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Visceral adiposity is associated
with altered myocardial glucose uptake
measured by ^{18}F FDG-PET in 346 subjects
with normal glucose tolerance,
prediabetes, and type 2 diabetes

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Directed by Professor Mijin Yun

The Master's Thesis
submitted to the Department of Medicine
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in partial fulfillment of the requirements for the degree of
Master of Medical Science

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ABSTRACT

Visceral adiposity is associated with altered myocardial glucose uptake measured by 18 FDG-PET in 346 subjects with normal glucose tolerance, prediabetes, and type 2 diabetes

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Purpose: To evaluate the glucose uptake in hearts among subjects with normoglycemia, impaired fasting glucose (IFG) and type 2 diabetes mellitus (T2D) using [¹⁸F] fluorodeoxyglucose- positron emission tomography (¹⁸FDG-PET) and investigate its association with various clinical and laboratory profiles.

Methods and results: In a total of 387 participants (control, N=75; IFG, N=242; T2D, N=70), enrolled from the health promotion center at university-affiliated tertiary-care hospital, we assessed the myocardial glucose uptake by ¹⁸FDG-PET. Visceral and subcutaneous fat areas were evaluated by abdominal computed tomography (CT) and laboratory parameters including glucose, insulin, uric acid, free fatty acids (FFA), and triglycerides were measured. Myocardial glucose uptake was significantly decreased in subjects with T2D compared to control or IFG groups (P for trend=0.001). Individuals in the lowest tertile of myocardial glucose uptake tended to be having T2D (P=0.002). Correlation analysis showed that myocardial glucose uptake was related to body mass index, visceral and subcutaneous fat areas, HOMA-IR, uric acid and fasting/postprandial serum levels of glucose, insulin, triglycerides, and FFA. In a multiple linear regression model, visceral fat area (Standardized β [STD β]=-0.31, P<0.001), fasting FFA (STD β =-0.34, P<0.001), and uric acid levels (STD β =-0.21, P=0.007) were independent determinants of

myocardial glucose uptake. Multiple logistic analyses demonstrated that decreased myocardial glucose uptake was associated with the risk of diabetes (odds ratio= 2.56; 95% confidence interval: 1.15-5.70, P=0.021).

Conclusion: Myocardial glucose uptake evaluated by ^{18}F FDG-PET may predict T2D and was significantly associated with visceral adiposity and fatty acid or uric acid metabolism.

Key words : Myocardial metabolism, Type 2 diabetes mellitus, Positron emission tomography, Fluorodeoxyglucose

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I. INTRODUCTION

Type 2 diabetes (T2D) is a chronic burgeoning disorder worldwide characterized by hyperglycemia, insulin resistance, and increased lipid and lipoprotein abnormalities^{1,2}. Previous studies have shown that dysfunctional adiposity such as excess visceral fat is independently associated with risks of prediabetes and T2D³. T2D also has effects on long-term dysfunction and damage in various organs including eye, kidney, and heart⁴. The heart requires constant sources of energy including mainly free fatty acids (FFA) and glucose for continuous pumping and has known to be highly flexible in energy substrate metabolism^{5,6}. In T2D heart, alterations in myocardial substrate metabolism characterized by increased myocardial fatty acid metabolism and concurrently decreased glucose metabolism have been demonstrated⁶⁻⁹.

Positron emission tomographic (PET) represents increased uptakes of [¹⁸F] fluorodeoxyglucose (¹⁸FDG), a glucose analog, which is taken up by tissues via glucose transporters^{10,11}. Recent studies have reported that the disturbances of carbohydrate, fat, and protein metabolism altered biodistribution of FDG in patients with T2D^{12,13}. In addition, myocardial glucose uptake may differ according to the

status of whole-body insulin resistance such as impaired fasting glucose (IFG) and T2D, compared with normoglycemic control, as myocardial insulin sensitivity. Yet little research has been performed in relationship with myocardial glucose uptake using PET, adiposity, and other metabolic profiles in subjects with normoglycemia, IFG, and T2D.

Therefore, in the current study, we aimed to investigate myocardial glucose uptake by ^{18}F FDG-PET and its association with various clinical and laboratory parameters according to glycemic status.

II. MATERIALS AND METHODS

1. Patient Population

Between Jan 2008 and July 2014, a total of 387 individuals aged over 25 years enrolled from the health promotion center at university-affiliated tertiary-care hospital. We studied 75 healthy control subjects, 242 subjects with IFG, and 70 subjects with T2D defined by the 2011 revision of the American Diabetes Association (ADA) guidelines¹⁴. Patients with cardiovascular disease, previous or present abnormal renal or hepatic function, and other endocrine disorders were excluded.

2. Clinical and laboratory study and abdominal computed tomography

Body mass index (BMI) was defined as weight divided by the square of the height (kg/m^2). Blood pressure was obtained by averaging three times of blood pressure records in the sitting position after at least 5 minutes of rest. Blood samples were collected from each participant after overnight fasts. The fasting and 2-hour postprandial glucose, insulin, FFA, and triglycerides after a conventional meal were measured. Plasma glucose was measured using the glucose oxidase method. Plasma total cholesterol, triglyceride, high density lipoprotein cholesterol, free fatty acid, lipoprotein(a), and uric acid were assayed using a Hitachi 7600 auto analyzer (Hit-

achi Instruments Service, Tokyo, Japan). Low density lipoprotein cholesterol was calculated using the Friedewald equation [LDL-C (mg/dL) = TC (mg/dL) – HDL-C (mg/dL) – TG (mg/dL)/5]. Serum glycated albumin was determined by an enzymatic method as previously described¹⁵. HbA_{1c} was measured by high-performance liquid chromatography using VariantTM II Turbo (Bio-Rad Laboratories, Hercules, CA, USA). The reference ranges for HbA_{1c} was 4.0% to 6.0%, and for glycated albumin was 11.0% to 16.0%. Homeostasis Model Assessment of Insulin Resistance (HOMA-IR) was calculated using the following formula: [(Fasting plasma insulin (μU/mL) * Fasting plasma glucose (mg/dL))/405]¹⁶. An index of adipose tissue insulin resistance (Adipo-IR) was calculated as [Fasting plasma FFA (mmol/L) * Fasting plasma insulin (pmol/L)]^{17,18}.

