

Long-Term Follow-Up of Patients After Autologous Bone Marrow Cell Infusion for Decompensated Liver Cirrhosis

Ja Kyung Kim,* Soo-Jeong Kim,* Yuri Kim,* Yong Eun Chung,† Young Nyun Park,‡
Hyun Ok Kim,§ Jin Seok Kim,* Mi-Suk Park,† Isao Sakaida,¶ Do Young Kim,*
Jung Il Lee,* Sang Hoon Ahn,* Kwan Sik Lee,* and Kwang-Hyub Han*

*Department of Internal Medicine, Yonsei University College of Medicine, Seoul, South Korea

†Department of Radiology, Yonsei University College of Medicine, Seoul, South Korea

‡Department of Pathology, Yonsei University College of Medicine, Seoul, South Korea

§Department of Laboratory Medicine, Yonsei University College of Medicine, Seoul, South Korea

¶Department of Gastroenterology and Hepatology, Yamaguchi University Graduate School of Medicine, Ube, Yamaguchi, Japan

Although several human clinical trials using various bone marrow-derived cell types for cirrhotic or decompensated patients have reported a short-term benefit, long-term follow-up data are limited. We analyzed the long-term clinical outcomes of autologous bone marrow cell infusion (ABMI) for decompensated liver cirrhosis (LC). Patients enrolled in a pilot single-armed ABMI study were followed up more than 5 years. Bone marrow-derived mononuclear cells (BM-MNCs) from decompensated LC were harvested and after processing were infused into a peripheral vein. The laboratory test results and long-term clinical course including liver transplantation (LT), development of cancer, cause of death, and survival after ABMI were analyzed. Nineteen patients were followed up for a median of 66 months after ABMI. Liver function, including serum levels of albumin and Child–Pugh (CP) score, was improved at the 1-year follow-up. Liver volume was significantly greater, cirrhosis was sustained, and collagen content was decreased at the 6-month follow-up. Five years after ABMI, five patients (26.3%) maintained CP class A without LT or death, and five patients (26.3%) had undergone elective LT. Hepatocellular carcinoma (HCC) occurred in five patients (26.3%), and lymphoma and colon cancer occurred in one patient each. Three patients (15.8%) were lost to follow-up at months 22, 31, and 33, respectively, but maintained CP class A until their last follow-up. Five patients expired due to infection. While improved liver function was maintained in some patients for more than 5 years after ABMI, other patients developed HCC. Further studies of long-term follow-up cohorts after cell therapy for LC are warranted.

Key words: Adult stem cells; Bone marrow; Autologous cells; Transplantation; Liver regeneration; Liver cirrhosis (LC)

INTRODUCTION

The end point of liver disease, irrespective of its cause, is liver cirrhosis (LC). Because fibrous scars hardly disappear, a cirrhotic liver does not return to a normal state. Before decompensation, control of the underlying liver disease is important to recover to some degree. After decompensation, patients suffer from repeated and continuous complications, including hepatocellular carcinoma (HCC). At present, liver transplantation (LT) is the only treatment option for patients with HCC.

Although antifibrotic drugs have been developed, their efficacy is limited. Research into bone marrow cells

(BMCs) has facilitated the application of cellular therapeutics. In hepatology, hepatocytes from BM have been found in mouse^{1,2} and human^{3,4} livers after BMC transplantation, suggesting that BMCs could contribute to liver regeneration. The fate of BMCs injected into fibrotic or injured livers has been investigated^{5,6}. Injected cells migrate to the fibrous area, penetrate into cords of hepatocytes, and produce enzymes that ameliorate fibrosis⁶. Together with BMCs, mesenchymal stem cells (MSCs) have been investigated. Following enhancement of their immunomodulatory and antifibrotic effects in other organ systems, application of MSCs to treat liver fibrosis has

shown positive results⁷⁻¹⁰. For clinical application, MSCs must undergo *in vitro* selection and culture. A clinical trial using MSCs, therefore, could not start at the initiatory stage in cell therapy until assurance of safety during *in vitro* manipulation. Although early studies had suggested transdifferentiation of BMCs or MSCs into hepatocytes, mechanisms of action remain poorly understood. Paracrine effects supporting hepatocyte function and liver regeneration and immune modulation are thought to be possible mechanisms¹¹.

For clinical application, cell therapy similar to autologous BM stem cell transplantation was first evaluated¹², as this procedure has long been applied to the treatment of hematologic disease. Subsequent clinical trials of autologous BMCs selected by means of surface markers¹³ or injected through the hepatic artery¹⁴ or the portal vein¹⁵ have reported efficacy in terms of liver regeneration after liver surgery or transient improvement of hepatic function. Further trials using BMCs¹⁶⁻¹⁸, mobilized peripheral blood mononuclear cells (PBMCs)^{19,20}, BM-derived endothelial progenitor cells²¹, or BM-derived MSCs (BM-MSCs)²²⁻²⁴ for LC have reported positive short-term results, but the beneficial effects lasted for less than 2 years²³. However, no study has evaluated maintenance of liver function, survival, and HCC development in such patients at ≥ 3 years after treatment.

We previously reported the preliminary outcomes of patients who underwent autologous bone marrow cell infusion (ABMI) for advanced LC²⁵. Together with gradual improvement of liver function and volume, serial liver biopsies showed an activated progenitor cell compartment after ABMI. A planned single-armed pilot study has been completed, and enrolled patients were followed up for more than 5 years. In this study, the long-term clinical outcomes of patients who underwent ABMI therapy for decompensated LC were analyzed.

