

Biology of invasive mucinous adenocarcinoma of the lung

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Abstract: Invasive mucinous adenocarcinoma (IMA) is a unique histologic subtype of lung adenocarcinoma. Recent studies document distinctive genetic alterations (e.g., *NRG1* fusions) and a “mucinous gene signature” in IMAs, as well as differences in clinical responses to traditional chemotherapies in IMAs versus non-mucinous adenocarcinomas. Our understanding of the genetic and clinical characteristics of IMAs has expanded, confirming the uniqueness of IMAs. Accordingly, IMAs require different therapeutic approaches than do lung adenocarcinomas in general. Here, we review recent updates on the genetic and clinical profiles of IMA of the lung.

Keywords: Lung cancer; adenocarcinoma; mucinous; biology; genetics

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Introduction

A primary pulmonary invasive mucinous adenocarcinoma (IMA) is an adenocarcinoma variant according to the current World Health Organization (WHO) classification of lung tumors (1). IMAs are less prevalent than are invasive non-mucinous adenocarcinomas (INMAs), accounting for approximately 5% of lung adenocarcinomas (2). IMAs have clinical, radiological, pathological, and genetic characteristics distinct from those of INMAs. In this article, we review the recently reported genetic and clinical characteristics as well as the general features of IMA of the lung.

Histopathology and immunohistochemical features of IMAs

By definition, IMAs consist of goblet and/or columnar tumor cells containing intracytoplasmic mucin (*Figure 1*), which are rarely observed in other lung adenocarcinomas (1,2). Because cytologic atypia is usually inconspicuous or absent in IMAs, definitive diagnosis of malignancy via biopsy is frequently challenging. In transbronchial biopsy,

accurate targeting could be limited since most IMAs are located in the periphery of lower lobes. Moreover, submucosal mucinous glands of bronchial tissue and the tumor cells of IMA could mimic each other because of bland-looking cytomorphology and intracytoplasmic mucin. Percutaneous computer tomography (CT)-guided lung biopsy appears to be more effective way to obtain the diagnostic tissue, however biopsied specimen could be composed of acellular mucin pool only since the alveolar spaces at the tumor periphery are often filled with mucin, which may correspond to the lobar pneumonia-like area on the chest CT image. Even if tumor cells are included in the specimen, a small amount of mucinous cells with bland morphology could be insufficient to make a definitive diagnosis of malignancy.

IMAs may show the same heterogeneous mixture of lepidic, acinar, papillary, micropapillary, and solid growth patterns as do INMAs (2). However, a lepidic growth pattern with microscopic skip lesions is a characteristic feature of IMAs, which is why they were formerly termed mucinous bronchioloalveolar carcinomas. Mixed invasive mucinous and non-mucinous adenocarcinomas, which are

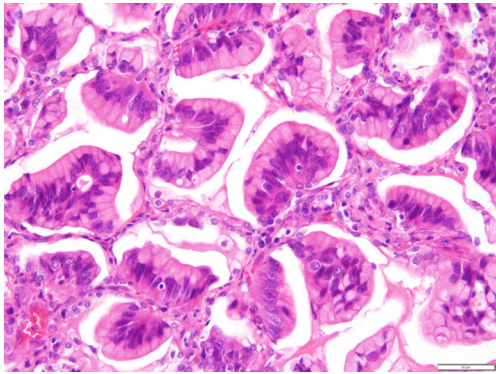


Figure 1 A representative photograph of an invasive mucinous adenocarcinoma (IMA) demonstrating goblet or columnar tumor cells with abundant intracytoplasmic mucin and basally located nuclei, characteristic of IMA. (Hematoxylin and eosin stain, magnification $\times 200$).

currently classified as a subcategory of IMAs by the WHO, have the histomorphologic features of both IMAs and INMAs (1). IMAs sometimes show depleted cytoplasmic mucin and aggravated cytologic atypia in areas of stromal invasion; thus, the non-mucinous components in the mixed tumors presumably arise from high-grade transformation of pre-existing mucinous components. However, further discussion and investigation are still required.

Colloid adenocarcinomas, in which abundant mucin pools replace air spaces, are also variants of lung adenocarcinomas (1). They differ from IMAs in two aspects: the mucin pools replace the underlying alveolar architecture, and scattered clusters of mucinous tumor cells are present (1). When diagnosing IMAs, the emphasis is made on intracellular mucin rather than extracellular mucin.

By immunohistochemistry, most IMAs express cytokeratin 7 (~88%) and cytokeratin 20 (~54%) (3-6). Expression rates of thyroid transcription factor-1 (TTF-1) and napsin A are variable in IMA but less than INMA, reported as approximately 40% and up to 33%, respectively (6,7). These findings support the premise that IMAs and INMAs have different cellular lineages. Previous studies categorized adenocarcinomas as terminal respiratory unit (TRU) type and non-TRU type, and IMAs are usually considered non-TRU type (8,9).