The abdominal adipose tissue areas were quantified by computed tomography (CT) (Tomoscan 350; Philips, Mahwah, NJ, USA). Total abdominal and visceral adipose tissue (VAT) area were measured at the L3-L4s level by a 10 mm CT slice scan in a supine position. VAT area was measured as intra-abdominal cavity at the internal aspect of the abdominal and oblique muscle walls and the posterior aspect of the vertebral body. The subcutaneous adipose tissue (SAT) area was acquired from the total adipose tissue area by subtracting the VAT area.

3. ¹⁸FDG-Positron Emission Tomography and image analysis

All patients fasted for more than six hours prior to the ¹⁸FDG-PET/CT scan. 5.5 MBq/kg of body weight of ¹⁸F-FDG were intravenously injected over 2 min. Serum glucose levels at the time of FDG injection did not exceed 150 mg/dL. After a 1-hour equilibration period, a low-dose CT scan for attenuation correction was performed, followed by acquisition of ¹⁸FDG-PET images from the skull base to the mid-thigh in the 3D mode at 3 min per bed position using a standard PET-CT scanner (Gemini, Philips Medical Systems or Discovery STe, GE Healthcare). PET images were reconstructed with attenuation correction. Two-dimensional regions of interest (ROI) were drawn the transaxial images to measure the standardized uptake

value (SUV) of the entire left ventricular myocardium within an inner edge; $SUV = (\text{peak kBq/mL in ROI}) / (\text{injected activity/g body weight})$. Myocardial ^{18}F FDG uptake was expressed as the maximal SUV (SUV_{max}). We also obtained the SUV of liver from the circular ROI in the central area of the right lobe. Liver ^{18}F FDG uptake was determined by averaging these values. The values of SUV of the heart to liver ^{18}F FDG uptake ratio (Heart SUV/ Liver SUV [$SUV_{\text{Heart/Liver}}$]) values were used to estimate myocardial glucose uptake.

4. Statistical Analysis

All continuous variables were presented as mean \pm standard deviation (SD) and categorical variables were presented as proportions. Differences were analysed using analysis of variance (ANOVA) for continuous variables and Chi-square test for categorical variables. Comparisons of myocardial glucose uptake across the status of diabetes were calculated with the Jonckheere- Terpstra trend test. Pearson's correlation coefficients were calculated to examine the relationships between myocardial glucose uptake and metabolic variables. Multiple linear regression analysis was performed to determine the independent relationship of the studied variables and Standardized β was represented as the coefficient β . The odds ratios (ORs) and 95% confidence intervals (CIs) for the factors associated with the risk of T2D were calculated using multivariate logistic regression analysis. In the Pearson's correlation, multiple linear regression, and logistic regression analysis, myocardial glucose uptake ($SUV_{\text{Heart/Liver}}$) was log-transformed. The receiver operating characteristic (ROC) curve of myocardial glucose uptake was used to assess the optimal cut-off value for the prediction of T2D with the areas under the curve (AUC). A $P < 0.05$ was considered statistically significant. Statistical analyses were performed using PASW Statistics version 20.0 for Windows (SPSS Inc., Chicago, IL, USA).

III. RESULTS

1. Baseline Characteristics of Study Population

Table 1. Characteristics of the study subjects

	NGT (N=75)	IFG (N=242)	T2D (N=70)	P
Age yrs	50.4 ± 10.6	58.8 ± 9.9	63.2 ± 10.7	< 0.001
Female n (%)	37 (49.3)	109 (45.0)	21 (30.0)	0.031
SBP (mmHg)	116.7 ± 11.9	120.8 ± 14.0	127.7 ± 14.6	< 0.001
DBP (mmHg)	74.2 ± 9.7	76.0 ± 10.2	78.0 ± 11.3	0.112
BMI (kg/m ²)	23.2 ± 3.1	24.2 ± 3.0	25.8 ± 3.9	< 0.001
Visceral fat area (cm ²)	104.3 ± 46.1	129.9 ± 59.6	195.7 ± 91.8	< 0.001
Subcutaneous fat area (cm ²)	140.9 ± 37.8	142.4 ± 56.7	157.8 ± 76.5	0.361
HbA _{1c} (%)	5.5 ± 0.2	5.8 ± 0.2	7.1 ± 1.2	< 0.001
Glycated albumin (%)	11.0 ± 1.2	11.8 ± 1.8	16.4 ± 6.3	< 0.001
Fasting glucose (mg/dL)	90.2 ± 8.1	97.0 ± 9.9	121.3 ± 35.5	< 0.001
Postprandial glucose (mg/dL)	98.2 ± 11.5	123.6 ± 36.9	214.0 ± 63.5	< 0.001
Fasting insulin (μU/mL)	5.7 ± 3.3	7.0 ± 4.2	9.4 ± 7.4	0.001
Postprandial insulin (μU/mL)	20.9 ± 17.4	38.5 ± 37.5	55.9 ± 51.7	< 0.001
HOMA-IR	1.3 ± 0.8	1.7 ± 1.0	2.9 ± 2.4	< 0.001
Adipo-IR	26.9 ± 20.4	30.3 ± 23.1	40.2 ± 36.3	0.083

