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**Toll-like receptor 4 signaling-mediated
responses are critically engaged in
optimal host protection against highly
virulent *Mycobacterium tuberculosis*
K infection**

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Department of Medical Science

The Graduate School, Yonsei University

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Directed by Professor Sung Jae Shin

The Master's Thesis
submitted to the Department of Medical Science,
the Graduate School of Yonsei University
in partial fulfillment of the requirements for the degree of
Master of Medical Science

Jaehun Park

February 2020

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박재훈 올림

ABSTRACT

**Toll-like receptor 4 signaling-mediated responses are critically engaged
in optimal host protection against highly virulent
Mycobacterium tuberculosis K infection**

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(Directed by Professor Sung Jae Shin)

Toll-like receptors (TLRs) play critical roles in the innate recognition of *Mycobacterium* spp. by host immune cells. However, controversy has arisen regarding the role TLR4 and its related signaling pathways play in determining the outcomes of *Mycobacterium tuberculosis* (Mtb) infection. To address this controversy, the function of TLR4 in the induction of an optimal protective immune response against Mtb infection was comparatively investigated in C3H/HeJ (TLR4-deficient mutant) and C3H/HeN (TLR4-competent wild-type) mice by aerosol-mediated infection with the highly virulent K strain of Mtb. Interestingly, following Mtb

infection, C3H/HeJ mice showed a more severe disease phenotype than C3H/HeN mice, exhibiting reduced survival rates and a marked increase in bacterial burden along with necrotic lung inflammation. Further analysis of the immune cell composition revealed a significant increase in the number of neutrophils in the infected lung tissue and a high level of IL-10 production accompanied by the impairment of the protective Th1-type T cell response in C3H/HeJ mice. Reducing the neutrophil numbers by treating C3H/HeJ mice with an anti-Ly6G monoclonal antibody (mAb) and blocking IL-10 signaling with an anti-IL-10 receptor mAb reduced the excessive lung inflammation and bacterial burden in C3H/HeJ mice. Therefore, abundant IL-10 signaling and neutrophils have detrimental effects in TLR4-deficient mice during Mtb infection. However, the blockade of IL-10 signaling produced an increase in the CD11b^{hi}Ly6G^{hi} neutrophil population, but the phenotypes of these neutrophils were different from those of the CD11b^{int}Ly6G^{int} neutrophils from mice with controlled infections. These results indicate that TLR4 and its relevant downstream signals positively contribute to the generation of an optimal protective immune response against Mtb infection. Collectively, these results show that TLR4 plays an essential role in the optimal induction of a protective response against Mtb infection. Furthermore, investigating the TLR4-mediated response will provide insight for the development of effective control measures, such as host-directed therapy and vaccines against TB.

Keywords: *Mycobacterium tuberculosis*; TLR4, Neutrophil, Th1-type response, IL-10

I. INTRODUCTION

Tuberculosis (TB), caused by *Mycobacterium tuberculosis* (Mtb), is highly contagious and the leading infectious disease, causing 1.6 million human deaths worldwide in 2017. TB is a life-threatening disease that is newly diagnosed more than 10 million times every year; Mtb has more than 90 different virulence factors that induce host immune cell interactions with the pathogen. The design of an effective treatment and vaccine for TB control by dissecting various virulence factors requires investigations of the primary immune responses that protect the host and the immunological understanding of the susceptibility to TB. The incomplete understanding of the pathogenesis of Mtb infection makes it difficult to control this detrimental pathogen.

The innate recognition of mycobacterial products is the first step in a series of events leading to effective host defense against Mtb infection. Antigen-presenting cells, such as macrophages and dendritic cells, express pattern recognition receptors (PRRs) that recognize conserved molecular patterns, the so-called pathogen-associated molecular patterns (PAMPs)¹. Toll-like receptors (TLRs) are one of the well-characterized PRR families. Functionally, TLRs 1-10 in humans and TLRs 1-9 and 11-13 in mice have been discovered; consequently, studies of the immune response related to various bacteria, viruses and fungi are underway. Among the TLRs, TLR2, TLR4, and TLR9 are well known to be involved in the recognition of Mtb²⁻⁴. In addition, genetic risk for Mtb infection may be increased by defects or polymorphisms in the TLR family⁵.

Polymorphisms in TLR2 and TLR4 genes might cause a reduced macrophage response to bacterial components, resulting in increased susceptibility to TB⁶. In particular, the synergistic effect of the TLR4 Thr399Ile and TLR9-1486T/C polymorphism may increase susceptibility to TB⁷.

Appropriate immune responses mediated by host cells present resistance to TB⁸. In particular, interferon-gamma (IFN)- γ -secreting CD4⁺ T cells are essential for the protective immune response to mycobacterial infection^{9,10}. Various animal models with a disrupted IFN- γ response are unable to restrain the growth of Mtb and capitulate to the infection¹¹⁻¹³. Other types of inflammatory cytokines, such as TNF- α and interleukin (IL)-12, are important for limiting Mtb infection through granuloma formation and activating T cell responses¹⁴⁻¹⁶. In contrast to pro-inflammatory cytokines such as TNF- α and IL-12, anti-inflammatory cytokines such as IL-10 inhibit the immune response of the host, creating a favorable environment for the growth of Mtb¹⁷. Mtb induces IL-10 secretion via the TLR2-ERK pathway¹⁸. A number of studies have shown that IL-10 secreted in response to Mtb is associated with susceptibility to TB in human and mouse models¹⁹⁻²⁴. For example, excessive IL-10 secretion is known to induce a decreased Th1 response and macrophage function^{17,25-29}. The ablation of IL-10 signaling in mice infected with Mtb is beneficial for the control of bacterial growth and improves mouse survival due to the restoration of CD4⁺ T cells and the Th1 responses³⁰.

Although several antigens derived from Mtb or Mtb itself bind to TLR4³¹⁻³³, resulting in a variety of alterations in immunobiological responses such as immune activation or cell death promotion, there remains controversy around the functional role TLR4 plays in the pathogenesis of TB. C3H/HeJ and C3H/HeN mouse substrains were derived from the same parental strain C3H/He in 1947³⁴. C3H/HeJ mice carry a missense mutation in the TLR4 gene, which induces a single amino acid change in the cytoplasmic portion of TLR4, disrupts signal transduction and induces a phenotype similar to that of TLR4-knockout mice³⁵⁻³⁷. C3H/HeJ mice have a specific tolerance to lipopolysaccharide (LPS), unlike C3H/HeN, due to the TLR4 mutation^{35,38}. However, C3H/HeJ mice are highly susceptible to infection by bacteria, such as *E. coli*, *Salmonella typhimurium* and *Neisseria meningitides*^{39,40}.

Components of Mtb, such as 3- and 4-acylated lipomannan and 60- and 65-heat shock proteins, activate various immune cell pathways when they bind to TLR4⁴¹. In particular, TLR4 ligation plays a major role in inducing T helper (Th) 1 responses by activating dendritic cells, which interact with T cells⁴². Because of this characteristic, TLR4 agonists have been developed as vaccine adjuvant candidates for hepatitis B and papilloma^{43,44}. Adjuvants from TB are being studied using glucopyranosyl lipid adjuvant (GLA), which is a TLR4 agonist combined with a stable emulsion formulation, to increase the efficacy of the existing vaccine⁴⁵⁻⁴⁷. However, the role of TLR4 during Mtb infection is still under discussion. Previous studies have shown inconsistent results regarding the protective role TLR4 plays against Mtb infection.

For example, Abel *et al.* reported that TLR4 is necessary to control chronic Mtb infection in mice, indicating that TLR4 mediates the protective immune response during chronic TB⁴⁸. In addition, Branger *et al.* showed that TLR4 is required to produce IFN- γ upon antigen-specific stimulation, suggesting that TLR4 plays a protective role in host defense against lung infection by Mtb⁴⁹. However, other studies have demonstrated that TLR4 does not protect against Mtb infection. For example, Shim *et al.* showed that TLR4-competent and TLR4-deficient mice showed similar types of cell accumulation and lung histopathology⁵⁰. Furthermore, Reiling *et al.* showed that TLR4 signaling is not required to control Mtb growth⁵¹.

In this study, we challenged C57BL/6J, C3H/HeN, and C3H/HeJ mice to clarify the functional role of TLR4 and define the immunological alterations that are involved in host protection during clinical Mtb infection. High levels of IL-10 secretion and neutrophil recruitment were found in the absence of TLR4 in mice. These results indicate that TLR4 might be involved in susceptibility to TB. Thus, our research reveals that TLR4 could be necessary for host protection against pulmonary TB.

II. MATERIALS AND METHODS

1. Experimental animals and ethics statement

Specific pathogen-free 6- or 7-week-old C57BL/6J, C3H/HeN, and C3H/HeJ mice were purchased from Japan SLC, Inc. (Shizuoka, Japan) or the Jackson Laboratory (Bar Harbor, ME, USA) and strictly maintained under barrier conditions in a BSL-3 facility at Avison Biomedical Research Center at Yonsei University College of Medicine (Seoul, Korea). All animal studies were performed in accordance with the Korean Food and Drug Administration (KFDA) guidelines. The experimental protocols used in this study were approved by the Ethics Committee and Institutional Animal Care and Use Committee of the Laboratory Animal Research Center at Yonsei University College of Medicine (permit number: 2017-0342).

2. Mtb K strain and culture conditions

Mtb K strain was collected from the Korean Tuberculosis Research Institute (KIT, Osong, Chungcheongbuk-do) and was cultured for infection experiments as described previously.

