

Pathology of C3 Glomerulopathy

Su-Jin Shin, M.D., Ph.D.
Yoonje Seong*
Beom Jin Lim, M.D., Ph.D.

Department of Pathology, Yonsei University College of Medicine, Korea

*This author is a student of Yonsei University College of Medicine and contributed to this work during the sub-internship program 2019.

Corresponding author:

Beom Jin Lim, M.D., Ph.D.
Department of Pathology, Yonsei University College of Medicine, Gangnam Severance Hospital 211 Eonju-ro, Gangnam-gu, 06273, Seoul, Korea
Tel: +82-2-2019-3540
Fax: +82-2-3463-2103
E-mail: bjlim@yuhs.ac

Received: 14 September 2019
Revised: 7 October 2019
Accepted: 10 October 2019

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © 2019 The Korean Society of Pediatric Nephrology

C3 glomerulopathy is a renal disorder involving dysregulation of alternative pathway complement activation. In most instances, a membranoproliferative pattern of glomerular injury with a prevalence of C3 deposition is observed by immunofluorescence microscopy. Dense deposit disease (DDD) and C3 glomerulonephritis (C3GN) are subclasses of C3 glomerulopathy that are distinguishable by electron microscopy. Highly electron-dense transformation of glomerular basement membrane is characteristic of DDD. C3GN should be differentiated from post-infectious glomerulonephritis and other immune complex-mediated glomerulonephritides showing C3 deposits.

Key words: C3 glomerulopathy, C3 glomerulonephritis, Membranoproliferative glomerulonephritis, Dense deposit disease, Alternative complement pathway

Introduction

Glomerulonephritides due to alternative complement pathway dysregulation are presently categorized as C3 glomerulopathy¹⁾. However, the scope of C3 glomerulopathy is somewhat blurred for the following reasons: (i) Some entities, such as atypical hemolytic uremic syndrome (aHUS), may share alternative pathway dysregulation yet differ distinctly in clinicopathologic features²⁾; (ii) Various glomerulopathies may show predominance or co-dominance of C3 deposition (relative to other immunoglobulins or complement types) by immunofluorescence (IF), post-infectious glomerulonephritis (PIGN) being a prime example³⁾; and (iii) Laboratory testing for alternative pathway dysregulation is not widely available at present, leaving the current IF standard of measure as the sole determinant of C3 glomerulopathy⁴⁾. Although this subject is complex, the historical aspects of C3 glomerulopathy, its pathogenesis, and pertinent renal biopsy findings are addressed herein.

Historical aspects of disease classification

C3 glomerulopathy is a relatively new disease class established by international consensus in 2013⁵⁾. Prior to this time, little was known of the related pathophysiology, so many cases previously diagnosed as membranoproliferative glomerulonephritis (MPGN) on morphologic grounds would now qualify as C3 glomerulopathy. Membranoproliferative is a descriptive term, refer-

ring to the most common light microscopic finding in such instances and signifying that both glomerular basement membrane (GBM) and related cellular components (mesangial, endothelial, and endocapillary) are duly altered⁶.

Repeated past efforts to subclassify MPGN have gradually advanced the understanding and optimal treatment strategies for each subgroup. Despite the impression of homogeneity by light microscopy (LM), three subgroups of MPGN (I–III) emerged, defined by electron microscopy (EM). Subendothelial deposits were characteristic of MPGN type I⁷; and dense deposit disease (DDD), named for its unique electron-dense transformation of GBM, corresponded with MPGN type II⁸. MPGN type III included two distinct forms: the Burkholder variant, showing additional subepithelial immune deposits⁹; and the Strife and Anders variant, demonstrating thickened glomerular capillary walls and destruction of GBM by intramembranous deposits^{10,11}. MPGN type II (or DDD) was deemed the most distinctive of these subgroups, given its hallmark EM findings. However, studies conducted during the 2000's revealed that membranoproliferative patterns by LM were confined to a minority (25–44%) of patients with DDD^{12,13}, suggesting substantial differences in the pathophysiology of DDD versus other types of MPGN.

In addition to LM and EM features, IF studies have provided critical etiologic clues in this setting fueling the C3 glomerulopathy disease concept. Early investigations from the 1970's had delineated two distinct IF patterns in instances of MPGN^{7,14,15}, one of which glomerular deposits of immunoglobulin and C3 both occur and the other is limited to isolated C3 deposition. The former was explained by triggering of complement activation via classical (immune complex-based) or lectin (foreign substance-induced) pathways, whereas isolated C3 deposition was attributed to alternative pathway complement activation.

