysis of individual patient data. *Lancet Oncol* 2016;17:779-790.
23. Le Bris Y, Struski S, Guieze R, et al. Major prognostic value of complex karyotype in addition to TP53 and IGHV mutational status in first-line chronic lymphocytic leukemia. *Hematol Oncol* 2017;35:664-670.
24. Rigolin GM, Saccenti E, Guardalben E, et al. In chronic lymphocytic leukaemia with complex karyotype, major structural abnormalities identify a subset of patients with inferior outcome and distinct biological characteristics. *Br J Haematol* 2018;181:229-233.
25. Herling CD, Klaumunzer M, Rocha CK, et al. Complex karyotypes and KRAS and POT1 mutations impact outcome in CLL after chlorambucil-based chemotherapy or chemoimmunotherapy. *Blood* 2016;128:395-404.
26. Wierda WG, O'Brien S, Wang X, et al. Prognostic nomogram and index for overall survival in previously untreated patients with chronic lymphocytic leukemia. *Blood* 2007;109:4679-4685.
27. Hallek M, Cheson BD, Catovsky D, et al. iwCLL guidelines for diagnosis, indications for treatment, response assessment, and supportive management of CLL. *Blood* 2018;131:2745-2760.

Supplementary Table 1. Clinical characteristics at the time of chemotherapy and treatment outcomes

Characteristic	Total (n = 90)
Stage (Rai system)	
o	23 (25.6)
I	13 (14.4)
II	14 (15.6)
III	20 (22.2)
IV	19 (21.1)
Unknown	1 (1.1)
Stage (Binet system)	
A	45 (50.0)
B	11 (12.2)
C	33 (36.7)
Unknown	1 (1.1)
Constitutional symptom (n = 88)	12 (13.3)
Response to 1st line chemotherapy	
CR	16 (17.8)
PR	47 (52.2)
SD	20 (22.2)
PD	4 (4.4)
Unknown	2 (2.2)
Death in cytopenia	1 (1.1)

Values are presented as number (%).

CR, complete remission; PR, partial remission; SD, stable disease; PD, progressive disease.