3. Mtb K strain infections and antibody treatments

Mice were infected with the Mtb K strain via aerosolization as previously described. Briefly, mice were exposed to the Mtb K strain in the calibrated inhalation chamber of an airborne infection apparatus for 60 min delivering a predetermined dose (Glas-Col, Terre Haute, IN, USA); approximately 150 viable bacteria were delivered to the

lungs. After 2 weeks, 250 $\mu\text{g}/\text{mouse}$ anti-IL-10 receptor mAb (1B1.3A; Bio X Cell, West Lebanon, NH, USA) in PBS or 200 $\mu\text{g}/\text{mouse}$ for the blockade of IL-10 signaling, and 250 $\mu\text{g}/\text{mouse}$ anti-Ly6G mAb (1A8; Bio X Cell, West Lebanon, NH, USA) in PBS or 200 $\mu\text{g}/\text{mouse}$ to reduce the neutrophil number, were injected intraperitoneally 3 times per week. All mice were sacrificed 3 or 4 weeks after Mtb K strain infection.

4. Antibodies and reagents

A LIVE/DEAD[®] Fixable Near-IR Dead Cell Stain Kit; Green Dead Cell Stain Kit; and Aqua Dead Cell Stain Kit were purchased from Molecular Probes (Carlsbad, CA, USA). A phycoerythrin (PE)-conjugated mAb against IFN- γ ; violet 450-conjugated mAb against CD44; brilliant violet (BV) 605-conjugated mAb against Thy1.2; PerCP-Cy5.5-conjugated mAb against CD4; BV 421-conjugated mAb against CD45; and a V 450-conjugated mAb against Ly6G were purchased from BD Bioscience (San Jose, CA, USA) and a PerCP-Cy5.5-conjugated mAb against CD11b; APC-Cy7-conjugated mAb against I-Ab; PE-conjugated mAb against CD64; and a PE-Dazzle-conjugated mAb against CD11c were purchased from Biolegend (San Diego, CA, USA) for flow cytometry analyses. Recombinant mouse granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-4 were purchased from JW CreaGene (Daegu, Korea).

5. Generation of bone marrow-derived dendritic cells

Red blood cells (RBCs) among whole bone marrow cells from C3H/HeJ and C3H/HeN mice were lysed using RBC Lysis Buffer (Sigma-Aldrich, St. Louis, MO, USA) and then the cells were maintained in Petri dishes (1×10^6 cells/mL; 10 mL/Petri dish) and cultured at 37 °C in the presence of 5% CO₂ using complete-RPMI 1640 (c-RPMI 1640) supplemented with 100 U/mL penicillin/streptomycin (Lonza, Basel, Switzerland), 10% fetal bovine serum (Lonza, Basel, Switzerland), GM-CSF (20 ng/mL) and IL-4 (0.5 ng/mL). To culture plates was added 10 mL of c-RPMI 1640 medium per well on day 3, which was replaced with 10 mL c-RPMI 1640 medium on day 6 of culture. After 9 days of culture, the cells were harvested and used for experiments.

6. Preparation of single cells and immune cell analysis

Lung single-cell suspensions were prepared from mice 4 weeks after Mtb infection. To obtain a single-cell suspension, the lung tissue was minced into 2-4 mm pieces with scissors. The lung tissue was incubated in 3 mL of cellular dissociation buffer RPMI medium (Biowest, Nuaille, France) containing 0.1% collagenase type IV (Worthington Biochemical Corporation, Lakewood, NJ, USA), 1 mM CaCl₂ and 1 mM MgCl₂ for 30 min at 37 °C. Lung and spleen cells were separated through 40- μ m cell strainers (Corning, NY, USA), and RBCs were lysed with ACK lysis buffer (Lonza, Basel, Switzerland) for 3 min at room temperature. After the cells were with c-RPMI 1640, a single-cell suspension was prepared.

7. Cytokine measurement by ELISA

Single cells from the lungs and spleens of Mtb-infected or immunized mice were stimulated with early secreted antigenic targets of 6 kDa (ESAT-6) for 12 hours at 37 °C. A sandwich enzyme-linked immunosorbent assay (ELISA) was used to detect TNF- α , IFN- γ , IL-6, IL-1 β , IL-12p70, IL-10, and IL-17AF (Invitrogen, San Diego, CA, USA).

8. Analysis of IFN- γ secreting CD4⁺ T cells by intracellular staining

Lung single-cell suspensions were prepared from immunized and Mtb-infected mice and stimulated with 1 μ g/mL ESAT-6 at 37 °C for 12 hours in the presence of GolgiPlug and GolgiStop (BD Bioscience, San Jose, CA, USA). First, the cells were washed with PBS, and the Fc receptor was blocked with an anti-CD16/32 blocking antibody at 4 °C for 15 min. Surface molecules were stained with fluorochrome-conjugated antibodies against Thy1.2, CD4, CD44, and a Live and DEADTM Fixable Dead Cell kit for 30 min at 4 °C. After being washed with PBS, the cells were fixed and permeabilized with Cytotfix/Cytoperm (BD Biosciences, San Jose, CA, USA) for 30 min at 4 °C. The permeabilized cells were washed twice with Perm/Wash (BD Biosciences, San Jose, CA, USA) and stained with PE-conjugated anti-IFN- γ , for 30 min at 4 °C. The cells were washed twice with Perm/Wash and fixed with intracellular (IC) fixation buffer (eBioscience, San Diego, CA, USA) until analysis by flow cytometry.

9. Histopathological analysis and enumeration of Mtb

The disease severity and phenotype were evaluated through histopathology and bacterial growth in the lungs and spleen. The organs were removed to determine protection at 4 weeks after infection. For the lung histopathology, the right-superior lobes were preserved overnight in 10% formalin and embedded in paraffin. The lungs were sectioned at 4-5 μm and H&E stained. The severity of lung inflammation was examined using the ImageJ program (National Institutes of Health, USA) as previously described⁵². For bacterial growth analysis, the lungs and spleens were homogenized, and serially diluted samples were plated onto Middlebrook 7H10 agar (Becton Dickinson, Franklin Lakes, NJ, USA) supplemented with 10% OADC (Difco Laboratories, Detroit, MI, USA), 2 $\mu\text{g}/\text{mL}$ 2-thiophenecarboxylic acid hydrazide (Sigma-Aldrich, St. Louis, MO, USA) and amphotericin B (Sigma-Aldrich, St. Louis, MO, USA). After 4 weeks of incubation at 37 °C, bacterial colonies were counted.

10. Statistical analyses

Statistical analyses were performed using GraphPad Prism V7.0 (GraphPad Software, La Jolla, CA, USA). The results are expressed as the mean \pm standard deviation (SD). Comparisons between two groups were conducted by an unpaired *t*-test. Statistical significance was determined as **p* < 0.05, ***p* < 0.01 or ****p* < 0.001.

III. RESULTS

1. TLR4-deficient mutant C3H/HeJ mice are more susceptible than TLR4-competent C57BL/6J and C3H/HeN mice to Mtb infection

To confirm the deficiency in TLR4 signaling, bone marrow-derived dendritic cells (BMDCs) differentiated from C3H/HeN and C3H/HeJ mice were treated with TLR2 and TLR4 agonists, respectively. BMDCs differentiation from C3H/HeN and C3H/HeJ mice was confirmed by flow cytometry (Figure 1A), and more than 90% of the cells were normally differentiated into BMDCs (Figure 1B). The LPS stimulation-induced pro-inflammatory cytokine secretion was lower in C3H/HeJ mice than C3H/HeN mice (Figure 1C). No LPS-induced cytokine secretion was observed in BMDCs differentiated from C3H/HeJ mice, which are TLR4 deficient. However, the secretion of TNF- α , IL-12, IL-6, IL-10 and IL-1 β was observed in BMDCs differentiated from C3H/HeJ mice, and significantly increased IL-10 was detected in BMDCs differentiated from C3H/HeJ mice compared to BMDCs differentiated from C3H/HeN mice (Figure 2).

To study the role of TLR4 during Mtb infection, we observed the pathology through H&E staining, and the bacterial burden in the lungs and spleen was analyzed at 3 and 4 weeks postinfection. The formation of granuloma structures in the lungs is a symbol of the confinement of the Mtb bacterial burden in Mtb-resistant strains of mice. C57BL/6J and C3H/HeN mice display the typical advancement of the disease, but C3H/HeJ mice deviated from the normal progression at 4 weeks postinfection. C3H/HeJ mice showed a greater inflamed lung lesion width than C57BL/6J and

C3H/HeN mice during Mtb K strain infection, which was not the case at 3 weeks postinfection (Figures 3A, B). The inflamed lesions in C3H/HeJ mice were similar to the necrotic lesions in C3HeB/FeJ mice, which are susceptible to Mtb. Differences in pathology between TLR4-competent and TLR4-deficient mutant mice were evident; thus, we investigated whether dissimilarities in the bacterial burden occur during the infection phase. The bacterial burden of the Mtb K strain in the lungs of three strains was analyzed. The bacterial growth in the lungs and spleen was significantly increased in C3H/HeJ mice compared to C57BL/6J and C3H/HeN mice at 4 weeks after Mtb infection (Figure 3C). Interestingly, the infection progression in the lungs of C3H/HeJ was greatly accelerated at 4 weeks postinfection. Mtb growth was controlled in the lungs of C57BL/6J and C3H/HeN mice to a similar extent. The loss of weight during Mtb infection was more pronounced in C3H/HeJ mice than C57BL/6J and C3H/HeN mice from 3 to 4 weeks (Figure 3D). These results demonstrate that TLR4 provides stronger protection against Mtb K strain infection in a mouse model.