Fakhouri et al. subsequently introduced C3 glomerulopathy as a new disease class imposed by isolated C3 glomerular deposition (in the absence of immunoglobulin), with or without visible membranoproliferative features¹⁶. They also defined two subsets of C3 glomerulopathy: DDD bearing dense osmiophilic deposits of mesangium, GBM, and tubular basement membrane and C3 glomerulonephritis (C3GN), a relatively rare and heterogeneous variant with less discrete EM deposits. Although morpho-

logically disparate, DDD and C3GN are similar in etiology and fall within the same disease spectrum¹⁶.

Countless efforts to reach an international consensus on the C3 glomerulopathy paradigm finally bore fruit during the first C3 Glomerulopathy Meeting held in August, 2012⁵. This gathering of experts in nephrology, renal pathology, complement biology, and complement therapeutics sought to provide a formal definition of the disease and establish guidelines for future research⁵. Unlike the criteria first proposed, they described C3 glomerulopathy as “a disease process due to abnormal control of complement activation, deposition, or degradation and characterized by predominant glomerular C3 fragment deposition with electron-dense deposits in EM”. This description implies that small amount of immunoglobulin deposits can be observed in C3 glomerulopathy and that diagnosis of C3 glomerulopathy must rely on IF, LM, EM, and clinical features⁵.

Pathophysiology of C3 glomerulopathy

As an important innate defense mechanism against pathogenic stimuli, the complement system includes >30 components and regulators¹⁷. There are three complement activation pathways: classical, lectin, and alternative. The classical pathway, incorporating C1q, C1r, C1s, C4, and C2 components, is activated by antigen-antibody immune complexes. The lectin pathway, in which C2 and C4 also have roles, is activated by attachment of mannose-binding lectin to microbial carbohydrate groups^{17,18}. The alternative pathway is unique in its continual activation by spontaneous C3 hydrolysis, forming C3 convertase (so-called “tick-over” process)¹⁹. Generation of C3 convertase and its downstream products is tightly regulated by various components of the complement system, so any inherent defects or autoantibody assaults may eventuate in alternative pathway dysregulation^{17,20}. The mechanism of alternative pathway dysregulation in the context of C3 glomerulopathy is highly complex, as is regulating the complement system itself. Factors contributing to this dysfunction may be categorized by mechanism of action.

1. Autoantibodies to alternative pathway regulators

C3 nephritic factors (C3NeFs) are IgG or IgM autoanti-

bodies targeting neopeptides of alternative pathway C3 convertase (C3bBb)^{21,22}. C3bBb converts C3 to C3b in conjunction with complement factor B (FB) and complement factor D; and in an amplification loop, factor B attached to C3b converts more C3 to C3b²³. C3 convertase is degraded very quickly by negative regulators, such as decay-accelerating factor (CD55), complement factor H (FH), and complement receptor 1, thus preventing uncontrolled tissue injury. C3NeFs prolong the half-life of C3bBb through interference with these regulators²⁴⁻²⁶. They are also found in many patients with DDD (70–80%) or C3GN (40–55%)²⁶. C5 nephritic factors (C5NeFs) are autoantibodies that stabilize C5 convertase (C3bBbC3b) of the alternative pathway, thereby affecting serum C5b-9 levels. Marinozzi et al. have demonstrated C3NeFs in more than half of patients with C3GN, although C3NeFs and C5NeFs in combination or C5NeFs alone may be present; and C5NeFs are more frequently associated with C3GN than with DDD²⁷. C4 nephritic factors (C4NeFs) stabilize the C3 convertase (C4bC2a) of classical and lectin pathways. There is evidence of sporadic C4NeF positivity in patients with C3GN, although the recorded prevalence varies^{28,29}. The functions of these nephritic factors often require the presence of properdin^{17,26,27}. In addition to previous therapeutic trials like plasmapheresis, anti-cellular immune therapy and eculizumab⁵, properdin is currently under scrutiny for its therapeutic utility³⁰. In addition to nephritic factors, autoantibodies aimed at other regulators or components of the complement pathway (i.e., FH and FB) have been detected^{31,32}.