MATERIALS AND METHODS

Patients

Patients enrolled in a pilot trial of ABMI therapy for decompensated LC between June 2006 and February 2011 were analyzed. All patients provided informed consent and met the following inclusion criteria: age 18–75 years, biopsy-proven LC with Child–Pugh (CP) class B or C, platelet count $\geq 50,000/\mu\text{l}$ and total bilirubin ≤ 3.0 mg/dl, and without evidence of HCC on magnetic resonance imaging (MRI). Patients who had a history of HCC without recurrence 2 years after curative treatment were eligible. Patients with a medical condition that made them ineligible for general anesthesia during BM collection were excluded. Patients maintained all medications, with the exception of intravenous albumin. The diuretic dose was adjusted according to the patient's volume status after ABMI. Biochemical tests were performed

for 12 months after ABMI. Thereafter, patients were followed up according to their individual clinic visit schedule. Long-term follow-up data were collected retrospectively. In patients who consented to follow-up, LT and development of malignancy, including hepatic function reserve, were monitored. To avoid potential bias, ABMI was considered to have failed in patients who underwent LT.

This study conformed to the ethical guidelines of the Declaration of Helsinki and was approved by the institutional review board (IRB) of Severance Hospital in the Yonsei University Health System (No. 4-2006-0087). This study was conducted with the cooperation of Yamaguchi University Graduate School of Medicine in Japan, under a memorandum of understanding.

Cell Preparation

Under general anesthesia, 500–750 ml of BM was aspirated from both ilia and collected in a plastic bag containing heparin for harvesting of marrow for BMC transplantation. Collected BM was filtered to remove fat and bony particles using a collection and gravity-flow filtration system (BM collection kit; Fenwal Inc., Lake Zurich, IL, USA). Red blood cells and plasma were depleted, and mononuclear cells were collected in a closed system using a COBE 2991 Cell Processor (Gambro, Lakewood, CO, USA). The final volume was ~ 100 ml. Processed cells were enumerated and tested for viability and contamination. Fluorescence-activated cell sorting (FACS) analysis was performed using a flow cytometer (Cytomics FC 500; Beckman Coulter, Inc., Brea, CA, USA) to evaluate cell populations using cell surface markers [cluster of differentiation (CD) 34, CD45, and CD133 (Miltenyi Biotec Inc., Auburn, CA, USA); CD117 (Immunotech, Beckman Coulter, Marseille, France)]. The final cell product was infused into a peripheral vein over a 1-h period.

Clinical Evaluation

After obtaining informed consent, screening tests were performed to meet the inclusion criteria. Twelve months of clinical follow-up with serial evaluations was conducted. Follow-up visits were scheduled weekly for 2 weeks, monthly for the first 6 months, and then every 3 months for the following 6 months. At each visit, laboratory parameters (complete blood count, serum levels of blood glucose, total protein, albumin, bilirubin, aminotransferases, alkaline phosphatase, γ -glutamyl transpeptidase, urea, creatinine, cholesterol, α -fetoprotein levels, and prothrombin time) were evaluated. Indocyanine green clearance testing was performed before and 6 and 12 months after ABMI.

Liver MRI and biopsies were conducted before and 1, 3, and 6 months after ABMI. All MRI scans to

screen for HCC and quantify ascites and liver volume were performed using a 1.5-T MRI scanner (Gyrosan Intera; Philips Medical Systems, Best, The Netherlands). Routine T1-weighted gradient echo, T2-weighted turbo spin echo, T2*-weighted gradient echo, and three-phase, dynamic three-dimensional (3D) T1-weighted images after injection of gadobenate dimeglumine (dose, 0.1 mmol/kg body weight; MultiHance; Bracco SpA, Milan, Italy) were obtained. Total liver volume was measured using commercially available image postprocessing software (Voxel plus 2; Mevisys, Daeduk, South Korea) with one radiologist applying the summation of areas technique using a 10-mm reconstruction thickness. The radiologist was blinded to the patients' clinical information.

Liver biopsies were performed using an ultrasound-guided 16-gauge gun. Transjugular liver biopsy was attempted as an alternative method in cases with massive ascites or a high risk of bleeding. The biopsied liver tissues were routinely processed and stained with hematoxylin and eosin (H&E) and Masson's trichrome. Staining was performed by an autostainer (Symphony staining instrument; Ventana Medical Systems, Tucson, AZ, USA). Unstained slides, available after routine histologic evaluation, were analyzed using a computer-assisted, staining-free method for liver fibrosis assessment by Genesis[®] 200 (Histoindex, Singapore)²⁶.

Statistical Analysis

Serial data from baseline to follow-up visits after ABMI were analyzed. After normality test of data, continuous variables are presented as means \pm standard deviations or median (range) accordingly, and categorical variables are presented as absolute values and percentages. Paired-sample *t*-tests were used to compare values. Data of liver volume did not follow normal distribution; change of liver volume was compared with Wilcoxon's matched-pairs signed-ranks test. All statistical procedures were conducted using PASW Statistics 17.0 (IBM, Armonk, NY, USA). Analyses were based on two-tailed hypothesis tests, and a value of $p < 0.05$ was considered to indicate statistical significance.