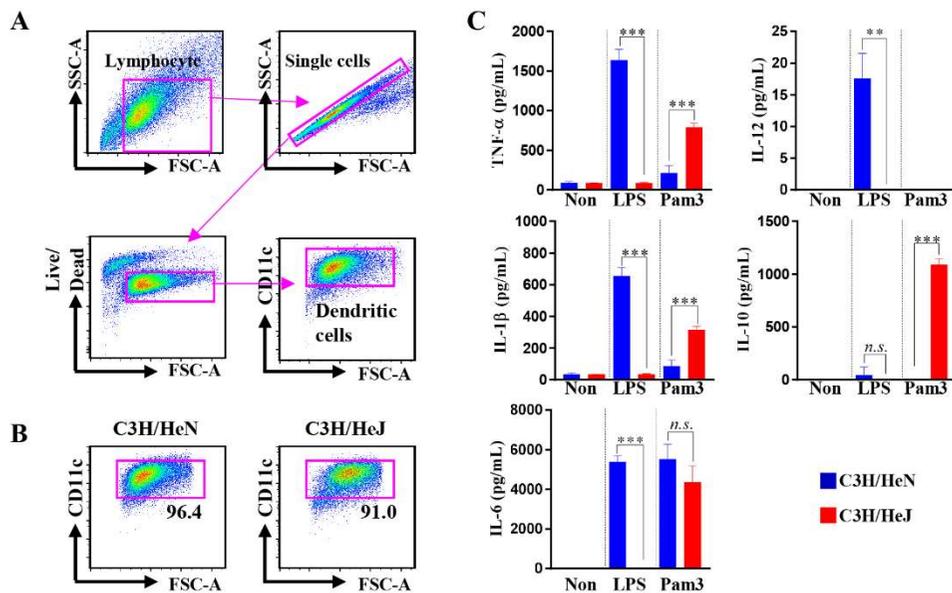


Figure 1. Confirmation of the TLR4 response in bone marrow-derived dendritic cells differentiated from C3H/HeN and C3H/HeJ mice after agonist stimulation. BMDCs from C3H/HeN and C3H/HeJ mice were treated with LPS (100 ng/mL) or Pam3CSK4 (300 ng/mL) for 24 hours. (A) A gating strategy was established to show BMDC differentiation. (B) The purity of BMDCs differentiated from C3H/HeN and C3H/HeJ mice confirmed through flow cytometry analysis. (C) TNF- α , IL-6, IL-12p70, IL-1 β , and IL-10 production was measured by ELISA. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value <0.05 was considered statistically significant; *n.s.*: not significant, ***p* < 0.01 and ****p* < 0.001 .

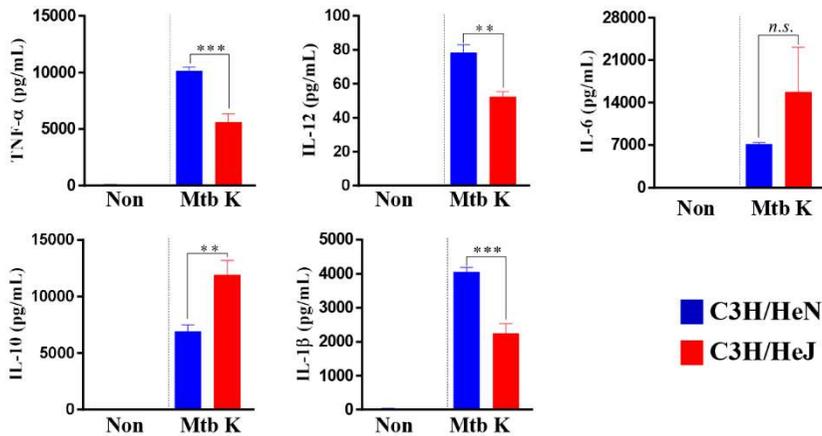


Figure 2. Differential cytokine profiles in bone marrow-derived dendritic cells differentiated from C3H/HeN and C3H/HeJ mice following Mtb K infection.

BMDCs differentiated from C3H/HeN and C3H/HeJ were infected with the Mtb K strain (MOI: 0.5) for 24 hours. TNF- α , IL-6, IL-12p70, IL-1 β , and IL-10 production was observed by ELISA. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value $<$ 0.05 was considered statistically significant; *n.s.*: not significant, ***p* $<$ 0.01 and ****p* $<$ 0.001.

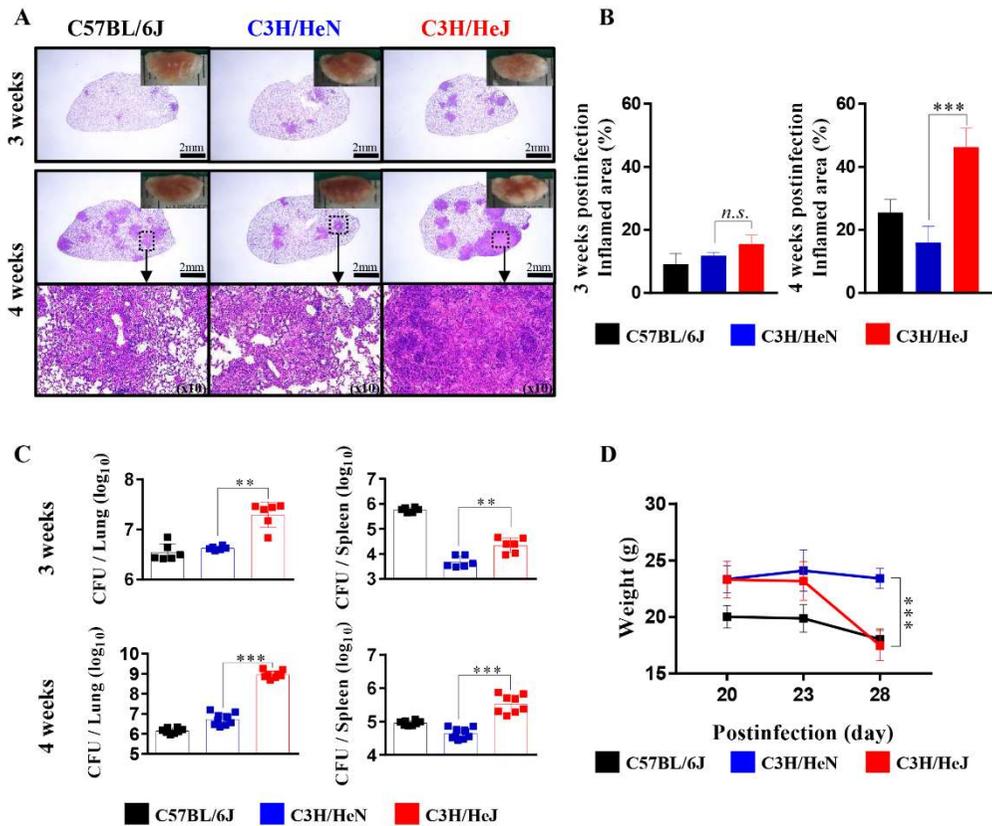


Figure 3. Comparative analysis of the pathogenesis in C3H/HeN and C3H/HeJ mice in terms of bacterial burden and inflammation in the lungs following infection with the K strain of Mtb. Mice were infected with 150 CFU of the Mtb K strain via aerosolization, and the lungs were removed at 3 and 4 weeks postinfection for analysis. (A) The superior lobe of the right lung stained with H&E at 3 and 4 weeks after Mtb K strain challenge. Gross lung pathology is shown with H&E staining. (B) The inflammation area of the H&E stained samples was quantified by percentage and presented as bar graphs. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with

an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant. *n.s.*: not significant and ****p* < 0.001. (C) The CFUs in the lungs and spleens of each group at 3 and 4 weeks postinfection were analyzed by counting the bacteria. The data are presented as the mean ± SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value <0.05 was considered statistically significant. ***p* < 0.01 and ****p* < 0.001. (D) Changes in body weight were measured from 3 weeks after infection for 1 week until autopsy. The data are presented as the mean ± SD from four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value <0.05 was considered statistically significant. ****p* < 0.001 compared to C3H/HeN and C3H/HeJ.

2. Lack of TLR4 signaling reduces the recruitment of T cells and promotes the influx of neutrophils into the lungs of C3H/HeJ mice during Mtb infection

We determined that the bacterial burden, inflammation and weight loss was greater in C3H/HeJ mice than C57BL/6J and C3H/HeN mice due to the absence of TLR4 signaling. To identify the cell compartment that mediates the severe pathology and increased bacterial burden after infection, the influx of immune cells into the lung compartment was assessed through flow cytometry. The myeloid cells (dendritic cells, alveolar macrophages, macrophages, and neutrophils) and lymphoid cells (T cells) were examined by following a gating strategy (Figure 4). CD11c⁺Siglec-F⁺ alveolar macrophages, which contribute to the first line of defense against pathogens, were not changed during Mtb infection from weeks 3 to 4 in C57BL/6J, C3H/HeN and C3H/HeJ mice. The population of CD11c⁺MHCII⁺ dendritic cells, which directly interact with activated T cells, was decreased after Mtb K strain infection at 4 weeks in C3H/HeJ mice; however, C3H/HeJ mice contained comparable populations at 3 weeks postinfection. The populations of other first-line defense-related immune cells, neutrophils and macrophages, were significantly increased in C3H/HeJ mice from 3 to 4 weeks postinfection. Interestingly, the neutrophil population increased 10-fold over 3 to 4 weeks postinfection; however, the number of neutrophils in C57BL/6J and C3H/HeN mice tended to decrease or slightly increase (Figure 5A). The number of T cells, which regulate the adaptive immune response, was dramatically reduced in C3H/HeJ mice compared to C57BL/6J and C3H/HeN mice at 4 weeks postinfection (Figures 5B, C). There was no significant difference in T cell numbers between

C3H/HeJ and C57BL/6J, C3H/HeN mice at 3 weeks postinfection. These results suggest that the uncontrolled accumulation of neutrophils and insufficient number of T cells might correlate with TLR4 and Mtb susceptibility.