2. Genetic alterations

Apart from acquired causes of C3 glomerulopathy thus far discussed, aberrant complement-related genes are similarly implicated, including variants of CFH (affecting FH), CFI (affecting factor I), CFB, and C3 (affecting C3 convertase and C5 convertase)^{1,20}. Factor H-related (FHR) proteins are other modulators of the complement pathway, some members (FHR1, FHR2, FHR3, and FHR5) forming dimeric complexes under normal conditions. Abnormal FHR fusion proteins due to genetic alterations may result in C3GN or DDD^{1,33}. The above genetic defects cited for C3GN largely overlap with those of aHUS. However, aHUS is marked by solid-phase endothelial cell activation of the

alternative complement pathway, unlike the largely fluid-phase alternative pathway activation of C3GN^{18,34,35}. Alternative pathway dysregulation may also stem from monoclonal immunoglobulin production in patients with monoclonal gammopathy of renal significance, multiple myeloma, and other lymphoproliferative disorders^{36,37}. Under such circumstances, monoclonal proteins act as autoantibodies targeting FH or FB and then culminate in C3 glomerulopathy (monoclonal immunoglobulin-associated C3 glomerulopathy)³⁸.

Pathologic features of C3 glomerulopathy

C3 glomerulopathy is typified by C3 accumulation deposited within glomerular mesangium and capillary walls. Little or no immunoglobulin is present. IF studies confirm an active glomerulonephritis, showing mostly granular or semi-linear C3 deposits more intensely stained (2-level at least) than other complement or immunoglobulin proteins. IgG, IgM, IgA, and C1q deposits may well be present, but not at levels comparable to C3 (Fig. 1A, B), and C4d deposition is rarely seen^{5,39}.

Electron-dense glomerular deposits are identified in all forms of C3 glomerulopathy and serve to differentiate the two major subtypes of C3 glomerulopathy: DDD and C3GN⁵. DDD is defined by highly electron-dense and osmiophilic intramembranous deposits and resultant GBM lamina densa transformation. The deposits appear ribbon- or sausage-like and may even inhabit tubular basement membrane or Bowman's capsule. In patients with C3GN, these deposits are ill-defined and not as highly electron-dense, involving mesangium and subendothelial regions (Fig. 1C, D). Rarely are they subepithelial or intramembranous in nature. They resemble those commonly seen in immune complex-mediated glomerulonephritis. So-called "hump-like" subepithelial deposits, similar to those manifested in PIGN, may be identified at times.

LM findings of C3 glomerulopathy are diverse, ranging from near-normal morphology to features of proliferative (mesangial, membranous, or endocapillary), crescentic, or sclerosing glomerulonephritis. The pattern witnessed in C3GN is usually that of MPGN (Fig. 1E, F)⁴⁰⁻⁴³, characterized by mesangial expansion with hypercellularity and

thickened (i.e., doubly contoured) glomerular capillary walls. In one series, MPGN patterns accounted for 71% of patients with C3GN⁴². The others primarily showed mesangial proliferation, without changes in glomerular capillaries. Some will lack mesangial proliferation altogether, mimicking minimal change disease. The glomeruli may show endocapillary proliferation, with variable inflammatory cell infiltrates, and crescents may be encountered^{5,42,44}.

In a large series of patients with DDD, the most common histologic pattern was mesangial proliferative GN (43.4%), followed by membranoproliferative (24.6%), crescentic (17.4%), and endocapillary proliferative (11.6%) glomerulonephritis¹³. Other cohorts have most often displayed features of MPGN (43.8–77.8%), followed by mesangial proliferative glomerulonephritis^{12,45,46}.