Total cholesterol (mg/dL)	190.8 ± 36.4	186.9 ± 39.3	162.5 ± 42.0	0.029
HDL cholesterol (mg/dL)	49.8 ± 11.8	49.3 ± 12.4	43.5 ± 10.60	0.001
LDL cholesterol (mg/dL)	113.6 ± 33.7	111.3 ± 34.4	94.1 ± 32.1	< 0.001
Fasting TG (mg/dL)	105.0 ± 71.2	114.4 ± 58.2	132.4 ± 70.3	0.029
Postprandial TG (mg/dL)	96.4 ± 73.9	104.9 ± 50.0	131.3 ± 80.0	0.051
Fasting FFA (μ Eq/L)	683.7 ± 381.1	626.1 ± 242.0	652.2 ± 286.3	0.569
Postprandial FFA (μ Eq/L)	173.2 ± 183.0	135.9 ± 106.0	196.3 ± 162.3	0.055
Lipoprotein(a) (mg/dL)	21.6 ± 24.2	23.9 ± 25.6	29.1 ± 33.8	0.246
Uric acid (mg/dL)	5.0 ± 1.4	5.5 ± 1.3	5.5 ± 1.0	0.020
Statin users	5 (6.8)	48 (19.8)	32 (45.7)	< 0.001

Data were shown as mean ± standard deviation (SD) or n(%).

NGT, Normal glucose tolerance; IFG, Impaired fasting glucose; T2D, Type 2 diabetes; SBP, Systolic blood pressure; DBP, Diastolic blood pressure; BMI, Body mass index; HOMA-IR, Homeostasis model assessment of insulin resistance; Adipo-IR, An index of adipose tissue insulin resistance; HDL, High density lipoprotein; LDL, Low density lipoprotein; TG, Triglycerides; FFA, Free fatty acid

For all subjects, the mean age of patients was 58.9 ± 10.9 years, 43.2 % were women, and the average BMI was 24.2 ± 3.0 kg/m². Among T2D patients, oral hypoglycemic agent users and insulin users were 56.9 % and 10.0 %, respectively. The proportion of subjects with dipeptidyl peptidase-4 inhibitors was 32.8 %, subjects with thiazolidinedione was 13.8 %, subjects with metformin was 46.6 %, and subjects with sulfonylurea was 22.4 % in subjects using oral hypoglycemic agents. Subjects with IFG (N=242) or T2D (N=70) tended to be older, to have higher systolic blood pressure, more obese, and to have metabolically unhealthy factors compared to subjects with normal glucose tolerance (NGT; N=75). Subjects

with IFG or T2D were more likely to have higher levels of uric acid and larger visceral fat area compared to subjects with NGT.

2. Myocardial glucose uptake in NGT, IFG, and T2D

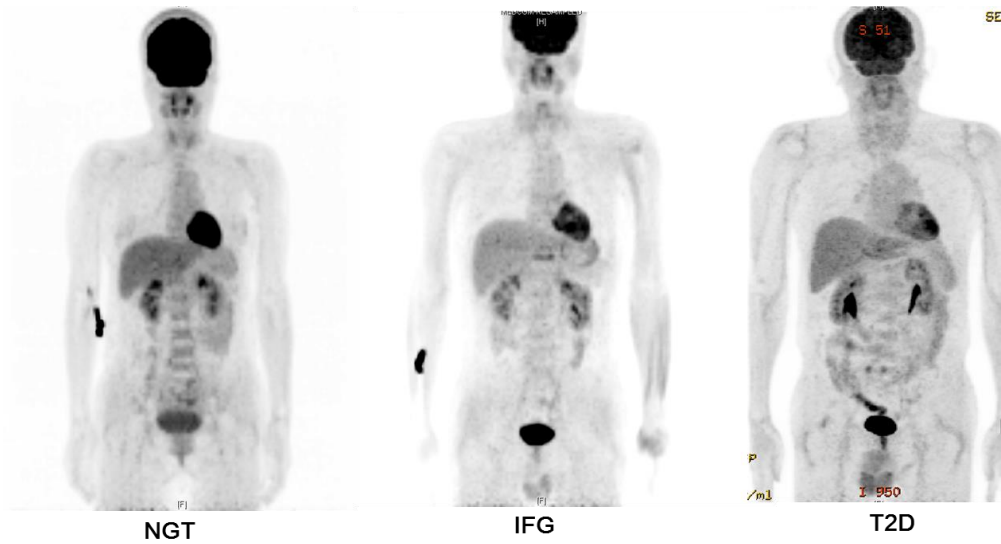


Figure 1. Altered myocardial glucose uptake of FDG in subjects by diabetic status. Each subject has median value within the highest tertile of myocardial glucose uptake in NGT, IFG, and T2D.

Figure 1 presents the status of myocardial glucose uptake in subjects by glycemic status. Compared to NGT or IFG groups, myocardial glucose uptake significantly tended to be decreased in subjects with T2D (NGT, IFG, T2D; mean $SUV_{\text{Heart/Liver}}$ = 1.89, 1.60, 1.12, respectively; P for trend=0.001, Figure 1, 2). The proportion of lowest tertile of myocardial glucose uptake was significantly higher in patients with T2D ($P=0.002$).