RESULTS

Characteristics of Patients and Cells Infused for ABMI

Twenty patients underwent screening tests, one of whom was excluded according to the exclusion criteria. Therefore, 19 patients underwent ABMI and were followed up. The baseline characteristics of the patients are shown in Table 1. The mean age of the 9 male and 10 female patients was 52 years. Eighteen patients (94.7%) had B-viral LC, and one had alcoholic LC. Sixteen patients had stable hepatitis B with (14 patients) or without (2 patients) antiviral medications. Two patients started antiviral medications, and four patients experienced viral

Table 1. Baseline Characteristics of Patients ($N=19$)

Variable	Values
Age (years)	52 \pm 8
Sex (male/female)	9:10
Etiology (HBV/alcohol)	18:1
Ascites	14 (73.7)
AST (IU/L)	46 (28–196)
ALT (IU/L)	39.5 (21–130)
Albumin (g/dl)	2.9 \pm 0.4
Total bilirubin (mg/dl)	1.6 \pm 0.6
Cholesterol (mg/dl)	123.2 \pm 27.1
Hemoglobin (g/dl)	11.6 \pm 1.5
Prothrombin time (%)	70.9 \pm 11.2
Child–Pugh score	7.6 \pm 1.2
Child–Pugh class (B/C)	18:1
ICG R15 (%)	48.0 \pm 13.8
Liver stiffness (kPa)*	26.5 \pm 11.5

Variables are expressed as mean \pm standard deviation, median (range), or *n* (%). HBV, hepatitis B virus; AST, aspartate aminotransferase; ALT, alanine aminotransferase; ICG, indocyanine green clearance test; kPa, kilopascal.

*Liver stiffness was measured only in the patients without ascites ($n=5$).

breakthrough during the follow-up period; the antiviral medications of these patients were changed. Three patients had HCC prior to undergoing ABMI. Two patients underwent resection of HCC, and one patient received transarterial chemoembolization and sustained a complete response for >2 years. Fourteen patients (73.7%) had ascites before ABMI. The average CP score of the patients was 7.6, and all but one had cirrhosis of CP class B.

The characteristics of infused cells are shown in Table 2. The mean cell number per unit body weight was 0.925×10^8 cells/kg. The average final cell volume after red blood cell (RBC) depletion and concentration was 124.1 ml. These cells were infused over a 1-h period, and no adverse reactions were noted. The majority of the cell population was CD45⁺ hematopoietic stem cells (83.43%), while CD133⁺, CD117⁺, and CD34⁺ hematopoietic stem cells each made up $\sim 1\%$ of the population. There was no association between cell population and hepatic function improvement.

Table 2. Composition of Infused Bone Marrow Cells

Variables	Values
Total infused nucleated cells/weight ($\times 10^8$ cells/kg)	0.9250 \pm 0.3244
CD34 ⁺ cells (%)	0.64 \pm 0.44
CD45 ⁺ cells (%)	83.43 \pm 15.91
CD117 ⁺ cells (%)	0.85 \pm 0.59
CD133 ⁺ cells (%)	1.10 \pm 1.00

Variables are expressed as mean \pm standard deviation. CD, cluster of differentiation.