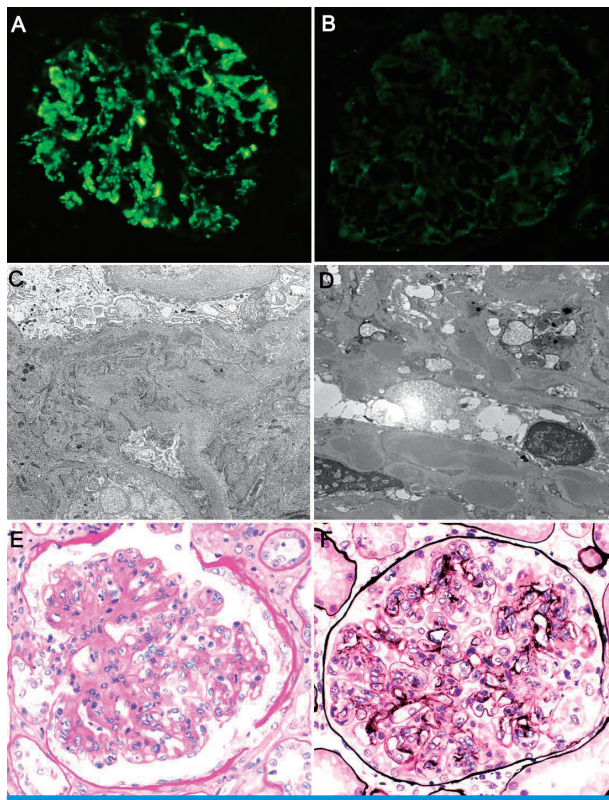


Fig. 1. Typical morphologic features of C3 glomerulopathy: Immunofluorescence preparations show (A) strong granular staining of glomerular C3 deposits (within mesangium and along capillary walls) as well as (B) trace IgG. Electron microscopic examination confirms multiple electron-dense deposits of (C) mesangium ($\times 6,000$) and (D) subendothelial regions ($\times 7,000$). Light microscopy displays (E) mesangial proliferation (Periodic acid-Schiff, $\times 400$) and (F) double-contoured capillary walls (methenamine silver, $\times 400$), indicating membranoproliferative glomerulonephritis.

Differential diagnosis of C3 glomerulopathy

1. C3 glomerulonephritis versus dense deposit disease

DDD is sometimes difficult to distinguish from C3GN^{5,47}. As noted earlier, C3 glomerulopathy in the absence of highly electron-dense, ribbon-like deposits by EM is classifiable as C3GN; and the electron-dense deposits of C3GN are more often found in mesangial and subendothelial regions, seldom occupying the intramembranous and subepithelial areas typical of DDD⁴⁸. Assessments of deposit density are subjective, frequently sparking disagreement among pathologists in their classification of DDD vs C3GN^{5,47,49-52}. Furthermore, the ribbon-like deposits of DDD are generally discontinuous, creating the potential for sampling error and subsequent misdiagnosis as C3GN. There are no other specific LM criteria for differentiating between DDD and C3GN, except when intramembranous deposits stain intensely by Periodic acid-Schiff^{8,53}. Such deposits are fuchsinophilic in trichrome stains and lack methenamine silver positivity⁵³.

The ratio of DDD to C3GN is reportedly about 1:3^{42,54}. It may be necessary to distinguish between DDD and C3GN for prognostic or therapeutic studies that compare respective outcomes. There is greater likelihood of C3NeFs⁴² and lower circulating C3 levels in patients with DDD (vs C3GN)⁵⁴. Otherwise, no clear clinical differences are evident.

2. Immune complex-mediated glomerulonephritis

By definition, glomerular C3 deposition in patients with C3 glomerulopathy is at least 2-level more intensely stained in IF preparations than are other complement or immunoglobulin deposits. However, the distinction between C3 glomerulopathy and MPGN is not always obvious on this basis. In terms of the latter, glomerular staining of C4d (a byproduct of activated classical and lectin pathways) may be helpful⁵⁵. C4d staining is negative or trace in instances of C3 glomerulopathy, whereas immune complex-mediated glomerulonephritis yields positive results⁵⁵⁻⁵⁷.

3. Post-infectious glomerulonephritis

PIGN is a self-limited glomerulonephritis commonly triggered by streptococcal infection⁵⁸ and reflecting a bacterial immunologic response. The findings are those of immune complex-mediated disease. Diffuse endocapillary

glomerulonephritis, deposition of IgG and C3, and subendothelial and subepithelial hump-like deposits are usually visible by LM, IF, and EM, respectively.

