

Invited Mini Review

Mitophagy: a balance regulator of NLRP3 inflammasome activation

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The NLRP3 inflammasome is activated by a variety of external or host-derived stimuli and its activation initiates an inflammatory response through caspase-1 activation, resulting in inflammatory cytokine IL-1 β maturation and secretion. The NLRP3 inflammasome activation is a kind of innate immune response, most likely mediated by myeloid cells acting as a host defense mechanism. However, if this activation is not properly regulated, excessive inflammation induced by over-activated NLRP3 inflammasome can be detrimental to the host, causing tissue damage and organ dysfunction, eventually causing several diseases. Previous studies have suggested that mitochondrial damage may be a cause of NLRP3 inflammasome activation and autophagy, which is a conserved self-degradation process that negatively regulates NLRP3 inflammasome activation. Recently, mitochondria-selective autophagy, termed mitophagy, has emerged as a central player for maintaining mitochondrial homeostasis through the elimination of damaged mitochondria, leading to the prevention of hyperinflammation triggered by NLRP3 inflammasome activation. In this review, we will first focus on the molecular mechanisms of NLRP3 inflammasome activation and NLRP3 inflammasome-related diseases. We will then discuss autophagy, especially mitophagy, as a negative regulator of NLRP3 inflammasome activation by examining recent advances in research. [BMB Reports 2016; 49(10): 529-535]

INTRODUCTION

The immune system is a host protection mechanism against invading pathogens, such as viruses and bacteria. Macrophages, the front line defenders who recognize invading pathogens,

not only kill pathogens by phagocytosis, but also initiate inflammation producing inflammatory cytokines upon infections. The maturation and secretion of IL-1 β , one of the strongest pro-inflammatory cytokines, is enhanced by the inflammasome in macrophages, leading to inflammation and cell death (1, 2).

Inflammasomes are molecular platforms that trigger activation of caspase-1, leading to pro-inflammatory cytokine maturation. One of the best characterized inflammasome is the NLRP3 inflammasome. Even though NLRP3 inflammasome activation is important for protecting cells from pathogenic microbes, excessive NLRP3 inflammasome may cause hyperinflammation resulting in tissue damage and organ failure. NLRP3 inflammasome is also activated by danger signals released from stressed cells and dysregulation of NLRP3 inflammasome activation causes many diseases, such as neurodegenerative diseases, metabolic diseases and sepsis. Therefore, mechanisms to control NLRP3 inflammasome activation are necessary for health.

Upon bacterial infection, innate immune cells, such as macrophages and neutrophils, take up bacteria and generate reactive oxygen species (ROS) (3-5). The 2 main sources of ROS are NADPH oxidase-mediated ROS and mitochondria-derived ROS in innate immune cells (6-8). Some studies have proposed that mitochondrial dysfunction is closely associated with NLRP3 inflammasome activation. Infection causes mitochondrial damage through an unknown mechanism and the damaged mitochondria release mitochondrial DNA (mtDNA) and mitochondrial reactive ROS (mtROS) and are thought to work as danger signals (9, 10). Cells have a regulatory mechanism, 'mitophagy', a mitochondria-selective autophagic process to eliminate damaged or unwanted mitochondria, to maintain mitochondrial homeostasis against stress. This review will discuss how the balance between pro-inflammatory response and anti-inflammatory response is maintained in cells focusing on NLRP3 inflammasome activation and mitophagy.