IMAs were recently shown to selectively express hepatocyte nuclear factor 4 α (HNF4 α) (10,11), a transcription factor present in normal and malignant gastrointestinal mucosa (12). Such selective expression could discriminate IMAs from other mucin-producing adenocarcinomas

such as those harboring *EGFR* or *ALK* mutations (11,13). A subset of adenocarcinomas with *ALK* rearrangements extensively produces extracellular mucin along with intracytoplasmic mucin, as do IMAs. However, because these adenocarcinomas differ from IMAs in terms of mutational status and consequently treatment strategies, accurate diagnosis is important and requires a comprehensive understanding of the genetic and histologic features including immunohistochemical profiles and cytomorphology (13).

Genetic alterations and expression profiles

In addition to their striking histologic features, IMAs also have a distinct molecular signature. Recent high throughput analyses revealed several genetic abnormalities in IMAs including *KRAS*, *BRAF*, *ERBB2*, and *PIK3CA* mutations and *NRG1*, *BRAF*, *NTRK1*, *ALK*, *RET*, and *ERBB4* rearrangements (14,15). *KRAS* mutations are the most frequent oncogenic driver mutations in IMAs (up to 86%) (14,15). However, IMAs and INMAs have different types of *KRAS* mutations: the most common types are G12D and G12V in IMAs and G12C in INMAs (15). G12D and G12V are most common in colorectal and pancreatobiliary carcinomas, suggesting that IMA of lung may be biologically more similar to pancreatobiliary and intestinal tract cancers. Although common in INMAs, *EGFR* mutations are very rare in IMAs (only 0–5%) (Figure 2) (14–17).

NRG1 fusions have been recently identified in IMAs, with an estimated frequency of 7–27% (Figure 2) (14,18,19). The fusion partners for *NRG1* in IMAs thus far include *CD74* (the most common), *SLC3A2*, and *VAMP2* (14,15,18,19). IMAs also harbor, albeit at low frequencies, *ERBB2* mutations, *BRAF* mutations, *BRAF* fusions, and *NTRK1* fusions (14,15). Drugs selectively targeting these kinases may be useful for treating IMAs, particularly given the success of kinase inhibitors in the treatment of lung cancers with *EGFR* mutations or *ALK* rearrangements.

Maeda *et al.* and Snyder *et al.* reported that *KRAS* lung cancer mouse models with reduced expression of *NKX2-1* (also known as *TTF-1*) developed mucinous lung tumors mimicking human IMA (20,21). Recently, Hwang *et al.* found that frameshift or nonsense mutations in *NKX2-1* occurred in approximately 19% (4/21) of IMAs as determined via next-generation sequencing and Sanger sequencing (16). Owing to these mutations, the TTF-1 protein was not expressed, which is a common finding in IMAs. INMAs, on the other hand, express wild-type *NKX2-1*,

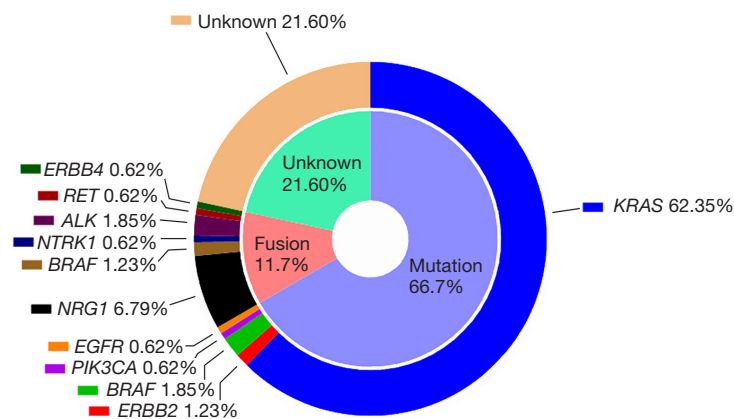


Figure 2 Pie chart showing the percentages of invasive mucinous adenocarcinomas (n=162) that harbor the indicated driver genes [combined data from references (14) and (15) are shown].

whose protein product (TTF-1) suppresses the expression of *FOXA3*, *SPDEF*, and *HNF4A* and, consequently, the expression of the mucin-related genes *MUC5AC*, *MUC5B*, and *MUC3* (22). In IMAs, loss of TTF-1 expression owing to *NKX2-1* mutation would de-repress the transcription of these downstream genes, thus accounting for the presence of *HNF4a* (11) and *MUC5AC* (9) in these tumors.