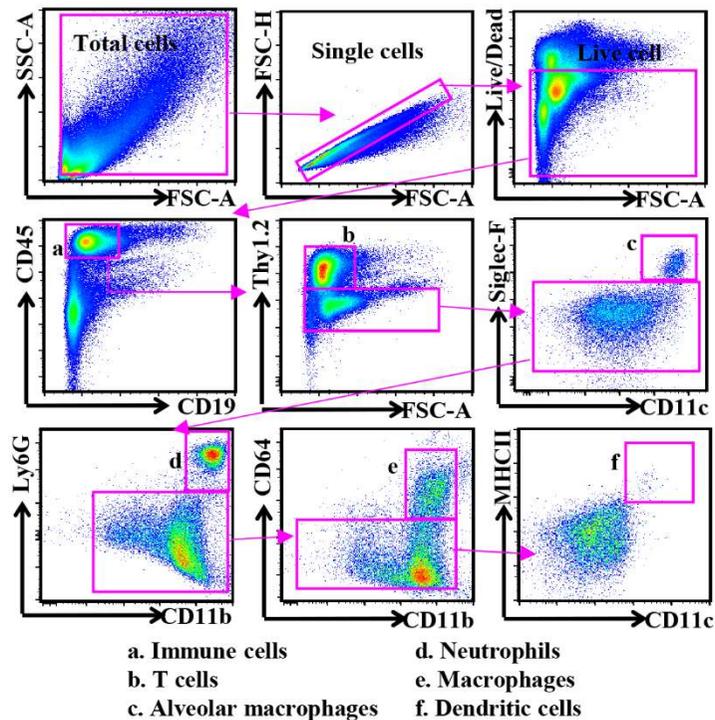


Figure 4. Flow cytometry gating strategies for the analysis of various types of immune cells. The immune cell population was first gated based on their characteristic forward scatter (FSC) and side scatter (SSC) patterns. Single cells were gated based on equivalent FSC height (FSC-H) and FSC area (FSC-A) values to exclude doublets and large cell aggregates. For the analysis of live immune cells, single-cell-gated leukocytes were stained with live and dead stains. (A) CD45⁺ immune cells were gated and then sub-gated into Thy1.2⁺ cells for the T cell population analysis. Thy1.2⁻ cells were gated into Siglec-F⁺ and CD11c⁺ cells to identify alveolar macrophages. Siglec-F⁻ cells were gated into Ly6G⁺ and CD11b⁺ cells for the neutrophil population analysis, and the remaining cells were further gated

into CD64⁺ and CD11b⁺ cells for the identification of macrophages. CD64-negative cells were gated into CD11c⁺MHCII⁺ cells for the analysis of the dendritic cell population.

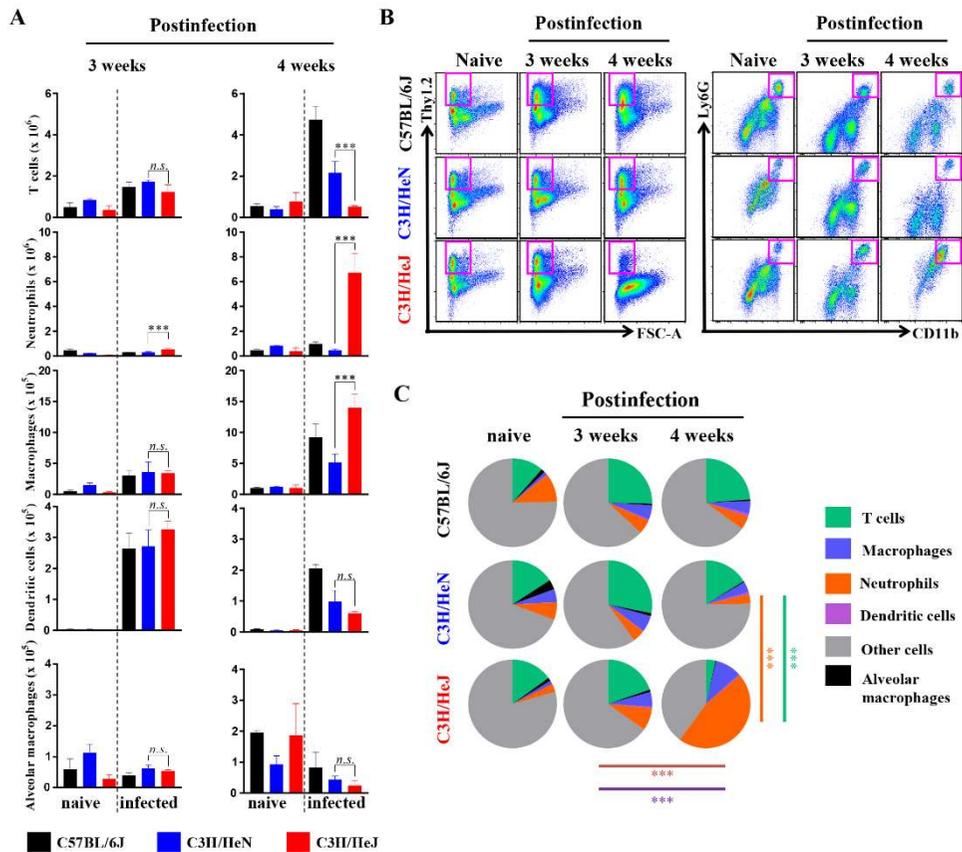


Figure 5. Analysis of cellular compartments in the lungs of C57BL/6, C3H/HeN and C3H/HeJ mice after challenge with Mtb K at 3 and 4 weeks post-infection. (A-B) The immune cell population from lung of B6, HeN and HeJ was analyzed with flow cytometry and shown as bar graphs. The data are presented as the mean \pm SD from four to five mice in each group. The significance of differences was determined with an unpaired *t*-test. *p* value <0.05 was considered statistically significant. *p* value <0.05 was considered statistically significant. *n.s.*: not significant and $***p<0.001$. (C) Pie charts indicated the mean frequencies of influx immune cells in the lungs at 3 and 4 weeks post-infection. The data are presented as the mean \pm SD from four to

five mice in each group. The significance of differences was determined with an unpaired *t*-test. *p* value <0.05 was considered statistically significant. The significance of differences was determined with an unpaired *t*-test. *p* value <0.05 was considered statistically significant. *n.s.*: not significant and **p* < 0.01. Orange line: significance of the neutrophil proportion between C3H/HeN and C3H/HeJ mice 4 weeks postinfection. Green-blue line: significance of the T cell proportion between C3H/HeN and C3H/HeJ mice 4 weeks postinfection. Violet line: significance of the neutrophil proportion between 3 weeks and 4 weeks postinfection in C3H/HeJ mice. Brown line: significance of the T cell proportion between 3 weeks and 4 weeks postinfection in C3H/HeJ mice.

3. Absence of TLR4 signaling leads to a decrease in the protective Th1 response and an increase in IL-10 secretion

The secretion of IFN- γ by CD4⁺ T cells is essential for reducing host damage by Mtb infection. Thus, we measured the cytokine production capacity of lung cells at 3 and 4 weeks postinfection to understand the process contributing to the Mtb susceptibility of C3H/HeJ mice. The early increase in CD4⁺IFN- γ ⁺ T cells in the lungs of C57BL/6J mice correlated with protection against Mtb infection. The number of CD4⁺IFN- γ ⁺ T cells was analyzed by flow cytometry by following a gating strategy (Figure 6B). C3H/HeJ mice produced less IFN- γ than C57BL/6J and C3H/HeN mice at 3 and 4 weeks postinfection (Figures 7A, B top panel). However, the population of CD4⁺CD44⁺ T cells, which are activated T cells, was similar in C57BL/6J, C3H/HeN and C3H/HeJ mice at 3 and 4 weeks postinfection (Figure 7B bottom panel). In contrast, IL-10, which is an immunosuppressive cytokine, was detected at higher concentrations in C3H/HeJ mice than C57BL/6J and C3H/HeN mice at 4 weeks postinfection, in contrast to 3 weeks postinfection (Figure 7C). These data suggest that the Th1 response was suppressed due to the high level of the immunosuppressive cytokine IL-10, which is one of the reasons that C3H/HeJ mice show increased susceptibility to Mtb infection.

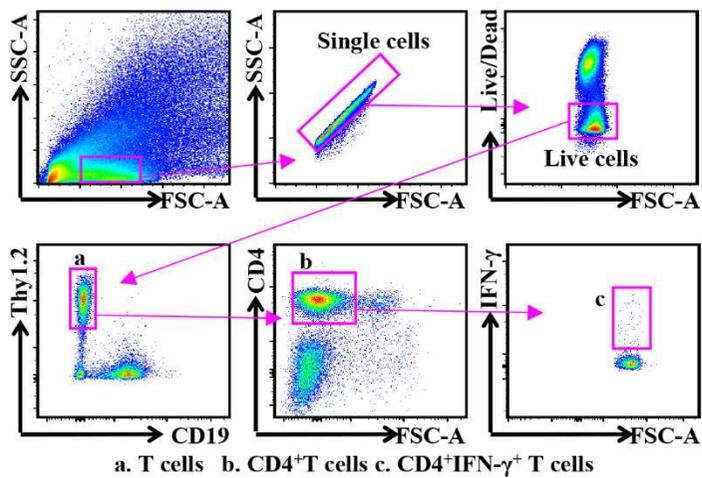


Figure 6. Flow cytometry gating strategies for the analysis of T cell response.