THE INFLAMMASOME

Inflammation is one of the first innate immune responses to

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combat foreign organisms entering the body that are able to cause disease (11). Most foreign organisms express various pathogen-associated molecular patterns (PAMPs) in their cell wall or on their cell surface (12). Therefore, it is possible that a host can distinguish between self and non-self by recognizing PAMPs. Moreover, under stressed conditions, cells release danger signals called danger-associated molecular patterns (DAMPs) to notify their urgent situation to the immune system. PAMPs and DAMPs are recognized by innate immune receptors pattern-recognition receptors (PRRs), such as Toll-like receptors (TLRs) and nucleotide-binding and oligomerization domain (NOD)-like receptors (NLRs) (13). Activation of PRRs lead to NF- κ B activation and inflammatory cytokine production. Many studies have shown that the inflammasome is a key regulator of inflammation (1, 14-17). Most inflammasomes contain NLR proteins and the NLR family can be classified into 3 subtypes based on their domain structures: the NODs (NOD1-5 and CIITA), the NLRPs (NLRP1-14) and the IPAF (IPAF also known as NLRC4, and NAIP) subtypes (18). The NLR family commonly contains a central nucleotide-binding and oligomerization (NACHT) domain. Most NLRs also contain C-terminal leucine-rich repeats (LRRs) and N-terminal caspase recruitment domains (CARD) or pyrin domains (PYD). NACHT plays an essential role in the activation and formation of signaling complexes and LRR mediates ligand sensing, while CARD and PYD function in homotypic protein-protein interactions for downstream events (18, 19). A non-NLR inflammasome member, absent in melanoma-2 (AIM2), was later identified that can form an inflammasome composed of AIM2, ASC and caspase-1. The function and mechanism of many NLR family members remain poorly understood, but some are well-characterized. For example, NOD1 and NOD2 are receptors for bacterial peptidoglycan fragments. NOD1 and NOD2 recognize D-glutamyl-meso-diaminopimelic acid (DAP) and muramyl dipeptide (MDP), respectively. Upon sensing their ligand, NOD1 and NOD2 oligomerize and recruit RIP2 via CARD-CARD interactions, triggering an inflammatory response (20, 21). CIITA uniquely acts as a transcription factor playing a key role in the regulation of class II MHC genes (22). The IPAF inflammasome is activated in response to gram-negative bacteria containing type III or IV secretion systems, such as *Pseudomonas aeruginosa*, *Salmonella typhimurium* and *Shigella flexneri* (23-25). The AIM2 inflammasome is activated by sensing cytosolic double-stranded DNA (dsDNA) through the HIN-200 domain of AIM2 (26, 27). As is demonstrated herein, each inflammasome is activated by different stimuli, but more studies concerning other NLR family members are required. In this review, the NLRP3 inflammasome, which is the most characterized of the inflammasome members, will be discussed in detail.

NLRP3 INFLAMMASOME

The NLRP3 inflammasome is activated in response to abundant

stimuli and 2 distinct steps are required for this activation. First, Nuclear factor- κ B (NF- κ B) activation by TLR ligands increases NLRP3 and pro-IL-1 β ; this step is called 'priming'. Second, activating stimuli derived from microbes or the host induce the assembly of NLRP3 inflammasome components (17). The NLRP3 inflammasome is composed of NLRP3, apoptosis-associated speck-like protein containing a CARD (ASC) and pro-caspase-1. ASC is an inflammasome adaptor containing N-terminal PYD and C-terminal CARD. NLRP3 inflammasome is assembled by the following steps: PYD of NLRP3 is oligomerized with ASC through PYD-PYD interactions. Subsequently, pro-caspase-1 is recruited and interacts with ASC through CARD-CARD interactions (28). After the formation of the NLRP3 inflammasome, pro-caspase-1 is autocleaved to active caspase-1 and the active caspase-1 matures cytokines, such as IL-1 β , to their bioactive and secreted forms (17). Currently, various sterile DAMP signals and pathogens, including mitochondrial ROS, mitochondrial DNA, potassium efflux, MDP, monosodium urate (MSU), cholesterol crystals, cathepsins, influenza virus, *Salmonella typhimurium*, *Mycobacterium tuberculosis* and others, are known to activate the NLRP3 inflammasome (29). Compared to NLRP3 inflammasome activators, negative regulators of the NLRP3 inflammasome are relatively less known. Recent studies suggest that nitric oxide (NO), Ca²⁺ and cyclic AMP negatively regulate the NLRP3 inflammasome but the detailed mechanisms are unclear (30, 31). If the upregulated NLRP3 inflammasome activation is not downregulated, inflammation will be persistently induced by NLRP3 inflammasome and contribute to a diverse array of diseases.

NLRP3 INFLAMMASOME IN DISEASE; NEURODEGENERATIVE DISEASE

Dysregulation of NLRP3 inflammasome activation is observed in neurodegenerative disease. Alzheimer's disease (AD), a representative neurodegenerative disease, is characterized by amyloid- β plaque accumulation. Halle et al., using *in vitro* and *in vivo* mouse models, identified that amyloid- β causes lysosomal damage releasing cathepsin B and the cathepsin B activated NLRP3 inflammasome (32). In addition to the mouse study, recently performed human studies showed that increased active caspase-1 expression is observed in the brains of AD patients (33). It has been reported that not only AD but also Parkinson's disease (PD) are associated with the NLRP3 inflammasome. Accumulation of Lewy bodies (LB) formed by α -synuclein (α Syn) aggregation is a main pathogenesis of PD. A recent study revealed that α Syn induces synthesis of pro-IL-1 β by an interaction with TLR2 and activates NLRP3 inflammasome resulting in caspase-1 activation and IL-1 β maturation in human primary monocytes (34). In contrast, it is reported that mitochondrial dysfunction may lead to neurodegenerative disease (35). These studies indicate that both the NLRP3 inflammasome and mitochondrial dysfunction are

involved in neurodegenerative disease though more studies are required to clarify the relationship among NLRP3 inflammasome, mitochondria and neurodegenerative diseases.