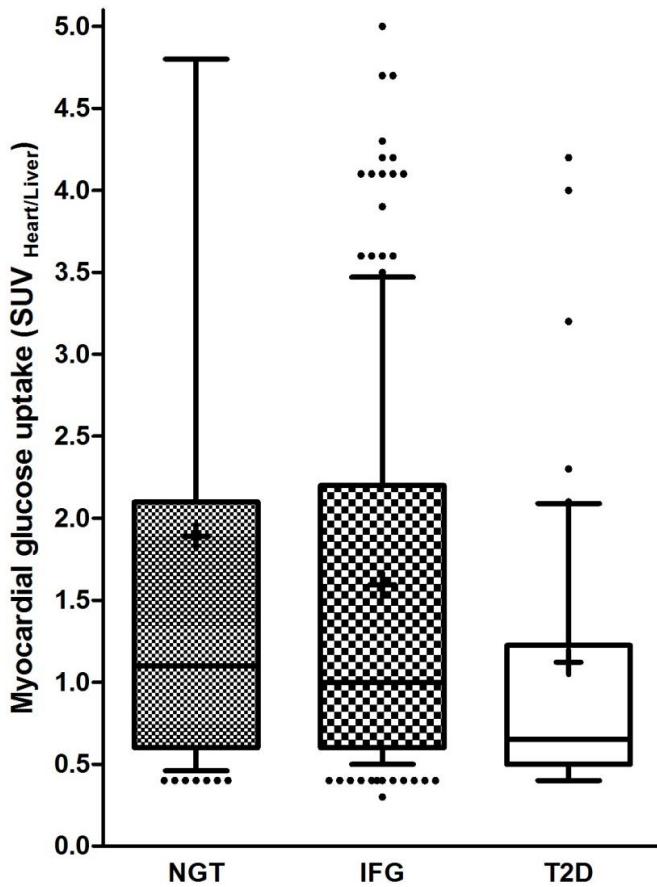


Figure 2. Myocardial glucose uptake by glycemic status Each + and horizontal line in the box indicate mean and median value of myocardial glucose uptake. Bar indicates 10 to 90 percentiles.

3. Relationship between myocardial glucose uptake and metabolic measures

Table 2. Correlation between myocardial glucose uptake and metabolic factors

	r	P
Age yrs	-0.02	0.757
SBP mmHg	-0.03	0.540
DBP mmHg	-0.03	0.571

BMI kg/m^2	-0.18	< 0.001
Visceral fat area cm^2	-0.24	< 0.001
Subcutaneous fat area cm^2	-0.16	< 0.001
HbA _{1c} %	-0.14	0.005
Glycated albumin %	-0.03	0.601
Fasting glucose mg/dL	-0.18	< 0.001
Postprandial glucose mg/dL	-0.13	0.013
Fasting insulin $\mu U/mL$	-0.12	0.035
Postprandial insulin $\mu U/mL$	-0.12	0.042
HOMA-IR	-0.15	0.006
Adipo-IR	-0.32	< 0.001
Total cholesterol mg/dL	0.00	0.995
HDL cholesterol mg/dL	0.02	0.776
LDL cholesterol mg/dL	0.04	0.499
Fasting triglycerides mg/dL	-0.13	0.010
Postprandial triglycerides mg/dL	-0.22	0.003
Fasting FFA $\mu Eq/L$	-0.30	< 0.001
Postprandial FFA $\mu Eq/L$	-0.16	0.031
Lipoprotein(a) mg/dL	0.04	0.309

SBP, Systolic blood pressure; DBP, Diastolic blood pressure; BMI, Body mass index; HOMA-IR, Homeostasis model assessment of insulin resistance; Adipo-IR, An index of adipose tissue insulin resistance; HDL, High density lipoprotein; LDL, Low density lipoprotein; FFA, Free fatty acids

To assess the relationship between myocardial glucose uptake and metabolic measures, univariate analysis was performed in Table 2. We observed myocardial glucose uptake correlated negatively with BMI, HbA_{1c}, fasting/postprandial levels of glucose, insulin, TG, HOMA-IR, Adipo-IR, and uric acid. Moreover, statistically significant inverse relations were found between myocardial glucose uptake and both visceral fat areas ($r=-0.24$, $P<0.001$), subcutaneous fat areas ($r=-0.16$, $P<0.001$), and fasting FFA ($r=-0.30$, $P<0.001$).

Table 3. Analysis of myocardial glucose uptake in association with metabolic parameters

	Model 1		Model 2		Model 3	
R ²	0.066		0.250		0.277	
	β	P	β	P	β	P
Visceral fat area <i>cm</i> ²	-0.30	<	-	-	-0.31	<
		0.001				0.001
Fasting FFA μ Eq/L	-	-	-0.34	<	-0.34	<
				0.001		0.001
Uric acid <i>mg/dL</i>	-	-	-0.26	<	-0.21	0.007
				0.001		
Postprandial glucose <i>mg/dL</i>	-	-	-0.15	0.031	-	-
Postprandial triglycerides <i>mg/dL</i>	-	-	-0.21	0.002	-	-

Model 1: adjusted for age, sex, BMI, and subcutaneous fat area; Model 2: adjusted for age, sex, fasting/postprandial glucose, insulin, C-peptide, FFA, triglycerids, HbA_{1c}, glycoalbumin, total cholesterol, HDL cholesterol, LDL cholesterol, lipoprotein(a), HOMA-IR, Adipo-IR, uric acid; Model 3: adjusted for age, sex, and variables derived from model 1 and 2.