The immune cell population was first gated based on their characteristic forward scatter (FSC) and side scatter (SSC) patterns. Single cells were gated based on equivalent FSC height (FSC-H) and FSC area (FSC-A) values to exclude doublets and large cell aggregates. For the analysis of live immune cells, single-cell-gated leukocytes were stained with live and dead stains. Thy1.2⁺ cells were gated into CD4⁺ T cells. CD4⁺ cells were gated to profile the IFN- γ -producing T cells.

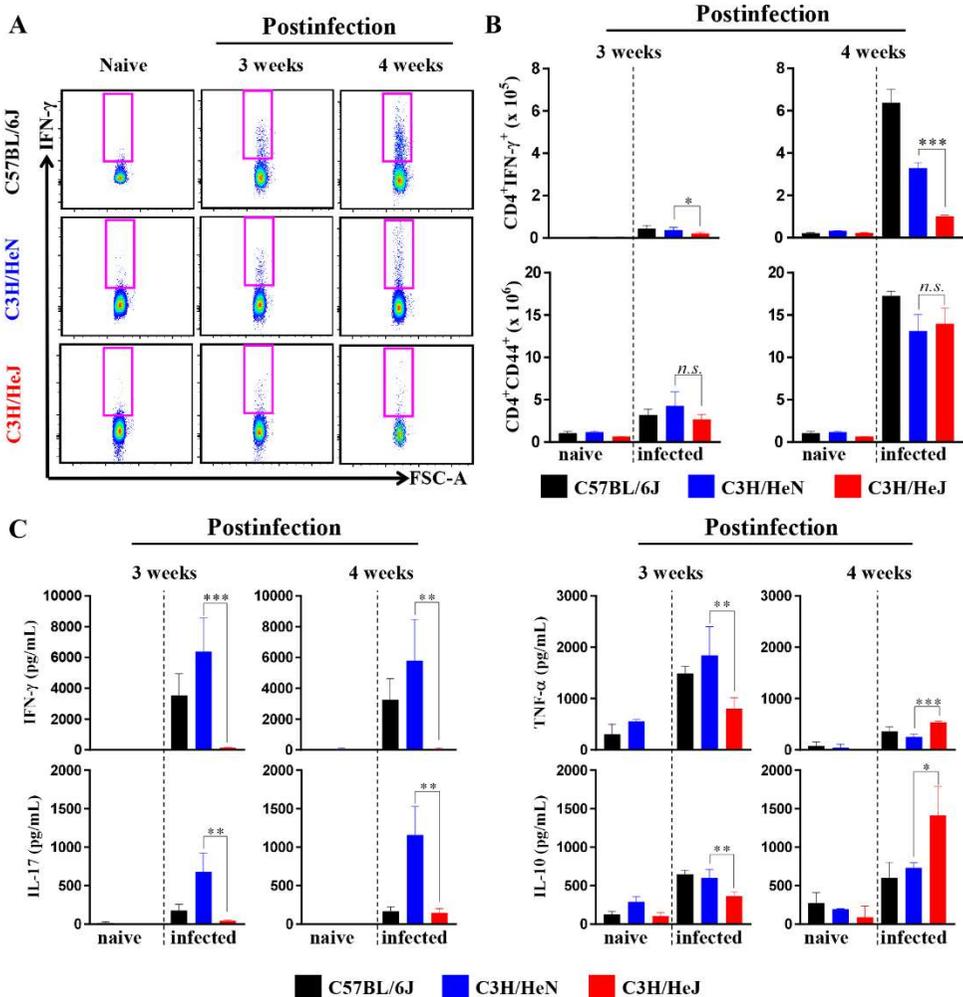


Figure 7. Comparative analysis of antigen-specific T cell responses and cytokine production in the lungs of C57BL/6J, C3H/HeN, and C3H/HeJ mice. At three and four weeks postinfection, mice from each group were euthanized, and their lung cells (1×10^6 cells) were stimulated with ESAT-6 ($1 \mu\text{g/mL}$) for 12 hours at 37°C in the presence of brefeldin A and monensin. (A-B) At 3 and 4 weeks postinfection, $\text{CD4}^+\text{CD44}^+$ T cells and $\text{IFN-}\gamma$ -secreting CD4^+ T cells in C57BL/6J, C3H/HeN and

C3H/HeJ mice were analyzed by flow cytometry, and the results are presented as bar graphs. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant. *n.s.*: not significant, $*p < 0.05$, and $***p < 0.001$. (C) The IFN- γ , IL-17AF, TNF- α , and IL-10 concentration in the supernatant were measured by ELISA. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant. $*p < 0.05$, $**p < 0.01$, and $***p < 0.001$.

4. Neutrophil removal contributes to the control of the excessive inflammation and bacterial burden in the lungs of C3H/HeJ mice

Compared to C3H/HeN mice, TLR4-deficient C3H/HeJ mice displayed a significantly increased neutrophil population. Thus, we designed an experiment to determine whether the markedly increased neutrophil population interferes with Mtb regulation in TLR4-deficient mice. The number of neutrophils in the lung was decreased after 2 weeks of Mtb infection with anti-Ly6G mAb administration (Figure 8A). The reduction in the neutrophil number led to the recovery of the IFN- γ production and a decrease in IL-10 production (Figure 8B). Notably, a decrease in the inflammation area was observed after anti-Ly6G mAb treatment in Mtb-infected C3H/HeJ mice (Figure 8C). Consistent with the reduction in the size of the inflamed area, the bacterial burden decreased significantly in response to anti-Ly6G mAb treatment after Mtb infection (Figure 8D).

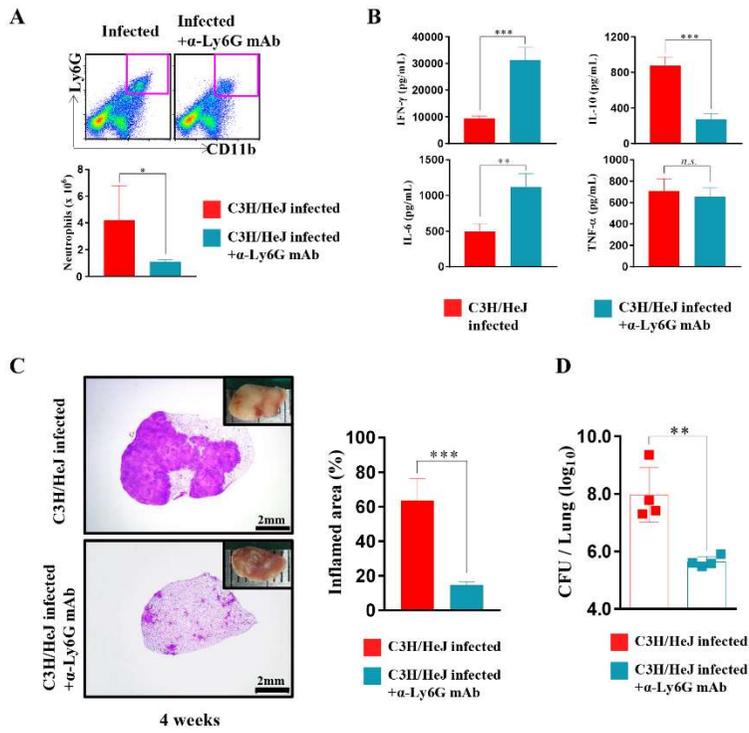


Figure 8. Effect of neutrophil depletion by treatment with an anti-Ly6G monoclonal antibody on bacterial burden and inflammation in C3H/HeJ mice. (A) Lung cells from infected C3H/HeJ and C3H/HeJ mice treated with an anti-Ly6G mAb were stained with the indicated markers, and the number of neutrophils at 4 weeks postinfection is shown as a bar graph. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value <0.05 was considered statistically significant; **p* < 0.05 . (B) At four weeks postinfection, mice from each group were euthanized, and their lung cells (1×10^6 cells) were stimulated with ESAT-6 ($1 \mu\text{g/mL}$) for 12 hours at 37°C . The concentration of IFN- γ , IL-6, TNF- α , and IL-10 in the

supernatant was measured by ELISA. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant; *n.s.*: not significant. (C) The superior lobe of the right lung stained with H&E at 4 weeks after Mtb K strain challenge. The gross lung pathology is shown with H&E staining. The data represent the percentages of the superior lobe of the right lung that showed inflammation and are presented as a bar graph. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant; ****p* < 0.001 . (D) The CFUs in the lungs of each group at 4 weeks postinfection were analyzed by counting the bacteria. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant; ***p* < 0.01 .