NLRP3 INFLAMMASOME IN DISEASE; METABOLIC DISORDER

It has been known that a High fat diet (HFD) can induce Type 2 diabetes (T2D) and obesity and chronic inflammation is thought to contribute to T2D and obesity (36). As such, there is a possibility that the NLRP3 inflammasome, which can lead to chronic inflammation, is related to these metabolic diseases. Recently Lee *et al.* observed the activated NLRP3 inflammasome in T2D patients and found that mitochondrial ROS is involved in this phenomenon (37). Other groups showed that saturated fatty acid-induced inflammation causes mitochondrial impairment in adipocytes, while saturated fatty acids can upregulate NLRP3 inflammasome activation (38, 39). Also, a higher level of NLRP3 inflammasome components were detected in obese patients, and active caspase-1 were increased with obesity development in adipose tissues (40). Interestingly, caspase-1 deficient mice gained less weight and they had less adipose tissue formation in the HFD-induced obesity mouse model (41). These data strongly suggest that the NLRP3 inflammasome and mitochondrial dysfunction are closely linked to metabolic disorders.

NLRP3 INFLAMMASOME IN DISEASE NLRP3; SEPSIS

Sepsis is a life-threatening systemic inflammatory condition caused by a host immune response to microbial infection. Normally, increased inflammatory response to infection should be resolved in a timely manner, but when it gets out of control, exaggerated activation of the inflammasome produces excessive inflammatory cytokines leading to sepsis. A few studies provided evidence that mitochondrial dysfunction is associated with sepsis. Damaged mitochondria of the cells treated with NLRP3 inflammasome activators release danger signals, such as mtROS and mtDNA, and these signals can activate the NLRP3 inflammasome (9, 10). Supporting these, increased mtDNA and inflammatory cytokines were detected in septic plasma samples (42). This evidence shows that both the NLRP3 inflammasome and mitochondria contribute to sepsis.

MITOCHONDRIA AND NLRP3 INFLAMMASOME

Mitochondria are double membrane organelles that play pivotal roles in cells, including energy production and regulation of cell death. In addition, recent studies have revealed that mitochondria are also involved in the innate immune response (43, 44). Mitochondria are the main sites of reactive oxygen species (ROS) production. Mitochondrial ROS are not just byproducts of aerobic respiration, but also

participate in signaling pathways. However, overproduced ROS can cause cell damage and contribute to pathologies. Some studies have shown that ATP and monosodium urate (MSU) crystals increase ROS production and these activate the inflammasome (45, 46). Silica and asbestos also increase ROS production and activate the NLRP3 inflammasome (47). In another study, increased mtROS by the inhibition of mitochondrial complex I or complex III, triggered NLRP3 inflammasome activation, indicating mtROS can be a main cause of NLRP3 inflammasome activation (48, 49). mtDNA also activates NLRP3 inflammasome. Under normal conditions, mtDNA is located in mitochondria; however, when mitochondria are damaged, mtDNA is released into the cytoplasm. Shimada *et al.* found that oxidized mtDNA released from damaged mitochondria directly induced the NLRP3 inflammasome (10). Mitochondrial outer membrane proteins are also thought to activate the NLRP3 inflammasome. Mitofusins, mitochondrial outer membrane proteins that are required for mitochondrial fusion, activate the NLRP3 inflammasome in influenza and encephalomyocarditis virus (EMCV) infection, while another mitochondrial outer membrane protein MAVS recruits NLRP3 to mitochondria and activates inflammasome activation in response to ATP, nigericin and poly I:C (50). All of these data indicate that mitochondria play a crucial role in NLRP3 inflammasome activation.

AUTOPHAGY AND NLRP3 INFLAMMASOME

As described above, there is a lot of evidence to suggest that damaged mitochondria contribute to disease, including NLRP3 inflammasome-related diseases. Therefore, the importance of mitophagy, which is a process to remove damaged mitochondria, can be considered as a key process to regulate NLRP3 inflammasome activation. Despite the importance of mitophagy, it has mostly only been studied in mitochondria-related gene overexpressed mammalian cell lines in recent years. To investigate its mechanism and function accurately, more *in vitro* and *in vivo* studies are required. Knowledge of macroautophagy (hereafter autophagy) is a prerequisite to understanding mitophagy. Autophagy is a self-degradation process to recycle cellular components under stressed conditions, such as lack of nutrients. Autophagy has been regarded as a nonselective process in the past; however, accumulating evidence shows that it can be selective to remove specific proteins and damaged organelles, such as mitochondria under certain conditions (51-53). Autophagy consists of 4 sequential steps: initiation of autophagosome formation, elongation and closure of autophagic membrane, fusion between autophagosome and lysome, and degradation. In many cellular cases, autophagic induction is regulated by the *mammalian* target of rapamycin (mTOR) and the AMP-activated protein kinase (AMPK). mTOR inactivates unc-51-like kinase 1/2 (ULK1/2) by phosphorylation under basal conditions, but mTOR is inhibited under stress