Multiple linear regression analysis was performed to determine whether myocardial glucose uptake was independently associated with various factors of body composition in addition with age, sex (Model 1), laboratory parameters in addition with age, sex (Model 2), and variables derived from model 1 and model 2 (Model 3; Table 3). In model 3, we found that visceral fat area (β =-0.31, P <0.001), fasting FFA (β =-0.34, P <0.001), and uric acid levels (β =-0.21, P =0.007) were remained to be as independent determinants of myocardial glucose uptake after adjustment for effective body composition and laboratory parameters from model 1 and model 2.

4. Assessment of diabetes risk in an association with myocardial glucose uptake

We further investigated significant determinants for the risk of diabetes by multiple logistic regression analyses. After sequential adjustment for confounding covariates, subjects with diabetes had significantly elevated odds ratios (ORs) for having decreased myocardial glucose uptake (Model 5, OR=2.56, P=0.021) and increased visceral fat area (1.02, P=0.008). The optimal cut-off value of myocardial glucose uptake for predicting diabetes was 0.726 (AUC 0.644, [95% CI]=[0.575-0.713]) with sensitivity of 62.8 % and specificity of 67.1 % (P=0.035, Figure 2).

IV. DISCUSSION

In the present study, we used ¹⁸FDG-PET and abdominal CT to examine myocardial glucose uptake and visceral/ subcutaneous adipose tissue area, respectively in subjects with normoglycemia, IFG, and T2D. Our results demonstrated that myocardial glucose uptake was markedly decreased by 40.7 %, while area of visceral fat was increased by 87.6 % in patients in T2D compared with normoglycemic control. Myocardial glucose uptake was negatively associated with levels of plasma FFA, uric acid, and visceral fat area. Furthermore, alteration of myocardial glucose uptake evaluated by ¹⁸FDG-PET can be a risk factor of T2D.

Previously, hyperglycemia, hyperinsulinemia, and disturbances of carbohydrates, fatty acids, and protein metabolism all have been demonstrated an association that has been found in prediabetes and T2D^{2,4}. Therefore, impaired glucose uptake and metabolism in not only skeletal muscle but also in heart, which consistently requires sources of energy from mainly FFA and glucose, could have a correlation with insulin resistance in the status of prediabetes and T2D, contributing to the development of hyperglycemia¹⁹. Moreover, visceral fat accumulation has been shown to correlate with insulin resistance in subjects with obesity and T2D^{20,21}. These findings are in line with the results of the current study that myocardial glucose uptake was significantly decreased, on the other hand, visceral adiposity

was increased with elevated levels of plasma FFA in patients with T2D.

Consistent with previous studies, we revealed that fasting FFA presented to be an independent predictor for myocardial glucose uptake^{22,23}. There have been conflicting findings of the uncertain relation between insulin resistance and myocardial glucose uptake in diabetic patients because relationship between a direct effect of myocardial insulin resistance on myocardial glucose uptake and independence of increased plasma FFA is not clear^{12,23-26}. For instance, there were reports that suppressed serum FFA concentrations by acipimox, a potent nicotinic acid derivative, effected the glucose uptake in myocardium²³, and decrease in the FFA levels by rosiglitazone therapy was associated with the improvement in myocardial glucose uptake²⁷. However, Yokoyama et al. showed whole-body glucose disposal rate (GDR) was independently related to myocardial FDG uptake, whereas FFA was not²⁵. Similar to this, Hicks et al. reported that relation between myocardial FDG uptake and GDR was significant than between myocardial FDG uptake and FFA in subjects with diabetes²⁸. Our study revealed myocardial glucose uptake independently related with not subcutaneous, but visceral fat tissue area, uric acid metabolism as well as FFA in multiple linear regression analysis. Various studies reported that increased visceral adipose tissue, not abdominal subcutaneous adipose tissue expected increased in insulin resistance²⁹⁻³¹. Moreover, visceral fat accumulation turned out to be closely related to the increased production of uric acid which may be linked to increased TG synthesis by liver in obese subjects³². Also, an increased serum uric acid was found to be independently associated with insulin resistance and increased risk factors of metabolic syndrome even in the normal subjects³³. Relation between serum uric acid concentration and insulin resistance seemed to be appeared by decreased urinary uric acid clearance but further study of the exact mechanism would be warranted³⁴. Of note, we found that myocardial glucose uptake showed a gradual decrease in proportion in subjects with insulin resistant IFG and T2D. Collectively, myocardial glucose uptake appears to be affected by insulin resistance in an association with visceral adiposity, uric acid

metabolism, and increased FFA itself.

Previous studies showed that in the normal heart, under fasting conditions, FDG uptake image showed variable myocardial glucose uptake because FFA is primary fuel whereas glucose utilization is relatively low for myocardial oxidative metabolism compared to glucose-loading conditions³⁵⁻³⁷. However, in T2D heart, regulation of glucose metabolism is different from normal heart, so that prior studies showed that myocardial FDG uptake was significantly decreased in diabetic patients compared to normal subjects consistent with our results^{13,35}. Cardiac myocytes utilize glucose via mostly insulin-sensitive glucose transporters (GLUT4), responsible for more than 50 % of all body glucose uptake³⁸. Reduced expression and mutations of GLUT4 have been associated with diabetes^{39,40}. Therefore, the probable mechanisms of decreased myocardial glucose uptake seem to be related with the presence of insulin resistance in patients with T2D¹³. Also, recent studies reported that influences of glucagon-like peptide-1 (GLP-1) caused improvement on myocardial glucose metabolism in diabetic patients^{41,42}. Several studies have reviewed that chronic shift of myocardial substrate preference has been noted in the diabetic heart, resulting in prominent decreased glucose and lactic acid oxidation and increased fatty acid oxidation⁶⁻⁹. Effects of diabetes on myocardial metabolism are very complex with systemic metabolic disturbances of hyperglycemia, increased FFA and down-regulation of glucose transporters, increased insufficient energy utilization of fatty acid oxidation, lipid accumulation, and lipotoxicity in cardiomyocyte^{6,43,44}. Accordingly, these may lead to diabetic cardiomyopathy^{43,45}. Further investigation in relationship between myocardial glucose uptake and cardiac function will be needed.