Difficulty may arise in distinguishing some cases of PIGN from C3GN. C3 amplification may outlast immunoglobulin, and C3-predominant or isolated C3 deposits may exist⁵⁹. C3GN may then sporadically present as a diffuse endocapillary proliferative pattern^{5,48,54,60}, and hump-like subepithelial deposits are frequent accompaniments of both MPGN and C3GN⁴⁸. The clinical course and laboratory findings are perhaps helpful in this regard. Patients with PIGN readily recover baseline kidney function and laboratory profiles in a matter of weeks, resolving hematuria, proteinuria, and hypocomplementemia without therapeutic intervention. Prolonged proteinuria and low serum C3 levels are clinically indicative of chronic glomerulonephritis. However, initial PIGN presentations have seemingly morphed into C3 glomerulopathy^{57,61,62}, transformed (it now appears) by complement activation via alternative pathway⁶¹⁻⁶⁴. Various sources have also noted the presence of nephritis-associated plasmin receptor (NAPlr) in instances of PIGN with C3 glomerulopathy features^{63,65,66}. As shown by Sethi et al., most cases of PIGN plagued by persistent hematuria and proteinuria have involved underlying defects, either genetic mutations, autoantibodies, or both, that impact alternative complement pathway regulation⁵⁷. Before conceding a diagnosis of C3 glomerulopathy in patients with C3-predominant PIGN, persistent hypocomplementemia, and proteinuria or declining renal function, further investigation of the alternative complement pathway is consequently advised^{3,5}.

Conclusion

C3 glomerulopathy is a recently established disease entity pathogenetically linked to alternative complement pathway dysregulation. The histologic patterns entailed are diverse, membranoproliferative being the most common. Based on distinctive EM features, DDD and C3GN are specific subtypes of C3 glomerulopathy, both requiring differentiation from other glomerulopathies marked by a predominance of C3 deposition.

Conflict of interest

The authors declare that they have no conflicts of interest.

ORCID

Beom Jin Lim. <https://orcid.org/0000-0003-2856-0133>

Su-Jin Shin. <https://orcid.org/0000-0001-9114-8438>

References

1. Smith RJH, Appel GB, Blom AM, Cook HT, D'Agati VD, Fakhouri F, et al. C3 glomerulopathy - understanding a rare complement-driven renal disease. *Nat Rev Nephrol* 2019;15:129-43.
2. Baines AC, Brodsky RA. Complementopathies. *Blood Rev* 2017; 31:213-23.
3. Khalighi MA, Wang S, Henriksen KJ, Bock M, Keswani M, Meehan SM, et al. Revisiting post-infectious glomerulonephritis in the emerging era of C3 glomerulopathy. *Clin Kidney J* 2016;9:397-402.
4. Ito N, Ohashi R, Nagata M. C3 glomerulopathy and current dilemmas. *Clin Exp Nephrol* 2017;21:541-51.
5. Pickering MC, D'Agati VD, Nester CM, Smith RJ, Haas M, Appel GB, et al. C3 glomerulopathy: consensus report. *Kidney Int* 2013; 84:1079-89.
6. Alchi B, Jayne D. Membranoproliferative glomerulonephritis. *Pediatr Nephrol* 2010;25:1409-18.
7. Levy M, Gubler MC, Sich M, Beziou A, Habib R. Immunopathology of membranoproliferative glomerulonephritis with subendothelial deposits (Type I MPGN). *Clin Immunol Immunopathol* 1978;10:477-92.
8. Habib R, Gubler MC, Loirat C, Maiz HB, Levy M. Dense deposit disease: a variant of membranoproliferative glomerulonephritis. *Kidney Int* 1975;7:204-15.
9. Burkholder PM, Marchand A, Krueger RP. Mixed membranous and proliferative glomerulonephritis. A correlative light, immunofluorescence, and electron microscopic study. *Lab Invest* 1970;23:459-79.
10. Strife CF, McEnery PT, McAdams AJ, West CD. Membranoproliferative glomerulonephritis with disruption of the glomerular basement membrane. *Clin Nephrol* 1977;7:65-72.
11. Anders D, Agricola B, Sippel M, Thoenes W. Basement membrane changes in membranoproliferative glomerulonephritis. II. Characterization of a third type by silver impregnation of ultra thin sections. *Virchows Arch A Pathol Anat Histol* 1977;376:1-19.
12. Nasr SH, Valeri AM, Appel GB, Sherwinter J, Stokes MB, Said SM, et al. Dense deposit disease: clinicopathologic study of 32 pe-