conditions, allowing ULK1/2 to be modified to their active forms by AMPK phosphorylation initiating autophagy. Next, VPS34 lipid kinase complex and phosphatidylinositol 3-phosphate (PI3P) are recruited to complete autophagosome formation. In the step of elongation and closure of the autophagic membrane, the ubiquitin-like proteins, ATG12 and ATG5, are conjugated and then the conjugated forms an E3-like ligase complex with ATG16L1. After that, ATG8 (LC3 and GABARAP subfamilies) is conjugated to the lipid phosphatidylethanolamine by the ATG16L1 ligase complex and in the case of LC3, LC3-I form changes to LC3-II form, which is often used as an autophagosome marker (44, 54). VPS34-Beclin 1 complex containing ultraviolet radiation resistance associated gene protein (UVRAG) is involved in the step of fusion between the autophagosome and lysosome. Once the autophagosome and lysosome are fused to the autolysosome, the cargo in the autolysosome is degraded through lysosomal hydrolase activity (55). When there are certain proteins or organelles to be eliminated, they are ubiquitinated by E3 ubiquitin ligases and the ubiquitin chains serve as 'eat-me' signals. Then, adaptor proteins, such as p62 and NBR1, which contain a ubiquitin-associated UBA domain and LC3-interacting region (LIR), capture the ubiquitinated cargos and recruit LC3-II to proceed to selective autophagy (54). Accumulating evidence has shown that autophagy is involved in the innate immune response and NLRP3 inflammasome activation. Inhibition of autophagy by 3MA, which is a PI3K inhibitor increased mtROS production resulting in NLRP3-dependent IL-1 β secretion in the absence of inflammasome stimuli (56, 57). Similarly, treatment of 3MA blocking autophagosome formation in *Mycobacterium tuberculosis*-infected cells led to increased IL-1 β secretion (58). Consistent with these data, bone marrow-derived macrophages from LC3 knockout mice and Beclin 1 knockout mice had more damaged mitochondria producing mtROS, and had more caspase-1 activation leading to increased IL-1 β secretion upon NLRP3 inflammasome activation (9). Also, Atg16L1 deficient mice were more susceptible to dextran sulfate sodium-induced colitis as a result of enhanced inflammasome activation (58). These data show that autophagy negatively regulates inflammasome activation. Moreover, as damaged mitochondria are detected at higher levels under autophagy inhibition (9, 56, 57), it is plausible to assume that the elimination of damaged mitochondria by autophagy is important to prevent excessive NLRP3 inflammasome activation.

MITOPHAGY AND NLRP3 INFLAMMASOME

As mentioned above, mitochondrial damage causes NLRP3 inflammasome, and mitophagy can remove damaged mitochondria selectively (Fig. 1). Since mitochondria are evolved from ancient bacteria, a comparison of differences between mitophagy and intracellular bacteria-selective autophagy referred to as xenophagy can provide valuable information to better

understand mitophagy. Distinguishing between self and non-self is achieved by different 'eat-me' signals and cargo receptors (52, 59). To date, at least 4 cargo receptors have been studied in detail: NDP52, p62, NBR1 and Optineurin (59). Invading bacteria can damage their host vacuoles in which the bacteria replicate exposing glycans from the host vacuoles, and the glycans are recognized by a danger receptor, Galectin-8. Galectin-8 functions as an 'eat-me' signal and interacts with NDP52 inducing xenophagy by recruiting LC3 attached to phagophores (60). NDP52 can also directly bind to ubiquitinated bacteria and recruit the autophagic machinery. Similarly, p62, NBR1 and Optineurin serves to connect ubiquitinated bacteria with LC3, facilitating autophagy. How the engulfed bacteria are ubiquitinated is still largely unknown but LRSAM1 has been identified as an E3 ubiquitin ligase that ubiquitinates *Salmonella typhimurium* recruiting cargo receptors (61). Another E3 ubiquitin ligase, PARKIN, has also been reported to be involved in xenophagy. PARKIN deficient mice and flies were susceptible to intracellular bacterial infections inducing less autophagy, and these indicate that PARKIN plays a crucial role in xenophagy (62). Interestingly, PARKIN is the best known E3 ubiquitin ligase in mitophagy. The PINK-PARKIN pathway is important in the mitophagic process (63, 64). PINK1 is a serine/threonine kinase containing a mitochondrial targeting sequence in the N-terminus.