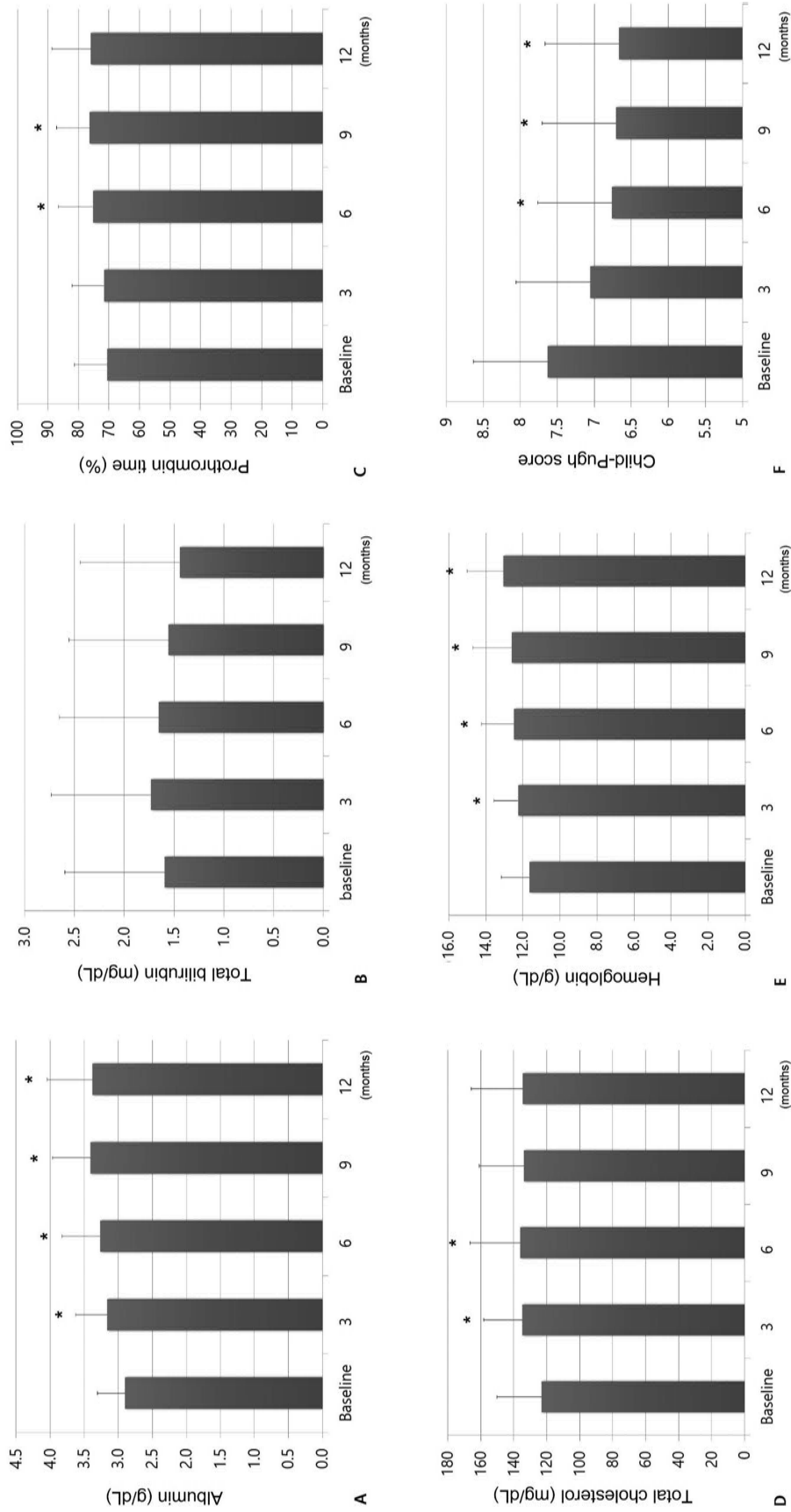


Figure 1. Results of serial blood tests after autologous bone marrow cell infusion (ABMI). (A) Serum levels of albumin gradually and significantly increased at 3, 6, 9, and 12 months after ABMI. (B) Total bilirubin levels did not change significantly. (C) Prothrombin time was significantly better at 6 and 9 months. (D) Total cholesterol level was significantly higher at 3 and 6 months. (E) Hemoglobin level was significantly higher after 3, 6, 9, and 12 months. (F) The Child–Pugh score was significantly better after 6, 9, and 12 months. * $p < 0.05$.

Changes in Liver Function

The results of serial blood tests after ABMI were compared to those before ABMI. The results of selected tests related to liver function are shown in Figure 1. The mean serum level of albumin before ABMI was 2.9 g/dl. The mean albumin level during the 6 months before ABMI was also 2.9 g/dl, and this was not improved with conservative management, including use of antiviral agents. This gradually and significantly increased to 3.2, 3.3, 3.4, and 3.4 g/dl at 3, 6, 9, and 12 months, respectively, after ABMI (Fig. 1A). The mean initial level of total bilirubin was 1.6 mg/dl, and it did not change significantly after ABMI (Fig. 1B). The average prothrombin time before ABMI was 70% and improved significantly to 75% at 6 months and 76% at 9 months after ABMI (Fig. 1C). The mean total cholesterol level increased from 123 to 134 mg/dl at 3 months and to 136 mg/dl at 6 months after ABMI (Fig. 1D). Hemoglobin levels increased significantly from 11.6 g/dl before ABMI to 12.3, 12.5, 12.5, and 13.1 g/dl at 3, 6, 9, and 12 months, respectively, after ABMI (Fig. 1E). The initial average CP score was 7.6 and improved significantly after 6, 9, and 12 months to 6.8, 6.7, and 6.7, respectively (Fig. 1F).

Twelve patients (63.2%) had achieved CP class A within 1 year. This improvement was significantly related only to an increased liver volume after 6 months ($p < 0.13$). At 24, 36, and 60 months, 11, 7, and 5 patients, respectively, had achieved CP class A without LT or death. Among the five patients with CP class A at 60 months, two had improved to that class after 24 months, while the others were of that class the full 60 months.

Changes in Liver Volume and Fibrosis

All patients underwent MRI at screening, but two patients refused to undergo MRI at the 6-month follow-up. The reconstructed liver volume of the other 17 patients increased significantly at 6 months after ABMI ($p = 0.049$) (Fig. 2A). Eleven patients (64.7%) showed an increased

liver volume compared to baseline. The greatest increase compared to baseline (Fig. 2B, left) was 42% after 6 months (Fig. 2B, right).

According to pathologic evaluation of serial biopsies, ABMI did not lead to resolution of cirrhosis. To compare changes in liver fibrosis, available unstained liver tissues were analyzed in terms of the level of collagen deposition. The percentage of portal collagen decreased significantly at 3 months (Fig. 3A). The septal collagen percentage showed a decreasing tendency after ABMI (Fig. 3B), as did the total fibrillar tissue collagen (fibrosis other than portal and septal fibrosis) percentage (Fig. 3C).

Long-Term Clinical Outcomes

Patients were followed up for a median of 66 months after ABMI (range: 2–113 months). Five patients (26.3%) underwent LT at 5–39 months, five patients (26.3%) died, and three patients (15.3%) were lost to follow-up. A survival curve of patients who did not undergo LT or die is shown in Figure 4A. The median duration of survival without LT was 48 months. Five patients died due to pneumonia ($n = 3$), lymphoma ($n = 1$), and spontaneous bacterial peritonitis ($n = 1$). Three patients (15.8%) were lost to follow-up at 22, 31, and 32 months; these patients did not exhibit deterioration of liver function at their final follow-up.