5. Blockade of the IL-10 receptor restores the protective Th1 response in the lungs of C3H/HeJ mice

To better understand the processes contributing to the well-controlled bacterial burden, we analyzed inflammatory cytokine secretion levels at 3 and 4 weeks postinfection. We examined the cytokine secretion of the lung cells and the number of CD4⁺IFN- γ ⁺ T cells after ESAT-6 stimulation. CD4⁺IFN- γ ⁺ T cells and IFN- γ production recovered, but the number of CD4⁺CD44⁺ T cells, which are activated T cells, did not differ from that in C3H/HeJ mice at 3 and 4 weeks postinfection after IL-10 receptor-blocking mAb treatment (Figure 9A). The production of immunosuppressive IL-10 did not decline at 3 and 4 weeks postinfection, but the secretion of IFN- γ recovered (Figure 9B). These data suggest that IL-10 signaling is associated with the reduced production of IFN- γ , and IL-10 plays a role in regulating Mtb in the absence of TLR4 signaling.

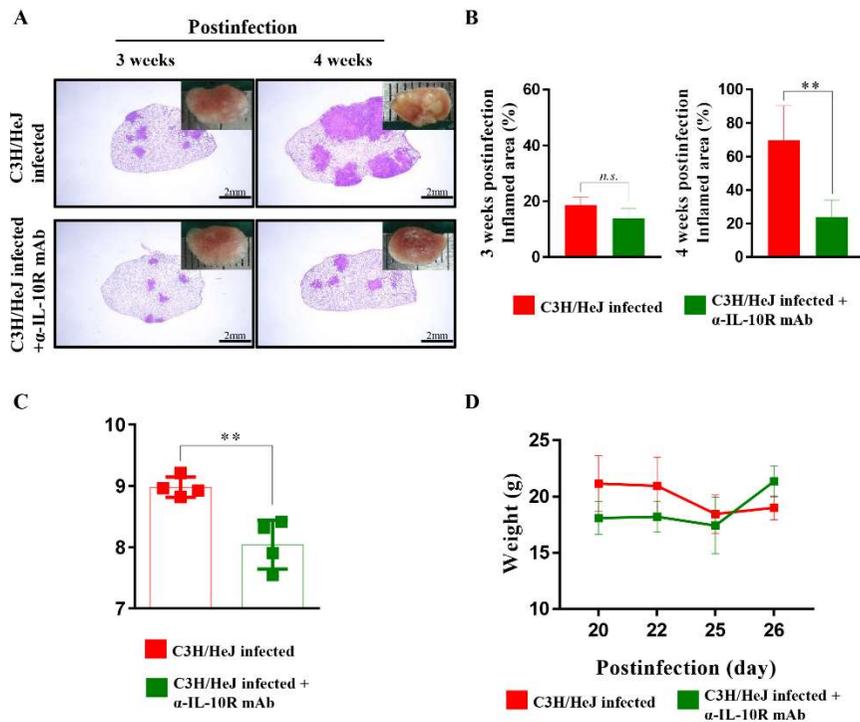


Figure 9. Effect of IL-10 receptor blockade on the protection against Mtb K infection in C3H/HeJ mice. C3H/HeJ and C3H/HeJ mice intraperitoneally administered an anti-IL-10 receptor mAb weekly 2 weeks postinfection were infected with 150 CFU of the Mtb K strain via aerosolization. Subsequently, the lungs were removed at 3 and 4 weeks postinfection for analysis. (A) The superior lobe of the right lung stained with H&E at 3 and 4 weeks after Mtb K strain challenge. The gross lung pathology is shown by H&E staining. (B) The inflamed area of the H&E-stained samples was quantified as the percentage and presented in bar graphs. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value $<$ 0.05 was considered

statistically significant; *n.s.*: not significant and $**p < 0.01$. (C) The CFUs in the lungs of each group at 4 weeks postinfection were analyzed by counting the bacteria. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant; $**p < 0.01$. (D) Changes in body weights were analyzed from 20 to 27 days after Mtb K strain infection.

6. Blockade of the IL-10 receptor restores the protective Th1 response in the lungs of C3H/HeJ mice

To better understand the processes contributing to the well-controlled bacterial burden, we analyzed inflammatory cytokine secretion levels at 3 and 4 weeks postinfection. We examined the cytokine secretion of the lung cells and the number of CD4⁺IFN- γ ⁺ T cells after ESAT-6 stimulation. CD4⁺IFN- γ ⁺ T cells and IFN- γ production recovered, but the number of CD4⁺CD44⁺ T cells, which are activated T cells, did not differ from that in C3H/HeJ mice at 3 and 4 weeks postinfection after IL-10 receptor-blocking mAb treatment (Figure 6A). The production of immunosuppressive IL-10 did not decline at 3 and 4 weeks postinfection, but the secretion of IFN- γ recovered (Figure 6B). These data suggest that IL-10 signaling is associated with the reduced production of IFN- γ , and IL-10 plays a role in regulating Mtb in the absence of TLR4 signaling.

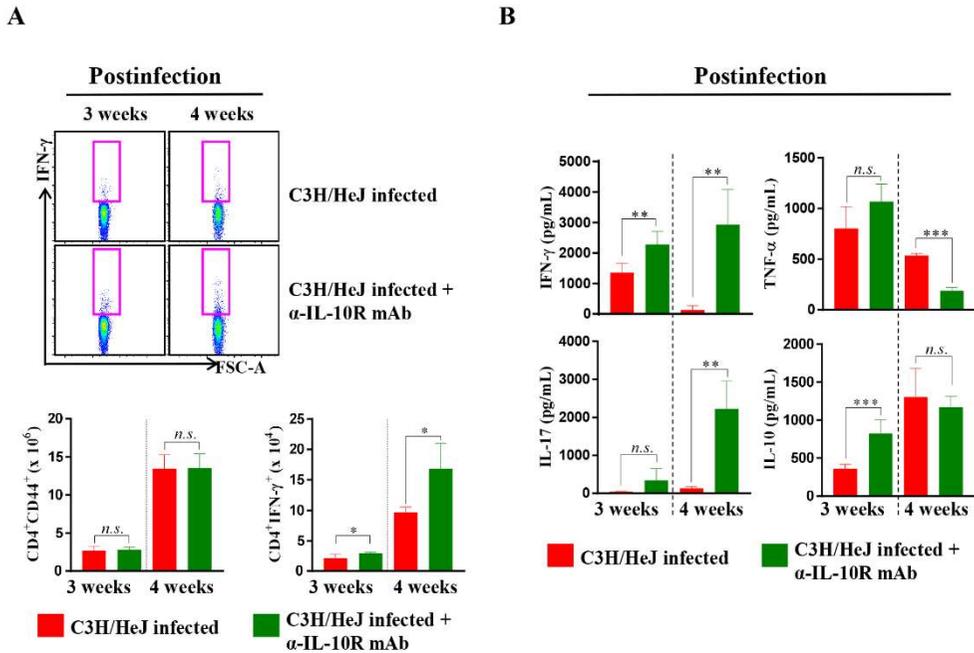


Figure 10. Effect of IL-10 receptor blockade in C3H/HeJ mice on the antigen-specific IFN- γ response and cytokine production in lung cells. Mice from each group were euthanized, and their lung cells (1×10^6 cells) were stimulated with ESAT-6 ($1 \mu\text{g/mL}$) for 12 hours at 37°C in the presence of brefeldin A and monensin. (A) At 3 and 4 weeks postinfection, the IFN- γ -secreting CD4⁺ T cells in the C57BL/6J, C3H/HeN and C3H/HeJ groups of mice were analyzed by flow cytometry, and the results are presented as bar graphs. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value <0.05 was considered statistically significant; *n.s.*: not significant and $*p < 0.05$. (B) IFN- γ , IL-17AF, TNF- α , and IL-10 concentrations in the supernatants were measured by ELISA. The data are presented as the mean \pm SD

of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value <0.05 was considered statistically significant; *n.s.*: not significant, ***p* < 0.01, and ****p* < 0.001.

7. Blockade of the IL-10 receptor retains the original CD11b^{hi}Ly6G^{hi} neutrophil phenotype, whereas the CD11b^{int}Ly6G^{int} neutrophil phenotype emerges in infection-controlled C3H/HeJ mice following Mtb infection

Next, we investigated the number of neutrophils since the blockade of the IL-10 receptor recovered the number of IFN- γ -producing CD4⁺ T cells (Figure 6), similar to the outcome of neutrophil depletion (Figure 4) in C3H/HeJ mice. As expected, the frequency of CD4⁺ T cells was restored, but the number of CD11b⁺Ly6G⁺ neutrophils unexpectedly increased in C3H/HeJ mice treated with the IL-10 receptor-blocking mAb at 4 weeks after Mtb infection (Figure 7A). We postulated that the increased population of neutrophils in C3H/HeJ mice treated with the IL-10 receptor-blocking mAb might display different phenotypes from those in C3H/HeJ mice infected with Mtb only. Intriguingly, C3H/HeJ mice treated with the IL-10 receptor blocking mAb showed a lower frequency and proportion of CD11b^{int}Ly6G^{int} cells at 4 weeks postinfection than the infection control group mice (Figures 7B,C). In other words, C3H/HeJ mice mostly had CD11b^{hi}Ly6G^{hi} cells considered neutrophils at 3 weeks postinfection, but CD11b^{int}Ly6G^{int} cells emerged as neutrophils at 4 weeks postinfection. Notably, the original CD11b^{hi}Ly6G^{hi} cells that appeared at 3 weeks postinfection in C3H/HeJ mice were retained at 4 weeks postinfection without phenotypic alteration (Figure 7D), suggesting that the disproportionate increase in CD11b^{int}Ly6G^{int} neutrophils from CD11b^{hi}Ly6G^{hi} cells may lead to the pathologic consequences in C3H/HeJ mice.