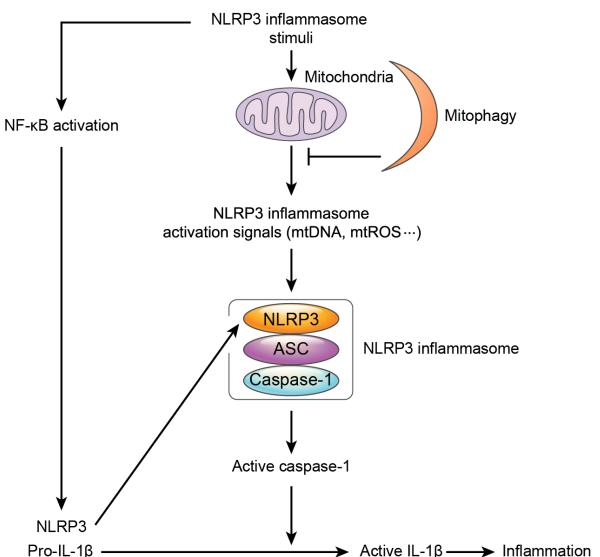


Fig. 1. Mitophagy downregulates aberrant NLRP3 inflammasome activation. NLRP3 inflammasome stimuli activates NF-κB producing NLRP3 and pro-IL-1 β . Mitochondria are damaged by NLRP3 inflammasome stimuli and release NLRP3 inflammasome activating signals, such as mtDNA and mtROS, triggering an inflammatory response. Mitophagy downregulates NLRP3 inflammasome activation by eliminating damaged mitochondria, blocking NLRP3 inflammasome activating signals.

Normally, PINK1 is imported into the mitochondria and anchored at the inner mitochondrial membrane (IMM) and then degraded by mitochondrial proteases. However, when mitochondria are damaged, PINK1 can't be imported to IMM, but instead accumulates on the outer mitochondria membrane (OMM) (65). The accumulated PINK1 recruits a cytosolic E3 ubiquitin ligase PARKIN and activates PARKIN by phosphorylation. Activated PARKIN ubiquitinates certain mitochondrial proteins or PARKIN itself. Then, the mitochondrial ubiquitination acts as an 'eat-me' signal and can be recognized by the adaptor p62 through its ubiquitin binding domain. Since p62 also has an LC3 interacting motif, p62 binds with LC3 on the autophagosome and facilitates the degradation of damaged mitochondria (44). However, not all mitophagy is processed by the PINK-PARKIN pathway. Since mitochondrial proteins BCL2/adenovirus E1B 19kDa interacting protein 3 (BNIP3), NIP3-like protein X (NIX/BNIP3L) and FUN14 domain containing 1 (FUNDC1) can directly interact with LC3-II, it is thought that other mechanisms exist to enhance mitophagy (66-69). So far we have discussed how mitochondria play a crucial role in the NLRP3 inflammasome activation, however not many studies have been performed concerning the detailed molecular mechanisms of mitophagy. Recently, some studies have shown how mitophagy is related to the control of NLRP3 inflammasome activation. Zhong *et al.* found that the cargo receptor p62 is increased by NF- κ B signaling, and that the increased p62 is translocated to damaged mitochondria, which are ubiquitinated by Parkin and induce mitophagy (70). Consistent with this finding, Kim *et al.* also observed that p62 is increased and translocated to damaged mitochondria in NLRP3 inflammasome activated cells. Not only p62 but also autophagic inducer Sestrin 2 (SESN2) is increased by NO and is translocated to damaged mitochondria and protects cells from hyperinflammation inducing mitophagy. Additionally, SESN2 increases ULK1 stability leading to the initiation of autophagy (71). These studies indicate that mitophagy is one of the self-limiting systems to protect cells from excessive inflammation.

CONCLUSION

In the past decade, there have been great advances in understanding of the NLRP3 inflammasome and autophagy. However, knowledge of the link between the NLRP3 inflammasome and autophagy, especially mitophagy, is poorly understood. Nevertheless we have discussed the role of mitophagy as a negative regulator of aberrant NLRP3 inflammasome activation, while detailed mechanisms remain largely unknown. There are questions that remain to be answered. Mitochondrial damage can activate the NLRP3 inflammasome, but inflammatory cytokines can also trigger mitochondrial damage, so it is ambiguous as to which one comes first. Since NLRP3 inflammasome activation has a beneficial effect so how to regulate activation appropriately it

should be considered for clinical application. Also, how damaged mitochondria serve as 'eat-me' signals in the case where PINK1 is not mediated should be investigated. Further studies are required for a better understanding of the molecular mechanisms of mitophagy, but mitochondria remain possible therapeutic targets to protect cells from aberrant inflammation.