- diatric and adult patients. *Clin J Am Soc Nephrol* 2009;4:22-32.
13. Walker PD, Ferrario F, Joh K, Bonsib SM. Dense deposit disease is not a membranoproliferative glomerulonephritis. *Mod Pathol* 2007;20:605-16.
 14. Belgiojoso GB, Tarantino A, Bazzi C, Colasanti G, Guerra L, Durante A. Immunofluorescence patterns in chronic membranoproliferative glomerulonephritis (MPGN). *Clin Nephrol* 1976;6:303-10.
 15. Pickering RJ, Herdman RC, Michael AF, Vernier RL, Gewurz H, Fish AJ, et al. Chronic glomerulonephritis associated with low serum complement activity (chronic hypocomplementemic glomerulonephritis). *Medicine (Baltimore)* 1970;49:207-26.
 16. Fakhouri F, Fremeaux-Bacchi V, Noel LH, Cook HT, Pickering MC. C3 glomerulopathy: a new classification. *Nat Rev Nephrol* 2010;6:494-9.
 17. Noris M, Remuzzi G. Overview of complement activation and regulation. *Semin Nephrol* 2013;33:479-92.
 18. Bomback AS, Markowitz GS, Appel GB. Complement-Mediated Glomerular Diseases: A Tale of 3 Pathways. *Kidney Int Rep* 2016;1:148-55.
 19. Nilsson B, Nilsson Ekdahl K. The tick-over theory revisited: Is C3 a contact-activated protein? *Immunobiology* 2012;217:1106-10.
 20. Wong EKS, Kavanagh D. Diseases of complement dysregulation—an overview. *Semin Immunopathol* 2018;40:49-64.
 21. Spitzer RE, Stitzel AE, Tsokos GC. Production of IgG and IgM autoantibody to the alternative pathway C3 convertase in normal individuals and patients with membranoproliferative glomerulonephritis. *Clin Immunol Immunopathol* 1990;57:10-8.
 22. Jozsi M, Reuter S, Nozal P, Lopez-Trascasa M, Sanchez-Corral P, Prohaszka Z, et al. Autoantibodies to complement components in C3 glomerulopathy and atypical hemolytic uremic syndrome. *Immunol Lett* 2014;160:163-71.
 23. Paixao-Cavalcante D, Lopez-Trascasa M, Skattum L, Giclas PC, Goodship TH, de Cordoba SR, et al. Sensitive and specific assays for C3 nephritic factors clarify mechanisms underlying complement dysregulation. *Kidney Int* 2012;82:1084-92.
 24. Ito S, Tamura N, Fujita T. Effect of decay-accelerating factor on the assembly of the classical and alternative pathway C3 convertases in the presence of C4 or C3 nephritic factor. *Immunology* 1989;68:449-52.
 25. Zhang Y, Nester CM, Holanda DG, Marsh HC, Hammond RA, Thomas LJ, et al. Soluble CR1 therapy improves complement regulation in C3 glomerulopathy. *J Am Soc Nephrol* 2013;24:1820-9.
 26. Corvillo F, Okroj M, Nozal P, Melgosa M, Sanchez-Corral P, Lopez-Trascasa M. Nephritic Factors: An Overview of Classification, Diagnostic Tools and Clinical Associations. *Front Immunol* 2019;10:886.
 27. Marinozzi MC, Chauvet S, Le Quintrec M, Mignotet M, Petitprez F, Legendre C, et al. C5 nephritic factors drive the biological phenotype of C3 glomerulopathies. *Kidney Int* 2017;92:1232-41.
 28. Zhang Y, Meyer NC, Fervenza FC, Lau W, Keenan A, Cara-Fuentes G, et al. C4 Nephritic Factors in C3 Glomerulopathy: A Case Series. *Am J Kidney Dis* 2017;70:834-43.