The present study has several distinguishable strengths. It is the first study to investigate myocardial glucose intake evaluated using ¹⁸FDG-PET accompanied with subcutaneous/ visceral adiposity by abdominal CT and other metabolic parameters in the point of view of insulin sensitivity according to glycemic status. It can be speculated that myocardial glucose uptake was decreased with insulin

resistance, demonstrating visceral adiposity, hyperuricemia, and elevated FFA in patients with T2D. ^{18}F FDG-PET is recently widely performed to detect of malignant tumors or ischemic heart disease for medical examination. Based on these results, furthermore, myocardial glucose uptake assessed by ^{18}F FDG-PET may be a useful tool to consider the risk for T2D.

The current study has several limitations. First, this cross-sectional study design is insufficient to lead the causal relationship in the development of impaired myocardial glucose uptake. Another limitation is that in the present study, we did not assess cardiac function in our patients. Correlation between myocardial glucose uptake and cardiac function would be an interesting topic of investigation.

V. CONCLUSION

In conclusion, we confirmed that myocardial glucose uptake was decreased in patients with T2D and there was an association with increased levels of free fatty acids, uric acid, and visceral adiposity. Importantly, alteration of myocardial glucose uptake evaluated by ^{18}F FDG-PET can be a risk factor of T2D. Additional studies would be needed to confirm the relationship of myocardial glucose metabolism on the development of diabetes and cardiac function.

REFERENCES

1. Smyth S, Heron A. Diabetes and obesity: the twin epidemics. *Nat Med* 2006;12:75-80.
2. DeFronzo RA. Pathogenesis of type 2 diabetes mellitus. *Med Clin North Am* 2004;88:787-835, ix.
3. Neeland IJ, Turer AT, Ayers CR, Powell-Wiley TM, Vega GL, Farzaneh-Far R, Grundy SM, Khera A, McGuire DK, de Lemos JA. Dysfunctional adiposity and the risk of prediabetes and type 2 diabetes in obese adults. *Jama* 2012;308:1150-1159.
4. Nolan CJ, Damm P, Prentki M. Type 2 diabetes across generations: from pathophysiology to prevention and management. *Lancet* 2011;378:169-181.
5. Neely JR, Rovetto MJ, Oram JF. Myocardial utilization of carbohydrate and lipids. *Prog Cardiovasc Dis* 1972;15:289-329.
6. van den Brom CE, Bulte CS, Loer SA, Bouwman RA, Boer C. Diabetes, perioperative ischaemia and volatile anaesthetics: consequences of derangements in myocardial substrate metabolism. *Cardiovasc Diabetol* 2013;12:42.
7. Rijzewijk LJ, van der Meer RW, Lamb HJ, de Jong HW, Lubberink M, Romijn JA, Bax JJ, de Roos A, Twisk JW, Heine RJ, Lammertsma AA, Smit JW, Diamant M. Altered myocardial substrate metabolism and decreased diastolic function in nonischemic human diabetic cardiomyopathy: studies with cardiac positron emission tomography and magnetic resonance imaging. *J Am Coll Cardiol* 2009;54:1524-1532.
8. Carley AN, Severson DL. Fatty acid metabolism is enhanced in type 2 diabetic hearts. *Biochim Biophys Acta* 2005;1734:112-126.
9. Stanley WC, Lopaschuk GD, McCormack JG. Regulation of energy substrate metabolism in the diabetic heart. *Cardiovasc Res* 1997;34:25-33.
10. Thorens B, Mueckler M. Glucose transporters in the 21st Century. *Am J Physiol Endocrinol Metab* 2010;298:E141-145.
11. Boellaard R, O'Doherty MJ, Weber WA, Mottaghy FM, Lonsdale MN, Stroobants SG, Oyen WJ, Kotzerke J, Hoekstra OS, Pruim J, Marsden PK, Tatsch K, Hoekstra CJ, Visser EP, Arends B, Verzijlbergen FJ, Zijlstra JM, Comans EF, Lammertsma AA, Paans AM, Willemsen AT, Beyer T, Bockisch A, Schaefer-Prokop C, Delbeke D, Baum RP, Chiti A, Krause BJ. FDG PET and PET/CT: EANM procedure guidelines for tumour PET imaging: version 1.0. *Eur J Nucl Med Mol Imaging* 2010;37:181-200.
12. Yokoyama I, Ohtake T, Momomura S, Yonekura K, Kobayakawa N, Aoyagi T, Sugiura S, Yamada N, Ohtomo K, Sasaki Y, Omata M, Yazaki Y. Insulin action on heart and skeletal muscle FDG uptake in patients with hypertriglyceridemia. *J Nucl Med* 1999;40:1116-1121.
13. Ozguven MA, Karacalioglu AO, Ince S, Emer MO. Altered biodistribution of FDG in patients with type-2 diabetes mellitus. *Ann Nucl Med* 2014;28:505-511.
14. Standards of medical care in diabetes--2011. *Diabetes Care* 2011;34 Suppl 1:S11-61.
15. Lee YH, Kwon MH, Kim KJ, Lee EY, Kim D, Lee BW, Kang ES, Cha BS, Lee HC. Inverse Association between Glycated Albumin and Insulin Secretory Function May Explain Higher Levels of Glycated Albumin in Subjects with Longer Duration of Diabetes. *PLoS One* 2014;9:e108772.

16. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985;28:412-419.
17. Groop LC, Bonadonna RC, DelPrato S, Ratheiser K, Zyck K, Ferrannini E, DeFronzo RA. Glucose and free fatty acid metabolism in non-insulin-dependent diabetes mellitus. Evidence for multiple sites of insulin resistance. *J Clin Invest* 1989;84:205-213.
18. Gastaldelli A, Cusi K, Pettiti M, Hardies J, Miyazaki Y, Berria R, Buzzigoli E, Sironi AM, Cersosimo E, Ferrannini E, DeFronzo RA. Relationship between hepatic/visceral fat and hepatic insulin resistance in nondiabetic and type 2 diabetic subjects. *Gastroenterology* 2007;133:496-506.
19. DeFronzo RA, Gunnarsson R, Bjorkman O, Olsson M, Wahren J. Effects of insulin on peripheral and splanchnic glucose metabolism in noninsulin-dependent (type II) diabetes mellitus. *J Clin Invest* 1985;76:149-155.
20. Cnop M, Landchild MJ, Vidal J, Havel PJ, Knowles NG, Carr DR, Wang F, Hull RL, Boyko EJ, Retzlaff BM, Walden CE, Knopp RH, Kahn SE. The concurrent accumulation of intra-abdominal and subcutaneous fat explains the association between insulin resistance and plasma leptin concentrations : distinct metabolic effects of two fat compartments. *Diabetes* 2002;51:1005-1015.
21. Despres JP, Moorjani S, Lupien PJ, Tremblay A, Nadeau A, Bouchard C. Regional distribution of body fat, plasma lipoproteins, and cardiovascular disease. *Arteriosclerosis* 1990;10:497-511.
22. Boden G, Chen X, Ruiz J, White JV, Rossetti L. Mechanisms of fatty acid-induced inhibition of glucose uptake. *J Clin Invest* 1994;93:2438-2446.
23. Knuuti MJ, Maki M, Yki-Jarvinen H, Voipio-Pulkki LM, Harkonen R, Haaparanta M, Nuutila P. The effect of insulin and FFA on myocardial glucose uptake. *J Mol Cell Cardiol* 1995;27:1359-1367.
24. Ng CK, Soufer R, McNulty PH. Effect of hyperinsulinemia on myocardial fluorine-18-FDG uptake. *J Nucl Med* 1998;39:379-383.
25. Yokoyama I, Yonekura K, Ohtake T, Kawamura H, Matsumoto A, Inoue Y, Aoyagi T, Sugiura S, Omata M, Ohtomo K, Nagai R. Role of insulin resistance in heart and skeletal muscle F-18 fluorodeoxyglucose uptake in patients with non-insulin-dependent diabetes mellitus. *J Nucl Cardiol* 2000;7:242-248.
26. Kobayashi Y, Kumita S, Fukushima Y, Ishihara K, Suda M, Sakurai M. Significant suppression of myocardial (18)F-fluorodeoxyglucose uptake using 24-h carbohydrate restriction and a low-carbohydrate, high-fat diet. *J Cardiol* 2013;62:314-319.
27. Lautamaki R, Airaksinen KE, Seppanen M, Toikka J, Luotolahti M, Ball E, Borra R, Harkonen R, Iozzo P, Stewart M, Knuuti J, Nuutila P. Rosiglitazone improves myocardial glucose uptake in patients with type 2 diabetes and coronary artery disease: a 16-week randomized, double-blind, placebo-controlled study. *Diabetes* 2005;54:2787-2794.
28. Hicks RJ, Herman WH, Kalff V, Molina E, Wolfe ER, Hutchins G, Schwaiger M. Quantitative evaluation of regional substrate metabolism in the human heart by positron emission tomography. *J Am Coll Cardiol* 1991;18:101-111.
29. Banerji MA, Faridi N, Atluri R, Chaiken RL, Lebovitz HE. Body composition, visceral fat, leptin, and insulin resistance in Asian Indian men. *J Clin Endocrinol*