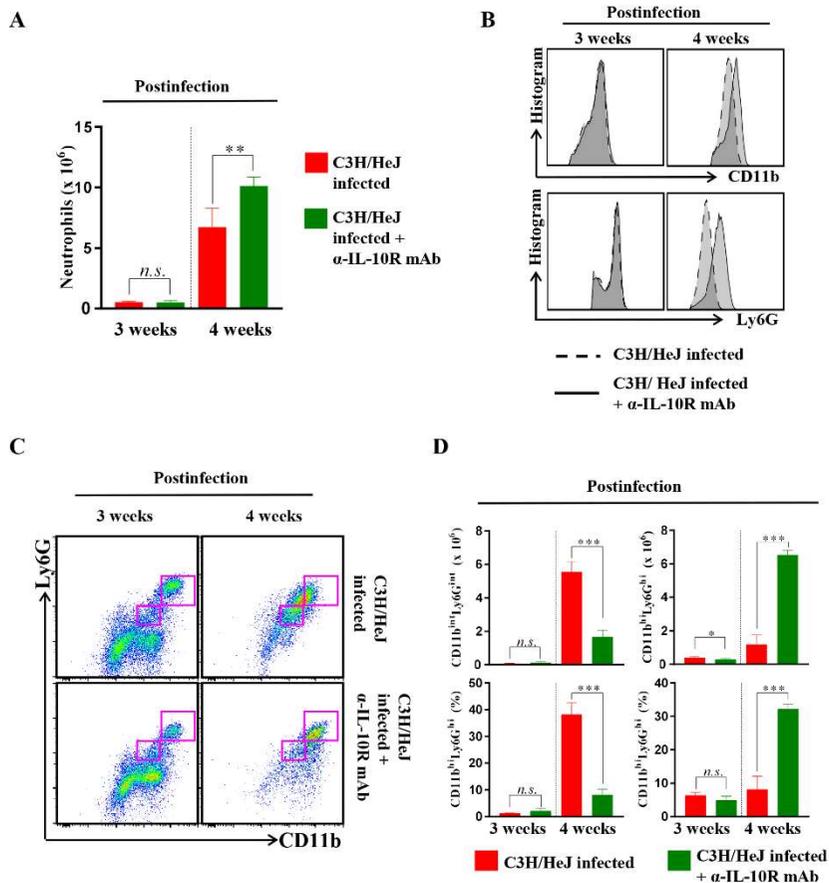


Figure 11. Analysis of neutrophil phenotype alterations produced by IL-10-receptor blocking in the lungs of C3H/HeJ mice. The mice in each group were sacrificed at 3 and 4 weeks postinfection, and their lung cells were analyzed by flow cytometry. (A) Lung cells from infected C3H/HeJ and C3H/HeJ mice treated with an IL-10 receptor-blocking mAb were stained with the indicated markers, and the neutrophil populations are shown as bar graphs at 3 and 4 weeks postinfection. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05

was considered statistically significant; *n.s.*: not significant and $**p < 0.01$. (B) The expression levels of CD11b and Ly6G in the neutrophil population (CD11b⁺Ly6G⁺) were analyzed by flow cytometry and are shown as histograms. (C) Neutrophil subpopulations (CD11b^{int}Ly6G^{int} and CD11b^{hi}Ly6G^{hi}) were gated according to the expression levels of CD11b and Ly6G. (D) Bar graph of CD11b^{int}Ly6G^{int} and CD11b^{hi}Ly6G^{hi} cells at 3 or 4 weeks postinfection. The data are presented as the mean \pm SD of four-to-five mice in each group. The significance of differences was determined with an unpaired *t*-test. A *p* value < 0.05 was considered statistically significant; *n.s.*: not significant, $*p < 0.05$, and $***p < 0.001$.

DISCUSSION

In this study, we attempted to elucidate the function of TLR4 during Mtb infection because its involvement in protective immune responses against Mtb infection remains controversial. The data in our current study support the hypothesis that TLR4 is required for an optimal protective Th1 response against Mtb infection, which is in good agreement with previous reports. In addition, we further found that Mtb infections were uncontrolled in TLR4-deficient mice due to abundant IL-10 production and an uncontrolled influx of neutrophils into the Mtb-infected lungs. The blockade of the IL-10 receptor and depletion of neutrophils restored the protective Th1 immune response against Mtb infection in the absence of TLR4. Therefore, our results clearly demonstrate that the susceptibility of TLR4-deficient C3H/HeJ mice to Mtb infection is caused by excessive IL-10 secretion and neutrophil recruitment into the lung, resulting in the disruption of the optimal host Th1 immune response.

Recent clinical studies of TLR2, TLR4, and TLR9 polymorphisms have reported their correlation with TB susceptibility in humans. In particular, the Asp299Gly mutation of TLR4 is known to increase susceptibility to TB because of a reduced CD4⁺ T cell frequency. Additionally, the Asp299Gly and Thr39Ile mutations in TLR4 are associated with increased bacillary load in the sputum and severe forms of TB in chest radiographs of TB patients. Additionally, *TLR2*, *IL-17*, and *TNFA* polymorphisms along with *TLR4* polymorphisms are associated with the TB infection risk. Although these clinical data support the hypothesis that various mutations in TLR4 are closely

associated with TB infection risk and TB progression, the immunological role of TLR4 in TB protection remains unclear.

Interestingly, previous studies conducted by different groups were performed using TLR4-competent C3H/HeN and TLR4-deficient C3H/HeJ mice, which were used in our current study. However, the outcomes between TLR4-competent C3H/HeN and TLR4-deficient C3H/HeJ mice infected with Mtb are clearly inconsistent among different researcher groups. Shim *et al.* and Reiling *et al.* reported that TLR4-competent C3H/HeJ and TLR4-deficient C3H/HeN mice infected with a low dose (50-100 CFU per mouse) of aerosolized Mtb H37Rv showed no significant differences over 100 days postinfection. Abel *et al.* also performed experiments under conditions similar to those of these two studies: (1) TLR4-competent C3H/HeN and TLR4-deficient C3H/HeJ mice were used, (2) the same Mtb strain H37Rv was used for the challenge, (3) the infection was carried out via aerosolization, and (4) the infection doses ranged from 50 to 100 per mouse. However, Abel *et al.* reported that C3H/HeJ mice showed a more severe TB disease phenotype than C3H/HeN mice, as demonstrated by a significantly increased bacterial load and lung inflammation⁴⁸. One putative reason for the controversial outcomes may be the virulence of the Mtb strain. Even in the same H37Rv Mtb strain, the virulence is altered by the batch or subculture via laboratory adaption processes⁵³. The pro- and anti-inflammatory cytokine secretion profile induced by TLR signaling is distinct depending on the strain and lineage of Mtb³³. The Mtb K strain is one of the causative strains of TB outbreaks and

is highly virulent¹⁴. We expected that infection with Mtb K might reveal the role of TLR4 because the Mtb K strain is associated with TB outbreaks, inducing TB progression more efficiently than the laboratory strain H37Rv^{54,55}. For these reasons, in our study, infection with 150 CFU of the Mtb K strain caused severely inflamed lesions and greater bacterial burden in TLR4-deficient mice than TLR4-expressing mice (Figure 1). The second reason for the conflicting results could be the production of IL-10 and Foxp3⁺ regulatory T cells induced by hypervirulent Mtb strains, such as HN878 and K, in the mouse model^{54,56-58}. BMDMs differentiated from TLR4-deficient mutant mice produced more IL-10 after Mtb strain HN878 infection than after H37Rv infection³³. Similarly, increased Foxp3⁺ regulatory T cell frequency and IL-10 production have been reported in TB patients⁵⁹. Thus, IL-10 production by hypervirulent Mtb strains may be associated with the different outcomes of TLR4-competent and TLR4-deficient mice during Mtb infection. In addition, infection with a high dose of the Edrman Mtb strain by intravenous administration (1×10^5 CFU), or the intranasal administration (1×10^5 CFU) of Mtb strain H37Rv produced higher bacterial burdens in TLR4-deficient C3H/HeJ mice than TLR4-competent C3H/HeN mice along with severe lung inflammation after 2 weeks of infection^{49,60}. The populations of CD4⁺IFN- γ ⁺ T cells, which are essential for protection against Mtb infection, were decreased from 1 to 4 weeks postinfection in TLR4-deficient mice compared to TLR4-competent mice according to Chackerian *et al*⁶⁰, which is consistent with our results. These differences between different studies also seem to be due to the infection dose and infection route. The host immune response may be

changed depending on the infection route⁶¹; for example, mice are more susceptible to Mtb infection by aerosolization than intravenous administration⁶².

Neutrophils are immune cells that are involved in early host protection against Mtb infection in the lung^{63,64}. These cells circulate in blood vessels and rapidly reach the infection site by following a gradient of CXCL8 or keratinocyte chemoattractant and kill pathogens⁶⁵. However, other studies have shown that neutrophils contribute to the progression of TB pathogenesis^{66,67}. In the cavity and sputum of active TB patients, neutrophils are major phagocytic cells that engulf Mtb bacilli⁶⁶. The absence of CCR2 or CCL2, which are involved in the egress of neutrophils from the bone marrow and migration to tissue, has been shown to effectively control Mtb^{68,69}. In our study, a number of neutrophils infiltrated the lung, and a wide range of necrotic lung lesions were found in TLR4-deficient C3H/HeJ mice (Figures 1,2), consistent with the results of previous studies^{48,49}. In addition, the depletion of the increased neutrophils after Mtb infection significantly alleviated the bacterial burden and lesion inflammation (Figure 4). Thus, further study is necessary to investigate the functional role of infiltrated neutrophils in TLR4-deficient C3H/HeJ mice.