HCC developed in five patients (26.3%) after ABMI. One patient had previously undergone HCC resection, while the others developed HCC at 16, 17, 18, and 56 months after ABMI. Four patients underwent successful nonsurgical treatment, and one received elective LT. In HCC-naïve patients, the cumulative incidence of HCC at 3 and 5 years was 23.1% and 35.9%, respectively (Fig. 4B). The patient with HCC recurrence exhibited improved liver function but was diagnosed with lymphoma at 49 months. The lymphoma was thought to be hepatitis B virus (HBV)-related lymphoproliferative disease and led to death of the patient.

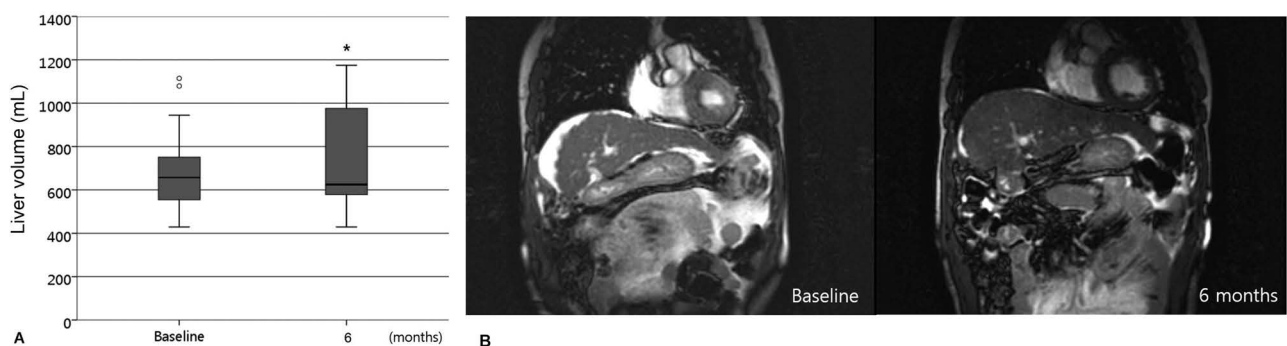


Figure 2. Changes in liver volume calculated using magnetic resonance imaging (MRI) scans at 6 months after ABMI. (A) Liver volume significantly increased after ABMI ($p = 0.049$). (B) Image of the most enlarged liver section image showed a 42% increase in liver volume and decreased ascites at 6 months after ABMI (right) compared to baseline (left). $*p < 0.05$.

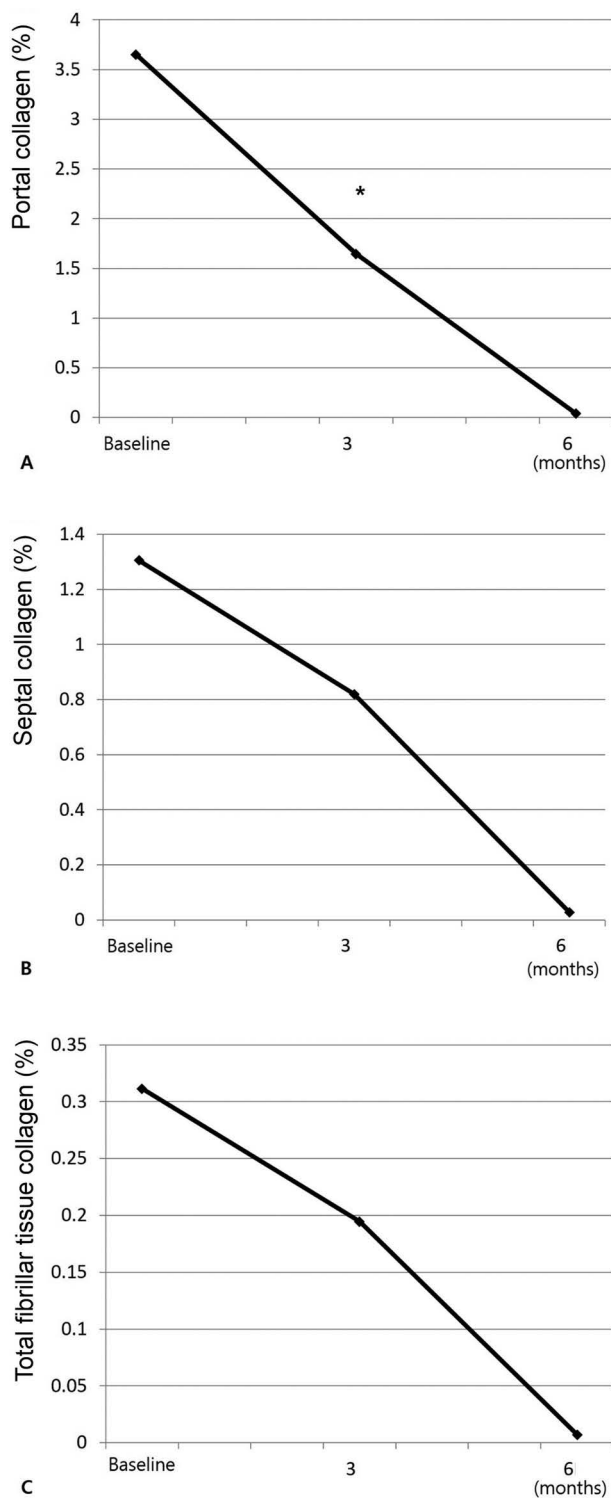


Figure 3. Collagen deposition as determined by a computer-assisted, staining-free method for assessment of liver fibrosis. (A) Percentage portal collagen was significantly lower at 3 months. (B) Septal collagen showed a decreasing tendency over time. (C) Total fibrillar tissue collagen (fibrosis other than portal and septal fibrosis) percentage also showed a decreasing tendency over time. * $p < 0.05$.