- Metab* 1999;84:137-144.
30. DeNino WF, Tchernof A, Dionne IJ, Toth MJ, Ades PA, Sites CK, Poehlman ET. Contribution of abdominal adiposity to age-related differences in insulin sensitivity and plasma lipids in healthy nonobese women. *Diabetes Care* 2001;24:925-932.
 31. Raji A, Seely EW, Arky RA, Simonson DC. Body fat distribution and insulin resistance in healthy Asian Indians and Caucasians. *J Clin Endocrinol Metab* 2001;86:5366-5371.
 32. Matsuura F, Yamashita S, Nakamura T, Nishida M, Nozaki S, Funahashi T, Matsuzawa Y. Effect of visceral fat accumulation on uric acid metabolism in male obese subjects: visceral fat obesity is linked more closely to overproduction of uric acid than subcutaneous fat obesity. *Metabolism* 1998;47:929-933.
 33. Yoo TW, Sung KC, Shin HS, Kim BJ, Kim BS, Kang JH, Lee MH, Park JR, Kim H, Rhee EJ, Lee WY, Kim SW, Ryu SH, Keum DG. Relationship between serum uric acid concentration and insulin resistance and metabolic syndrome. *Circ J* 2005;69:928-933.
 34. Facchini F, Chen YD, Hollenbeck CB, Reaven GM. Relationship between resistance to insulin-mediated glucose uptake, urinary uric acid clearance, and plasma uric acid concentration. *Jama* 1991;266:3008-3011.
 35. Hasegawa S, Kusuoka H, Uehara T, Yamaguchi H, Hori M, Nishimura T. Glucose tolerance and myocardial F-18 fluorodeoxyglucose uptake in normal regions in coronary heart disease patients. *Ann Nucl Med* 1998;12:363-368.
 36. vom Dahl J, Herman WH, Hicks RJ, Ortiz-Alonso FJ, Lee KS, Allman KC, Wolfe ER, Jr., Kalff V, Schwaiger M. Myocardial glucose uptake in patients with insulin-dependent diabetes mellitus assessed quantitatively by dynamic positron emission tomography. *Circulation* 1993;88:395-404.
 37. Gropler RJ. Methodology governing the assessment of myocardial glucose metabolism by positron emission tomography and fluorine 18-labeled fluorodeoxyglucose. *J Nucl Cardiol* 1994;1:S4-14.
 38. Aerni-Flessner L, Abi-Jaoude M, Koenig A, Payne M, Hruz PW. GLUT4, GLUT1, and GLUT8 are the dominant GLUT transcripts expressed in the murine left ventricle. *Cardiovasc Diabetol* 2012;11:63.
 39. Garvey WT, Hardin D, Juhaszova M, Dominguez JH. Effects of diabetes on myocardial glucose transport system in rats: implications for diabetic cardiomyopathy. *Am J Physiol* 1993;264:H837-844.
 40. Karim S, Adams DH, Lalor PF. Hepatic expression and cellular distribution of the glucose transporter family. *World J Gastroenterol* 2012;18:6771-6781.
 41. Nikolaidis LA, Elahi D, Hentosz T, Doverspike A, Huerbin R, Zourelis L, Stolarski C, Shen YT, Shannon RP. Recombinant glucagon-like peptide-1 increases myocardial glucose uptake and improves left ventricular performance in conscious dogs with pacing-induced dilated cardiomyopathy. *Circulation* 2004;110:955-961.
 42. Gejl M, Lerche S, Mengel A, Moller N, Bibby BM, Smidt K, Brock B, Sondergaard H, Botker HE, Gjedde A, Holst JJ, Hansen SB, Rungby J. Influence of GLP-1 on myocardial glucose metabolism in healthy men during normo- or hypoglycemia. *PLoS One* 2014;9:e83758.
 43. van den Brom CE, Bosmans JW, Vlasblom R, Handoko LM, Huisman MC, Lubberink M, Molthoff CF, Lammertsma AA, Ouwens MD, Diamant M, Boer C. Diabetic cardiomyopathy in Zucker diabetic fatty rats: the forgotten right ventricle. *Cardiovasc Diabetol* 2010;9:25.

44. Rodrigues B, Cam MC, McNeill JH. Metabolic disturbances in diabetic cardiomyopathy. *Mol Cell Biochem* 1998;180:53-57.
45. van den Brom CE, Huisman MC, Vlasblom R, Boontje NM, Duijst S, Lubberink M, Molthoff CF, Lammertsma AA, van der Velden J, Boer C, Ouwens DM, Diamant M. Altered myocardial substrate metabolism is associated with myocardial dysfunction in early diabetic cardiomyopathy in rats: studies using positron emission tomography. *Cardiovasc Diabetol* 2009;8:39.

ABSTRACT (IN KOREAN)

양전자방출단층촬영술 시행 시 보이는 공복 상태 심근의 FDG
섭취와 정상, 당뇨 전 단계, 제 2형 당뇨병 환자의 내장 지방과의
연관성에 대한 연구

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조관형

연구목적: 정상환자와 당뇨 전 단계, 그리고 제 2형 당뇨병환자에서 양전자방출단층촬영 시행 시에 관찰되는 심근의 FDG 섭취 정도를 다양한 임상적 특징과 혈액학적 검사와 비교하여 그 임상적 의미를 알아보고자 하였다.

대상 및 방법: 건강검진을 목적으로 양전자방출단층촬영술을 시행한 총 387명을 정상군, 당뇨 전 단계, 제 2형 당뇨병의 세 군으로 구분하여 공복시의 심근 포도당 섭취, 내장 및 피하 지방 영역 분석, 혈액 검사를 시행하였으며 이에 대한 유의미한 차이와 관련성을 조사하였다.

결과: 심근의 FDG 섭취는 제 2형 당뇨병 환자 그룹에서 유의하게 감소하였고(P for trend=0.001), 다변량 선형 회귀 분석에서는 내장 지방 면적($\beta = -0.31$, $P < 0.001$), 공복 FFA($\beta = -0.34$, $P < 0.001$) 및 요산 수치($\beta = -0.21$, $P = 0.007$)와 심근의 FDG 섭취와의 관련성이 보고되었다.

결론: 심근의 FDG 섭취는 제 2형 당뇨병을 예측할 수 있고, 내장 지방 및 지방산 그리고 요산의 대사와 유의미한 관련이 있음이 보였다.

핵심되는 말 : 심근 대사, 양전자방출단층촬영술, 제 2형 당뇨병