Interestingly, the accumulation of neutrophils represents the disease severity and progression of TB in various TB-susceptible mouse models, including genetically susceptible mice such as IFN- γ -knockout mice, TLR2-knockout mice, caspase recruitment domain-containing protein 9-knockout mice, immune-responsive gene 1-

knockout mice, and inbred mice such as I/St, C3HeB/FeJ, DBA/2, and CBA/J strains of mice. Neutrophils and polymorphonuclear-myeloid-derived suppressor cells (PMN-MDSCs) have similar phenotypical markers, CD11b⁺Ly6G⁺⁷⁰. IL-10 production by MDSCs suppresses T cell responses, especially IFN- γ production by CD4⁺ T cells^{71,72}. Recently, an increasing number of studies have reported that neutrophils are associated with the progression of TB, but the correlation of TB with MDSCs is unknown. Increased survival and reduced inflammation were observed in Mtb-infected C3H/HeJ mice after the depletion of neutrophils via anti-Ly6G mAb treatment. As shown in previous studies, the removal of neutrophils in TLR4-deficient mutant mice attenuated the lesion inflammation, reduced the Mtb burden and IL-10 production, and recovered the IFN- γ production in our study (Figure 4). Neutrophils are a source of IL-10 production¹, and the depletion of neutrophils by anti-Ly6G mAb treatment was accompanied by a decrease in IL-10 production (Figure 3). Therefore, the increased IL-10 production through neutrophils in TLR4-deficient mice might impair the immune response against Mtb infection. The decrease in neutrophil numbers might be associated with reduction of IL-10 production because neutrophils are one of the sources of IL-10⁷³. CD11b⁺Ly6G⁺ neutrophils in TLR4-deficient mice might play a similar role as MDSCs in Mtb infection, resulting in an immunosuppressive effect. IL-10 production via the markedly increased recruitment of neutrophils in TLR4-deficient mice could aggravate the prognosis of Mtb infection. Therefore, the removal of neutrophils could contribute to the control of Mtb infection^{13,74}.

IL-10 is an anti-inflammatory cytokine that inhibits the activity of immune cells such as Th1 cells, NK cells, and macrophages, which are essential for Mtb control⁷⁵. In humans and mouse models, IL-10 has been discussed as one of the factors that promote TB progression⁷⁶. The level of IL-10 was found to be elevated in the serum of TB patients with increased bacillary loads^{20,77}. In other studies, the neutralization of endogenous IL-10 was found to elevate Th1 responses in peripheral blood mononuclear cells (PBMCs) from TB patients⁷⁸⁻⁸⁰. Previous studies have suggested that IL-10 suppresses the host protective Th1 immune response against Mtb infection, eventually leading to TB progression. In addition, similar to the results of our study, the blockade of IL-10 signaling in the Mtb-susceptible strain CBA/J has been shown to decrease the bacterial burden and restore the CD4⁺ and CD8⁺ T cell frequency³⁰. In our study, significantly increased IL-10 production was observed in lung cells from antigen-stimulated TLR4-deficient mice at 4 weeks postinfection but not at 3 weeks postinfection (Figure 3C). The blockade of the IL-10 receptor led to a significant reduction in bacterial burden (Figure 5) and the restoration of the Th1 response (Figure 6). The results of previous studies and our results confirm that the control of Mtb growth by IL-10 is disturbed and that the removal of IL-10 signaling enhances the protection against Mtb infection in the absence of TLR4 signaling. Furthermore, the depletion of neutrophils markedly enhanced the protection against Mtb infection and recovered the Th1-type T cell response (Figure 4). Thus, we focused on the number of neutrophils rather than other immune cells, such as macrophages or

dendritic cells. TLR4-deficient mice with blocked IL-10 signaling showed a well-controlled Mtb infection with an increased number of neutrophils (Figure 7A). In the current study, we found that the blockade of IL-10 signaling in TLR4-deficient mice appeared to inhibit the alteration of the conventional neutrophil phenotype, from CD11b^{hi}Ly6G^{hi} to CD11b^{int}Ly6G^{int} (Figures 7B-D). Among the cells expressing Ly6G and CD11b, Gr-1^{dim}Ly6G^{dim}CD11b⁺-expressing cells inhibit T cell proliferation and IFN- γ production⁸¹. We hypothesized that CD11b^{int}Ly6G^{int} cell accumulation in TLR4-deficient mice could suppress the Th1-type T cell response. The phenotypic maintenance of neutrophils caused by the blockade of the IL-10 receptor may contribute to the restriction of the progression of TB. However, functional studies of the differences between CD11b^{hi}Ly6G^{hi} cells and CD11b^{int}Ly6G^{int} should be conducted, particularly by focusing on the effect on T cell-mediated immune responses.

In summary, our results suggest that TLR4 is necessary to elicit optimal protection against Mtb infection via the regulation of IFN- γ , IL-10 production, and neutrophil recruitment during Mtb infection. the disruption of IL-10 signaling and depletion of neutrophils contribute to restrict Mtb growth in TLR4-deficient mice and enhance the IFN- γ -producing CD4⁺ T cell response. Although the alteration of neutrophil phenotypes by IL-10 was associated with TB progression, it did not affect the function after Mtb infection. These findings could provide effective targets and immunological approaches for understanding TB pathogenesis.

V. CONCLUSION

According to World Health Organization, tuberculosis (TB), caused by *Mycobacterium tuberculosis* (Mtb), is the largest infectious disease cause leading human death in the world. Toll-like receptors (TLRs) are required for host protection against Mtb infection. However, the role of TLR4 is still unclear. In this study, we investigated the role of TLR4 during Mtb infection. TLR4-deficient mutant mice showed increase in bacterial burden along with necrotic lung inflammation compared to TLR4-competent mice. In addition, increased influx of neutrophil and IL-10 production in the lung of TLR4-deficient mice. Reduction of neutrophil and IL-10 signal blockade could contribute to control Mtb growth via restored Th1-type T cell response in TLR4-deficient mice. Therefore, IL-10 signal and neutrophil may be detrimental role in the absence of TLR4 signal during Mtb infection. Interestingly, neutrophil influx increased in TLR4-deficient mice after IL-10 blockade. TLR4-deficient mice with blocked IL-10 signaling displayed different neutrophil phenotype after Mtb infection. We found that phenotypical alteration of neutrophil was inhibited after IL-10 blockade. Therefore, the phenotypical maintenance of neutrophils could contribute to improve the host protection against Mtb infection. These results indicated that TLR4 plays a critical role to investigate the effective control against Mtb infection.

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ABSTRACT (IN KOREAN)

결핵균을 방어하기 위한 최적화된 면역반응을
매개하는 톨-유사 수용체4에 대한 연구

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박재훈

결핵균(*Mycobacterium tuberculosis*, Mtb)는 전 세계 인구의 30% 이상이 이미 감염되어 있으며, 결핵을 일으키는 병원성 세균이다. 세계보건기구에 따르면 매년 960만 명의 새로운 결핵 환자와 150만 명이 결핵으로 사망한다. 결핵의 발병은 매년 감소하고 있지만 여전히 환자에게 높은 치료 비용의 부담과 부작용으로 인류에 큰 피해를 주고 있다. 이렇게 인류의 보건을 위협하는 결핵을 제어하기 위한 전 세계의 많은 연구들이 진행되고 있다. 하지만, 아직도 결핵이라는 질병의 진행되는 과정에 대한 원인은 명확히 알려지지 않았다.

숙주가 결핵균을 인지하고 방어하기 위한 수용체의 역할에 대해 알아보았다. 면역세포들의 표면에 위치한 수용체들은 최초로 결핵균을 인지하고 제어하기 위해 필수적이다. 톨-유사수용체(Toll-like receptor, TLR)은 선천 및 후천성 면역반응에서 결핵균을 인식하는데 중추적인 역할을 하는 것으로 알려져 있다. 이전 연구들에서 TLR4의 역할에 대한 결과는 아직 논란의 여지가 있다. 따라서, 실제 질병의 진행에 TLR4의 역할을 알아보기 위해 TLR4 결핍 마우스에 결핵균을 감염하여 면역세포의 구성과 T 세포의 반응을 분석하였다. TLR4 결핍 마우스는 매우 악화된 폐의 염증 및 박테리아의 수가 증가함을 보였다. 면역세포의 유입에도 호중구의 수가 제어되지 않고 다량이 발견되었으며 T 세포의 수는 매우 감소하였다. 결핵균을 제어하는데 중추적인 사이토카인인 IFN- γ 생산이 또한 매우 낮았으며 면역반응을 억제하는 IL-10 사이토카인의 생산이 유의적으로 증가했다. 증가한 호중구와 IL-10 신호를 제거하였을 때 숙주는 효과적으로 결핵균을 제어할 수 있었다. 특이적으로, IL-10 신호의 제거는 호중구의 증가 및 호중구의 표현형적 차이를 동반했다. 이러한 결과를 종합해 볼 때, TLR4는 결핵균을 제어하는데 필수적임을 알 수 있다.

본 연구는 결핵균을 제어하기 위해 면역세포의 초기 인식 및 성장 인자에 의한 면역세포의 구성을 알아보았다. 호중구 및 IL-10이 결핵균 감수성을 결정하는 중요한 요소임을 알 수 있다. TLR4 매개 반응의 활용은 개인 맞춤 치료 요법 혹은 백신과 같은 효과적인 제어 수단의 개발에 대한 새로운 목표가 될 것이다.

핵심되는 말: 결핵균, Toll-유사수용체, 호중구, T세포, IL-10