In one patient who showed improvement to CP class A, cancer of the sigmoid colon was found during routine colonoscopy at 18 months after ABMI. This was removed by endoscopic mucosal resection, and no further recurrence or hepatic function deterioration was noted.

DISCUSSION

The findings of this work suggest that ABMI for decompensated LC could improve liver function and decrease the collagen level in liver tissue transiently and inconspicuously, although LC itself was not resolved. Some patients successfully bridged to LT, and others sustained an improved state without significant adverse events related to ABMI. However, some patients developed infection or HCC after >5 years.

We evaluated the effects of ABMI using various parameters, while other studies using BMCs or MSCs for LC have focused on blood test results. Functional changes were assessed by determining serum levels of albumin and CP class. Pathologic and computer-assisted evaluation of fibrosis and liver tissue were performed. Regenerative changes were assessed by measuring liver volume. The patients showed improvement in terms of biochemical parameters, CP class, and liver regeneration, as indicated by an increased liver volume.

A recent meta-analysis of autologous BM transplantation in decompensated livers showed a benefit after 1 year²⁷. Previous studies of cell therapy have reported only short-term effects because long-term monitoring of subjects was not possible. In the early stage of cell therapy research, long-term safety was a matter of concern because, in contrast to chemotherapeutic agents, injected stem cells have long half-lives, the possible ability to transdifferentiate and/or repopulate, and no modulating manner after injected. While use of autologous cells prevents adverse immunologic reactions, their effects on cancer development are unclear. Therefore, maintenance of the therapeutic effects of cell therapy and its influence on cancer development are unresolved issues.

In this cohort, three patients with HCC underwent successful treatment before ABMI. These patients were included in this study because they had no recurrence of HCC for 2 years before ABMI and showed no HCC on screening MRI. Among these three patients, one experienced HCC recurrence at 10 months after ABMI. This patient underwent successful HCC treatment; however, lymphoma developed 49 months after ABMI. A hematologist speculated that the lymphoma was likely related to HBV infection, as reported previously²⁸. However, it is also possible that ABMI contributed to the development of lymphoma. Because lymphoproliferative disease usually progresses rapidly, the 4 years between ABMI and the lymphoma diagnosis means that a causal relationship is possible but unlikely. Further research is warranted to

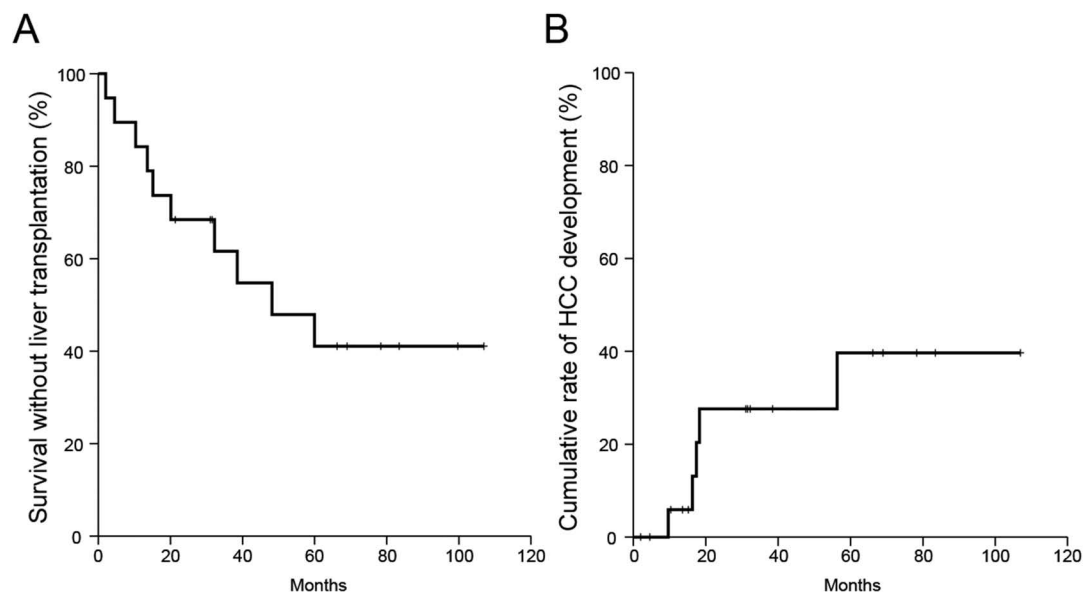


Figure 4. Survival curve without liver transplantation (LT) and cumulative rate of hepatocellular carcinoma (HCC) development. (A) The median survival duration without LT was 48 months. (B) In HCC-naïve patients, the cumulative rate of HCC development at 3 and 5 years was 23.1% and 35.9%, respectively.

resolve this issue. HCC did not recur in the other two patients with HCC before ABMI.

Other than the patient discussed above, HCC was newly diagnosed in four patients after ABMI. Because they had undergone regular evaluations, HCC was found at an early stage and treated successfully. Only one patient considered LT for treatment of HCC; the others elected nonsurgical treatments. The 5-year cumulative risk of HCC has been reported to be 15% in compensated HBV-related LC in a high-endemic area²⁹, and patients with a more advanced CP class had a higher risk of HCC³⁰. The rate of newly developed HCC in these patients (4/16, 25%) is comparable with that reported previously. However, the relatively high incidence of HCC within 2 years after ABMI warrants further study.