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REFERENCES

1. Strowig T, Henao-Mejia J, Elinav E and Flavell R (2012) Inflammasomes in health and disease. *Nature* 481, 278-286
2. Lamkanfi M and Dixit VM (2012) Inflammasomes and their roles in health and disease. *Annu Rev Cell Dev Biol* 28, 137-161
3. Rosen H and Klebanoff SJ (1979) Bactericidal activity of a superoxide anion-generating system. A model for the polymorphonuclear leukocyte. *J Exp Med* 149, 27-39
4. Chen X, Lee KA, Ren X *et al* (2016) Synthesis of a highly HOCl-selective fluorescent probe and its use for imaging HOCl in cells and organisms. *Nat Protoc* 11, 1219-1228
5. Yin J, Kwon Y, Kim D *et al* (2015) Preparation of a cyanine-based fluorescent probe for highly selective detection of glutathione and its use in living cells and tissues of mice. *Nat Protoc* 10, 1742-1754
6. Joo JH, Ryu JH, Kim CH *et al* (2012) Dual oxidase 2 is essential for the toll-like receptor 5-mediated inflammatory response in airway mucosa. *Antioxid Redox Signal* 16, 57-70
7. Kim MJ, Ryu JC, Kwon Y *et al* (2014) Dual oxidase 2 in lung epithelia is essential for hyperoxia-induced acute lung injury in mice. *Antioxid Redox Signal* 21, 1803-1818
8. Pelletier M, Lepow TS, Billingham LK, Murphy MP and Siegel RM (2012) New tricks from an old dog: mitochondrial redox signaling in cellular inflammation. *Semin Immunol* 24, 384-392
9. Nakahira K, Haspel JA, Rathinam VA *et al* (2011) Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat Immunol* 12, 222-230
10. Shimada K, Crother TR, Karlin J *et al* (2012) Oxidized mitochondrial DNA activates the NLRP3 inflammasome during apoptosis. *Immunity* 36, 401-414
11. Medzhitov R (2007) Recognition of microorganisms and activation of the immune response. *Nature* 449, 819-826
12. Akira S, Uematsu S and Takeuchi O (2006) Pathogen recognition and innate immunity. *Cell* 124, 783-801
13. Takeuchi O and Akira S (2010) Pattern recognition receptors and inflammation. *Cell* 140, 805-820
14. Wen H, Miao EA and Ting JP (2013) Mechanisms of NOD-like receptor-associated inflammasome activation. *Immunity* 39, 432-441