 29. Ravindran A, Fervenza FC, Smith RJH, De Vriese AS, Sethi S. C3 Glomerulopathy: Ten Years' Experience at Mayo Clinic. *Mayo Clin Proc* 2018;93:991-1008.
 30. Michels M, van de Kar N, van den Bos RM, van der Velden T, van Kraaij SAW, Sarlea SA, et al. Novel Assays to Distinguish Between Properdin-Dependent and Properdin-Independent C3 Nephritic Factors Provide Insight Into Properdin-Inhibiting Therapy. *Front Immunol* 2019;10:1350.
 31. Blanc C, Togarsimalemath SK, Chauvet S, Le Quintrec M, Moulin B, Buchler M, et al. Anti-factor H autoantibodies in C3 glomerulopathies and in atypical hemolytic uremic syndrome: one target, two diseases. *J Immunol* 2015;194:5129-38.
 32. Marinozzi MC, Roumenina LT, Chauvet S, Hertig A, Bertrand D, Olagne J, et al. Anti-Factor B and Anti-C3b Autoantibodies in C3 Glomerulopathy and Ig-Associated Membranoproliferative GN. *J Am Soc Nephrol* 2017;28:1603-13.
 33. Goicoechea de Jorge E, Caesar JJ, Malik TH, Patel M, Colledge M, Johnson S, et al. Dimerization of complement factor H-related proteins modulates complement activation in vivo. *Proc Natl Acad Sci U S A* 2013;110:4685-90.
 34. Martinez-Barricarte R, Heurich M, Valdes-Canedo F, Vazquez-Martul E, Torreira E, Montes T, et al. Human C3 mutation reveals a mechanism of dense deposit disease pathogenesis and provides insights into complement activation and regulation. *J Clin Invest* 2010;120:3702-12.
 35. Pickering M, Cook HT. Complement and glomerular disease: new insights. *Curr Opin Nephrol Hypertens* 2011;20:271-7.
 36. Ravindran A, Fervenza FC, Smith RJH, Sethi S. C3 glomerulopathy associated with monoclonal Ig is a distinct subtype. *Kidney Int* 2018;94:178-86.
 37. Sethi S, Rajkumar SV, D'Agati VD. The Complexity and Heterogeneity of Monoclonal Immunoglobulin-Associated Renal Diseases. *J Am Soc Nephrol* 2018;29:1810-23.
 38. Sethi S, Fervenza FC, Rajkumar SV. Spectrum of manifestations of monoclonal gammopathy-associated renal lesions. *Curr Opin Nephrol Hypertens* 2016;25:127-37.
 39. Hou J, Markowitz GS, Bomback AS, Appel GB, Herlitz LC, Barry Stokes M, et al. Toward a working definition of C3 glomerulopathy by immunofluorescence. *Kidney Int* 2014;85:450-6.
 40. Yagi K, Yanagida H, Sugimoto K, Kuwajima H, Tabata N, Morita K, et al. Clinicopathologic features, outcome, and therapeutic interventions in four children with isolated C3 mesangial proliferative glomerulonephritis. *Pediatr Nephrol* 2005;20:1273-8.
 41. Cook HT. C3 glomerulopathy. *F1000Res* 2017;6:248.
 42. Servais A, Noel LH, Roumenina LT, Le Quintrec M, Ngo S, Dragon-Durey MA, et al. Acquired and genetic complement abnormalities play a critical role in dense deposit disease and other C3 glomerulopathies. *Kidney Int* 2012;82:454-64.
 43. Viswanathan GK, Nada R, Kumar A, Ramachandran R, Rayat CS, Jha V, et al. Clinico-pathologic spectrum of C3 glomerulopathy-