Infection was the most frequent cause of death in our patients. Two patients with improved liver function expired from uncontrolled pneumonia, and two patients without improvement died due to pneumonia and spontaneous bacterial peritonitis. To the best of our knowledge, no previous study has evaluated the frequency of infections after cell therapy in LC patients.

The limitations of this study were its single-arm design and inclusion of a small population. However, this study was started as an exploratory pilot trial in its early days of cell therapy. From the result of this trial, a randomized controlled trial is underway and is expected to provide more confirmative evidence in terms of therapeutic effects and influence on cancer development.

Our results suggest that ABMI for decompensated LC could improve liver function and volume over the short

term. Over the long term, while some patients maintained improved liver function, HCC developed in other patients. Further studies of long-term follow-up cohorts after cell therapy for LC are warranted.

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REFERENCES

- Petersen BE, Bowen WC, Patrene KD, Mars WM, Sullivan AK, Murase N, Boggs SS, Greenberger JS, Goff JP. Bone marrow as a potential source of hepatic oval cells. *Science* 1999;284(5417):1168–70.
- Theise ND, Badve S, Saxena R, Henegariu O, Sell S, Crawford JM, Krause DS. Derivation of hepatocytes from bone marrow cells in mice after radiation-induced myeloablation. *Hepatology* 2000;31(1):235–40.
- Alison MR, Poulson R, Jeffery R, Dhillon AP, Quaglia A, Jacob J, Novelli M, Prentice G, Williamson J, Wright NA. Hepatocytes from non-hepatic adult stem cells. *Nature* 2000;406(6793):257.
- Theise ND, Nimmakayalu M, Gardner R, Illei PB, Morgan G, Teperman L, Henegariu O, Krause DS. Liver from bone marrow in humans. *Hepatology* 2000;32(1):11–6.
- Terai S, Sakaida I, Yamamoto N, Omori K, Watanabe T, Ohata S, Katada T, Miyamoto K, Shinoda K, Nishina H, Okita K. An in vivo model for monitoring trans-differentiation of bone marrow cells into functional hepatocytes. *J Biochem*. 2003; 134(4):551–8.
- Sakaida I, Terai S, Yamamoto N, Aoyama K, Ishikawa T, Nishina H, Okita K. Transplantation of bone marrow cells reduces CCl4-induced liver fibrosis in mice. *Hepatology* 2004;40(6):1304–11.