15. Lamkanfi M and Dixit VM (2014) Mechanisms and functions of inflammasomes. *Cell* 157, 1013-1022
16. Vanaja SK, Rathinam VA and Fitzgerald KA (2015) Mechanisms of inflammasome activation: recent advances and novel insights. *Trends Cell Biol* 25, 308-315
17. Guo H, Callaway JB and Ting JP (2015) Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nat Med* 21, 677-687
18. Schroder K and Tschoop J (2010) The inflammasomes. *Cell* 140, 821-832
19. Latz E, Xiao TS and Stutz A (2013) Activation and regulation of the inflammasomes. *Nat Rev Immunol* 13, 397-411
20. Kufer TA, Banks DJ and Philpott DJ (2006) Innate immune sensing of microbes by Nod proteins. *Ann N Y Acad Sci* 1072, 19-27
21. Moreira LO and Zamboni DS (2012) NOD1 and NOD2 Signaling in Infection and Inflammation. *Front Immunol* 3, 328
22. Masternak K, Muhlethaler-Mottet A, Villard J, Zufferey M, Steimle V and Reith W (2000) CIITA is a transcriptional coactivator that is recruited to MHC class II promoters by multiple synergistic interactions with an enhanceosome complex. *Genes Dev* 14, 1156-1166
23. Franchi L, Stoolman J, Kanneganti TD, Verma A, Ramphal, R and Nunez G (2007) Critical role for Ipaf in *Pseudomonas aeruginosa*-induced caspase-1 activation. *Eur J Immunol* 37, 3030-3039
24. Mariathasan S, Newton K, Monack DM et al (2004) Differential activation of the inflammasome by caspase-1 adaptors ASC and Ipaf. *Nature* 430, 213-218
25. Suzuki T, Franchi L, Toma C et al (2007) Differential regulation of caspase-1 activation, pyroptosis, and autophagy via Ipaf and ASC in *Shigella*-infected macrophages. *PLoS Pathog* 3, e111
26. Hornung V, Ablasser A, Charrel-Dennis M et al (2009) AIM2 recognizes cytosolic dsDNA and forms a caspase-1-activating inflammasome with ASC. *Nature* 458, 514-518
27. Roberts TL, Idris A, Dunn JA et al (2009) HIN-200 proteins regulate caspase activation in response to foreign cytoplasmic DNA. *Science* 323, 1057-1060
28. Lu A, Magupalli VG, Ruan J et al (2014) Unified polymerization mechanism for the assembly of ASC-dependent inflammasomes. *Cell* 156, 1193-1206
29. Franchi L, Munoz-Planillo R and Nunez G (2012) Sensing and reacting to microbes through the inflammasomes. *Nat Immunol* 13, 325-332
30. Hernandez-Cuellar E, Tsuchiya K, Hara H et al (2012) Cutting edge: nitric oxide inhibits the NLRP3 inflammasome. *J Immunol* 189, 5113-5117
31. Lee GS, Subramanian N, Kim AI et al (2012) The calcium-sensing receptor regulates the NLRP3 inflammasome through Ca²⁺ and cAMP. *Nature* 492, 123-127
32. Halle A, Hornung V, Petzold GC et al (2008) The NALP3 inflammasome is involved in the innate immune response to amyloid-beta. *Nat Immunol* 9, 857-865
33. Heneka MT, Golenbock DT and Latz E (2015) Innate immunity in Alzheimer's disease. *Nat Immunol* 16, 229-236
34. Codolo G, Plotegher N, Pozzobon T et al (2013) Triggering of inflammasome by aggregated alpha-synuclein, an inflammatory response in synucleinopathies. *PLoS One* 8, e55375
35. Voloboueva LA and Giffard RG (2011) Inflammation, mitochondria, and the inhibition of adult neurogenesis. *J Neurosci Res* 89, 1989-1996
36. Hotamisligil GS (2006) Inflammation and metabolic disorders. *Nature* 444, 860-867
37. Lee HM, Kim JJ, Kim HJ, Shong M, Ku BJ and Jo EK (2013) Upregulated NLRP3 inflammasome activation in patients with type 2 diabetes. *Diabetes* 62, 194-204
38. Wen H, Gris D, Lei Y et al (2011) Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nat Immunol* 12, 408-415
39. Vandana Magsar B, Youm YH, Ravussin A et al (2011) The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat Med* 17, 179-188
40. Yin Z, Deng T, Peterson LE et al (2014) Transcriptome analysis of human adipocytes implicates the NOD-like receptor pathway in obesity-induced adipose inflammation. *Mol Cell Endocrinol* 394, 80-87
41. Stienstra R, Joosten LA, Koenen T et al (2010) The inflammasome-mediated caspase-1 activation controls adipocyte differentiation and insulin sensitivity. *Cell Metab* 12, 593-605
42. Nakahira K, Kyung SY, Rogers AJ et al (2013) Circulating mitochondrial DNA in patients in the ICU as a marker of mortality: derivation and validation. *PLoS Med* 10, e1001577; discussion e1001577
43. Gurung P, Lukens JR and Kanneganti TD (2015) Mitochondria: diversity in the regulation of the NLRP3 inflammasome. *Trends Mol Med* 21, 193-201
44. Lazarou M (2015) Keeping the immune system in check: a role for mitophagy. *Immunol Cell Biol* 93, 3-10
45. Cruz CM, Rinna A, Forman HJ, Ventura AL, Persechini PM and Ojcius DM (2007) ATP activates a reactive oxygen species-dependent oxidative stress response and secretion of proinflammatory cytokines in macrophages. *J Biol Chem* 282, 2871-2879
46. Petrelli V, Papin S, Dostert C, Mayor A, Martinon F and Tschoop J (2007) Activation of the NALP3 inflammasome is triggered by low intracellular potassium concentration. *Cell Death Differ* 14, 1583-1589
47. Dostert C, Petrelli V, Van Bruggen R, Steele C, Mossman BT and Tschoop J (2008) Innate immune activation through Nalp3 inflammasome sensing of asbestos and silica. *Science* 320, 674-677
48. Huang LS, Cobessi D, Tung EY and Berry EA (2005) Binding of the respiratory chain inhibitor antimycin to the mitochondrial bc1 complex: a new crystal structure reveals an altered intramolecular hydrogen-bonding pattern. *J Mol Biol* 351, 573-597
49. Li N, Ragheb K, Lawler G et al (2003) Mitochondrial complex I inhibitor rotenone induces apoptosis through enhancing mitochondrial reactive oxygen species production. *J Biol Chem* 278, 8516-8525
50. Subramanian N, Natarajan K, Clatworthy MR, Wang Z and Germain RN (2013) The adaptor MAVS promotes NLRP3 mitochondrial localization and inflammasome