- an Indian experience. *Diagn Pathol* 2015;10:6.
44. Ravindran A, Fervenza FC, Smith RJH, Sethi S. C3 glomerulonephritis with a severe crescentic phenotype. *Pediatr Nephrol* 2017;32:1625-33.
 45. Prema KSJ, Kurien AA, Gopalakrishnan N, Walker PD, Larsen CP. Dense deposit disease: a greatly increased biopsy incidence in India versus the USA. *Clin Kidney J* 2019;12:476-82.
 46. Park SJ, Kim YJ, Ha TS, Lim BJ, Jeong HJ, Park YH, et al. Dense deposit disease in Korean children: a multicenter clinicopathologic study. *J Korean Med Sci* 2012;27:1215-21.
 47. Sethi S, Fervenza FC, Smith RJ, Haas M. Overlap of ultrastructural findings in C3 glomerulonephritis and dense deposit disease. *Kidney Int* 2015;88:1449-50.
 48. Cook HT, Pickering MC. Histopathology of MPGN and C3 glomerulopathies. *Nat Rev Nephrol* 2015;11:14-22.
 49. Sethi S, Fervenza FC, Zhang Y, Nasr SH, Leung N, Vrana J, et al. Proliferative glomerulonephritis secondary to dysfunction of the alternative pathway of complement. *Clin J Am Soc Nephrol* 2011;6:1009-17.
 50. Bridoux F, Desport E, Fremeaux-Bacchi V, Chong CF, Gombert JM, Lacombe C, et al. Glomerulonephritis with isolated C3 deposits and monoclonal gammopathy: a fortuitous association? *Clin J Am Soc Nephrol* 2011;6:2165-74.
 51. Hawfield A, Iskandar SS, Smith RJ. Alternative pathway dysfunction in kidney disease: a case report and review of dense deposit disease and C3 glomerulopathy. *Am J Kidney Dis* 2013;61:828-31.
 52. Strife CF, West CD, Witte DP. Crescentic glomerulonephritis in childhood: acute nonproliferative glomerulitis versus dense deposit disease. *Am J Kidney Dis* 2003;41:897; author reply 8.
 53. Churg J, Duffy JL, Bernstein J. Identification of dense deposit disease: a report for the International Study of Kidney Diseases in Children. *Arch Pathol Lab Med* 1979;103:67-72.
 54. Medjeral-Thomas NR, O'Shaughnessy MM, O'Regan JA, Traynor C, Flanagan M, Wong L, et al. C3 glomerulopathy: clinicopathologic features and predictors of outcome. *Clin J Am Soc Nephrol* 2014;9:46-53.
 55. Sethi S, Nasr SH, De Vriese AS, Fervenza FC. C4d as a Diagnostic Tool in Proliferative GN. *J Am Soc Nephrol*. 2015;26:2852-9.
 56. Goodship TH, Cook HT, Fakhouri F, Fervenza FC, Fremeaux-Bacchi V, Kavanagh D, et al. Atypical hemolytic uremic syndrome and C3 glomerulopathy: conclusions from a "Kidney Disease: Improving Global Outcomes" (KDIGO) Controversies Conference. *Kidney Int* 2017;91:539-51.
 57. Sethi S, Fervenza FC, Zhang Y, Zand L, Meyer NC, Borsa N, et al. Atypical postinfectious glomerulonephritis is associated with abnormalities in the alternative pathway of complement. *Kidney Int* 2013;83:293-9.
 58. Kanjanabuch T, Kittikowit W, Eiam-Ong S. An update on acute postinfectious glomerulonephritis worldwide. *Nat Rev Nephrol* 2009;5:259-69.
 59. Fish AJ, Herdman RC, Michael AF, Pickering RJ, Good RA. Epidemic acute glomerulonephritis associated with type 49 streptococcal pyoderma. II. Correlative study of light, immunofluorescent and electron microscopic findings. *Am J Med* 1970;48:28-39.
 60. Sethi S, Fervenza FC, Zhang Y, Zand L, Vrana JA, Nasr SH, et al. C3 glomerulonephritis: clinicopathological findings, complement abnormalities, glomerular proteomic profile, treatment, and follow-up. *Kidney Int* 2012;82:465-73.
 61. Sandhu G, Bansal A, Ranade A, Jones J, Cortell S, Markowitz GS. C3 glomerulopathy masquerading as acute postinfectious glomerulonephritis. *Am J Kidney Dis* 2012;60:1039-43.
 62. Vernon KA, Goicoechea de Jorge E, Hall AE, Fremeaux-Bacchi V, Aitman TJ, Cook HT, et al. Acute presentation and persistent glomerulonephritis following streptococcal infection in a patient with heterozygous complement factor H-related protein 5 deficiency. *Am J Kidney Dis* 2012;60:121-5.
 63. Suga K, Kondo S, Matsuura S, Kinoshita Y, Kitano E, Hatanaka M, et al. A case of dense deposit disease associated with a group A streptococcal infection without the involvement of C3NeF or complement factor H deficiency. *Pediatr Nephrol* 2010;25:1547-50.
 64. Prasto J, Kaplan BS, Russo P, Chan E, Smith RJ, Meyers KE. Streptococcal infection as possible trigger for dense deposit disease (C3 glomerulopathy). *Eur J Pediatr* 2014;173:767-72.
 65. Sawanobori E, Umino A, Kanai H, Matsushita K, Iwasa S, Kitamura H, et al. A prolonged course of Group A streptococcus-associated nephritis: a mild case of dense deposit disease (DDD)? *Clin Nephrol* 2009;71:703-7.
 66. Okabe M, Tsuboi N, Yokoo T, Miyazaki Y, Utsunomiya Y, Hosoya T. A case of idiopathic membranoproliferative glomerulonephritis with a transient glomerular deposition of nephritis-associated plasmin receptor antigen. *Clin Exp Nephrol* 2012;16:337-41.