Primary IMAs are often indistinguishable from metastatic adenocarcinomas of the gastrointestinal or pancreatobiliary tract as both tumor types may have *KRAS* mutations and produce mucin (7). Differentiation is especially challenging if these tumors, in the lung or other primary sites, express CDX-2 and have the same type of *KRAS* mutation (7). Furthermore, recent RNA sequencing analysis of IMAs revealed a “mucinous signature” similar to that of gastrointestinal mucinous tumors (22). However, *NKX2-1* mutations appear to be specific for pulmonary IMAs (16) and if present, may help differentiating a lung primary from metastasis from the other sites.

Interestingly, IMAs rarely exhibit *TP53* mutations, which may reflect the lower mutation burden of IMAs compared with that of INMAs (15). IMAs rarely express PD-L1 and predominantly occur in people who are not heavy smokers (18,22). In comparison, *KRAS*-mutated INMAs have a higher rate of *T53* mutation and PD-L1 expression and occur mainly in heavy smokers (23-25). Hence, IMAs and INMAs differ genetically in terms of mutation burden even in the presence of *KRAS* mutations. Thus far, comprehensive gene expression profiling of IMAs has fallen short, perhaps because of their lower prevalence than INMA. Furthermore, previous expression data did not consider histologic subtypes in detail. However, a recent

study demonstrated that IMAs has distinct expression profiles through RNA sequencing. IMAs express *VTCN1/B7-H4* mRNA, whose protein product regulates an immune checkpoint and hence is a potential immunotherapy target, while they do not appreciably express PD-L1, as do INMAs (22). This finding may open new avenues of treatment using immune checkpoint inhibitors aimed at proteins other than PD-1/PD-L1.

Clinical features, prognostic significance, and response to treatments

IMAs tend to present with multicentric opacities or consolidation and multi-lobar and bilateral involvement, mimicking pneumonia (1,26). Interestingly, primary IMAs are frequently found in the lower lobes of the lungs (26).

Survival data for IMA are limited owing to its low incidence and tend to be contradictory (15,27,28). Yoshizawa *et al.* classified IMAs as high-grade based on a relatively high recurrence rate (76% in 5 years) (27), although the data may lack definitive prognostic significance given the small number of IMAs and the study population limited to stage I tumors in their study. The high recurrence rate could be explained by microscopic skip lesions, a characteristic feature of IMA, which is relevant to the concept of spread through airspace (STAS) of tumor cells (29). STAS is a recently described concept defined as tumor cells within air spaces in the lung parenchyma beyond the edge of the main tumor (29). Besides the conventional concept of ‘invasion’ in lung cancer—infiltration of stroma, blood vessels, or pleura—STAS is a newly recognized pattern of invasion in lung cancer, based on its anatomical characteristics that lung

is an air-filled organ with supporting alveolar structures. When STAS was present in the sub-anatomically resected lung, both locoregional and distant recurrence rates were significantly increased (29). As STAS contributes to patient prognosis, especially local recurrence, even in small early-stage adenocarcinomas (29,30), the skip patterns of IMA may do likewise. In IMAs, it is hypothesized that tumor cells travel in the background of abundant alveolar mucin and become situated in the alveolar walls away from the primary lesion. In our previous study, however, the recurrence-free survival rate did not differ significantly between IMAs and other tumors (15). Interestingly, all recurrences were limited to the lungs with no extrapulmonary metastases (15). Another study showed that even patients with IMA had longer recurrence-free survival than patients with INMA (28), and recurrences after surgical resection appear to be associated with the size of non-lepidic invasive pattern, and the presence of pleural and/or vascular invasion (31). These findings suggest that IMAs may not be aggressive tumors. In support, in a study of stage IV IMAs, overall survival (OS) was significantly better in untreated IMA patients than in untreated INMA patients (26).

Patients with lung adenocarcinomas harboring *EGFR* mutations or *ALK/ROS1* rearrangements qualify for targeted treatments with tyrosine kinase receptor inhibitors (TKIs). However, IMA patients are usually ineligible for such treatments, as well as for clinical trials of TKIs, owing to the rarity or absence of targetable mutations. Instead, most patients with advanced-stage IMAs receive non-TKI, platinum-based conventional chemotherapy (CTx), which, according to our previous study, does not improve OS relative to no treatment (26). Non-TKI CTx did, however, improve OS in INMA patients, although to a lesser extent than did TKI-based therapy in INMA patients with targetable mutation (26). These results indicate that IMA patients do not appreciably benefit from non-TKI CTx, and hence, new therapeutic approaches are required for IMAs of the lung.

Conclusions

Our rapidly evolving knowledge of the genetic and clinical characteristics of IMAs confirms the uniqueness of this disease among other primary lung adenocarcinomas. New findings regarding the targetable genetic alterations and clinical profiles of IMAs are anticipated to result in better patient management. Via comprehensive clinicopathologic and molecular analyses, IMAs will be more accurately

diagnosed and more thoroughly investigated.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

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