- activation. *Cell* 153, 348-361
51. Komatsu M and Ichimura Y (2010) Selective autophagy regulates various cellular functions. *Genes Cells* 15, 923-933
52. Johansen T and Lamark T (2011) Selective autophagy mediated by autophagic adapter proteins. *Autophagy* 7, 279-296
53. Saito T and Sadoshima J (2015) Molecular mechanisms of mitochondrial autophagy/mitophagy in the heart. *Circ Res* 116, 1477-1490
54. Abounit K, Scarabelli TM and McCauley RB (2012) Autophagy in mammalian cells. *World J Biol Chem* 3, 1-6
55. Deretic V, Saitoh T and Akira S (2013) Autophagy in infection, inflammation and immunity. *Nat Rev Immunol* 13, 722-737
56. Harris J, Hartman M, Roche C et al (2011) Autophagy controls IL-1 β secretion by targeting pro-IL-1 β for degradation. *J Biol Chem* 286, 9587-9597
57. Zhou R, Yazdi AS, Menu P and Tschopp J (2011) A role for mitochondria in NLRP3 inflammasome activation. *Nature* 469, 221-225
58. Shi CS, Shenderov K, Huang NN et al (2012) Activation of autophagy by inflammatory signals limits IL-1 β production by targeting ubiquitinated inflammasomes for destruction. *Nat Immunol* 13, 255-263
59. Boyle KB and Randow F (2013) The role of 'eat-me' signals and autophagy cargo receptors in innate immunity. *Curr Opin Microbiol* 16, 339-348
60. Thurston TL, Wandel MP, von Muhlinen N, Foeglein A and Randow F (2012) Galectin 8 targets damaged vesicles for autophagy to defend cells against bacterial invasion. *Nature* 482, 414-418
61. Huet A, Heath RJ, Begun J et al (2012) The LRR and RING domain protein LRSAM1 is an E3 ligase crucial for ubiquitin-dependent autophagy of intracellular *Salmonella Typhimurium*. *Cell Host Microbe* 12, 778-790
62. Manzanillo PS, Ayres JS, Watson RO et al (2013) The ubiquitin ligase parkin mediates resistance to intracellular pathogens. *Nature* 501, 512-516
63. Narendra DP, Jin SM, Tanaka A et al (2010) PINK1 is selectively stabilized on impaired mitochondria to activate Parkin. *PLoS Biol* 8, e1000298
64. Vives-Bauza C, Zhou C, Huang Y et al (2010) PINK1-dependent recruitment of Parkin to mitochondria in mitophagy. *Proc Natl Acad Sci U S A* 107, 378-383
65. Jin SM, Lazarou M, Wang C, Kane LA, Narendra DP and Youle RJ (2010) Mitochondrial membrane potential regulates PINK1 import and proteolytic destabilization by PARL. *J Cell Biol* 191, 933-942
66. Schwartzen M, Mohrluder J, Ma P et al (2009) Nix directly binds to GABARAP: a possible crosstalk between apoptosis and autophagy. *Autophagy* 5, 690-698
67. Hanna RA, Quinsay MN, Orogo AM, Giang K, Rikka S and Gustafsson AB (2012) Microtubule-associated protein 1 light chain 3 (LC3) interacts with Bnip3 protein to selectively remove endoplasmic reticulum and mitochondria via autophagy. *J Biol Chem* 287, 19094-19104
68. Novak I, Kirkin V, McEwan DG et al (2010) Nix is a selective autophagy receptor for mitochondrial clearance. *EMBO Rep* 11, 45-51
69. Liu L, Feng D, Chen G et al (2012) Mitochondrial outer-membrane protein FUNDC1 mediates hypoxia-induced mitophagy in mammalian cells. *Nat Cell Biol* 14, 177-185
70. Zhong Z, Umemura A, Sanchez-Lopez E et al (2016) NF-kappaB Restricts Inflammasome Activation via Elimination of Damaged Mitochondria. *Cell* 164, 896-910
71. Kim MJ, Bae SH, Ryu JC et al (2016) SESN2/sestrin2 suppresses sepsis by inducing mitophagy and inhibiting NLRP3 activation in macrophages. *Autophagy* 12, 1272-1291