7. Hardjo M, Miyazaki M, Sakaguchi M, Masaka T, Ibrahim S, Kataoka K, Huh NH. Suppression of carbon tetrachloride-induced liver fibrosis by transplantation of a clonal mesenchymal stem cell line derived from rat bone marrow. *Cell Transplant*. 2009;18(1):89–99.
8. Rabani V, Shahsavani M, Gharavi M, Piryaei A, Azhdari Z, Baharvand H. Mesenchymal stem cell infusion therapy in a carbon tetrachloride-induced liver fibrosis model affects matrix metalloproteinase expression. *Cell Biol Int*. 2010;34(6):601–5.
9. Zhao DC, Lei JX, Chen R, Yu WH, Zhang XM, Li SN, Xiang P. Bone marrow-derived mesenchymal stem cells protect against experimental liver fibrosis in rats. *World J Gastroenterol*. 2005;11(22):3431–40.
10. Jang YO, Jun BG, Baik SK, Kim MY, Kwon SO. Inhibition of hepatic stellate cells by bone marrow-derived mesenchymal stem cells in hepatic fibrosis. *Clin Mol Hepatol*. 2015;21(2):141–9.
11. Behbahan IS, Keating A, Gale RP. Concise review: Bone marrow autotransplants for liver disease? *Stem Cells* 2013; 31(11):2313–29.
12. Terai S, Ishikawa T, Omori K, Aoyama K, Marumoto Y, Urata Y, Yokoyama Y, Uchida K, Yamasaki T, Fujii Y, Okita K, Sakaida I. Improved liver function in patients with liver cirrhosis after autologous bone marrow cell infusion therapy. *Stem Cells* 2006;24(10):2292–8.
13. am Esch JS 2nd, Knoefel WT, Klein M, Ghodsizad A, Fuerst G, Poll LW, Piechaczek C, Burchardt ER, Feifel N, Stoldt V, Stockschröder M, Stoecklein N, Tustas RY, Eisenberger CF, Peiper M, Häussinger D, Hosch SB. Portal application of autologous CD133+ bone marrow cells to the liver: A novel concept to support hepatic regeneration. *Stem Cells* 2005;23(4):463–70.
14. Lyra AC, Soares MB, da Silva LF, Fortes MF, Silva AG, Mota AC, Oliveira SA, Braga EL, de Carvalho WA, Genser B, dos Santos RR, Lyra LG. Feasibility and safety of autologous bone marrow mononuclear cell transplantation in patients with advanced chronic liver disease. *World J Gastroenterol*. 2007;13(7):1067–73.
15. Mohamadnejad M, Vosough M, Moossavi S, Nikfam S, Mardpour S, Akhlaghpour S, Ashrafi M, Azimian V, Jarughi N, Hosseini SE, Moeininia F, Bagheri M, Sharafkhan M, Aghdami N, Malekzadeh R, Baharvand H. Intraportal infusion of bone marrow mononuclear or CD133+ cells in patients with decompensated cirrhosis: A double-blind randomized controlled trial. *Stem Cells Transl Med*. 2016;5(1):87–94.
16. Lyra AC, Soares MB, da Silva LF, Braga EL, Oliveira SA, Fortes MF, Silva AG, Brustolim D, Genser B, Dos Santos RR, Lyra LG. Infusion of autologous bone marrow mononuclear cells through hepatic artery results in a short-term improvement of liver function in patients with chronic liver disease: A pilot randomized controlled study. *Eur J Gastroenterol Hepatol*. 2010;22(1):33–42.
17. Saito T, Okumoto K, Haga H, Nishise Y, Ishii R, Sato C, Watanabe H, Okada A, Ikeda M, Togashi H, Ishikawa T, Terai S, Sakaida I, Kawata S. Potential therapeutic application of intravenous autologous bone marrow infusion in patients with alcoholic liver cirrhosis. *Stem Cells Dev*. 2011;20(9):1503–10.
18. Bai YQ, Yang YX, Yang YG, Ding SZ, Jin FL, Cao MB, Zhang YR, Zhang BY. Outcomes of autologous bone marrow mononuclear cell transplantation in decompensated liver cirrhosis. *World J Gastroenterol*. 2014;20(26):8660–6.
19. Han Y, Yan L, Han G, Zhou X, Hong L, Yin Z, Zhang X, Wang S, Wang J, Sun A, Liu Z, Xie H, Wu K, Ding J, Fan D. Controlled trials in hepatitis B virus-related decompensate liver cirrhosis: Peripheral blood monocyte transplant versus granulocyte-colony-stimulating factor mobilization therapy. *Cytotherapy* 2008;10(4):390–6.
20. Pai M, Zacharoulis D, Milicevic MN, Helmy S, Jiao LR, Levicar N, Tait P, Scott M, Marley SB, Jestice K, Glibetic M, Bansal D, Khan SA, Kyriakou D, Rountas C, Thillainayagam A, Nicholls JP, Jensen S, Apperley JF, Gordon MY, Habib NA. Autologous infusion of expanded mobilized adult bone marrow-derived CD34+ cells into patients with alcoholic liver cirrhosis. *Am J Gastroenterol*. 2008;103(8):1952–8.
21. D'Avola D, Fernandez-Ruiz V, Carmona-Torre F, Mendez M, Perez-Calvo J, Prosper F, Andreu E, Herrero JJ, Inarrairaegui M, Fuertes C, Bilbao JJ, Sangro B, Prieto J, Quiroga J. Phase 1-2 pilot clinical trial in patients with decompensated liver cirrhosis treated with bone marrow-derived endothelial progenitor cells. *Transl Res*. 2016 [Epub ahead of print].
22. Jang YO, Kim YJ, Baik SK, Kim MY, Eom YW, Cho MY, Park HJ, Park SY, Kim BR, Kim JW, Soo Kim H, Kwon SO, Choi EH, Kim YM. Histological improvement following administration of autologous bone marrow-derived mesenchymal stem cells for alcoholic cirrhosis: A pilot study. *Liver Int*. 2014;34(1):33–41.
23. Peng L, Xie DY, Lin BL, Liu J, Zhu HP, Xie C, Zheng YB, Gao ZL. Autologous bone marrow mesenchymal stem cell transplantation in liver failure patients caused by hepatitis B: Short-term and long-term outcomes. *Hepatology* 2011; 54(3):820–8.
24. Vosough M, Moossavi S, Mardpour S, Akhlaghpour S, Azimian V, Jarughi N, Hosseini SE, Ashrafi M, Nikfam S, Aghdami N, Malekzadeh R, Mohamadnejad M, Baharvand H. Repeated intraportal injection of mesenchymal stem cells in combination with pioglitazone in patients with compensated cirrhosis: A clinical report of two cases. *Arch Iran Med*. 2016;19(2):131–6.
25. Kim JK, Park YN, Kim JS, Park MS, Paik YH, Seok JY, Chung YE, Kim HO, Kim KS, Ahn SH, Kim DY, Kim MJ, Lee KS, Chon CY, Kim SJ, Terai S, Sakaida I, Han KH. Autologous bone marrow infusion activates the progenitor cell compartment in patients with advanced liver cirrhosis. *Cell Transplant*. 2010;19(10):1237–46.
26. Wang TH, Chen TC, Teng X, Liang KH, Yeh CT. Automated biphasic morphological assessment of hepatitis B-related liver fibrosis using second harmonic generation microscopy. *Sci Rep*. 2015;5:12962.
27. Pankaj P, Zhang Q, Bai XL, Liang TB. Autologous bone marrow transplantation in decompensated liver: Systematic review and meta-analysis. *World J Gastroenterol*. 2015; 21(28):8697–710.
28. Yi HZ, Chen JJ, Cen H, Yan W, Tan XH. Association between infection of hepatitis B virus and onset risk of B-cell non-Hodgkin's lymphoma: A systematic review and a meta-analysis. *Med Oncol*. 2014;31(8):84.
29. Fattovich G, Stroffolini T, Zagni I, Donato F. Hepatocellular carcinoma in cirrhosis: Incidence and risk factors. *Gastroenterology* 2004;127(5 Suppl 1):S35–50.
30. Tsai JF, Jeng JE, Ho MS, Chang WY, Hsieh MY, Lin ZY, Tsai JH. Effect of hepatitis C and B virus infection on risk of hepatocellular carcinoma: A prospective study. *Br J Cancer* 1997;76(7):968–74.