G1 Cyclin

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Rb-E2F E

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The Expression of G1 Cyclins and Rb-E2F, and the Effect of Vitamin E on Hepatic Stellate Cells Activated by CCl4

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Background/Aims: It is obscure when the activation of hepatic stellate cells and the expression of its related factors occur in acute liver injury. Vitamin E is expected to prevent hepatic fibrosis. The aims of this study were to establish the model of hepatic stellate cell activation in acute liver injury and to confirm the effect of vitamin E for preventing hepatic fibrosis. Methods: Male Sprague-Dawley rats were classified into two groups. The one group received a single injection of CCl4 and the other group received injection of vitamin E daily and a single injection of CCl4. The serial changes of serum ALT, and [3H]thymidine uptake, -SMA, cyclin D1, CDK4, cyclin E, CDK2, Rb, E2F-1 and NF- B of stellate cells were measured. Results: The serial changes of serum ALT levels, [3H]thymidine uptake, and -SMA positive cells showed maximum increase at 32 hours after CCl4 injection. However, they were significantly decreased with injection of vitamin E. CDK4, cyclin E and CDK2 showed definite band at 16, 32, 48 hours after CCl4 injection, which diminished or disappeared with injection of vitamin E. Cyclin D1, Rb, E2F-1 and NF- B showed definite band at 32 hours after CCl4 injection, which also diminished or disappeared with injection of vitamin E. Conclusions: We established an *in vivo* model of hepatic stellate cell activation in acute liver injury and confirmed the effect of vitamin E in preventing hepatic fibrosis. (Korean J Gastroenterol 2001;38:262-269)

Key Words: Hepatic stellate cell, Hepatic fibrosis, CCl₄, Vitamin E,

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dose of 200 mg/kg from 2 days before CCl4 injection (group 2). Five rats were killed before injection of CCl4, at 8 hr, 16 hr, 32

hr, 48 hr, and 60 hr after injection of CCl4, respectively. They received [3H]thymidine at a dose 30 µ Ci at 4 hr before sacrifice.

가 2 200 mg/kgE(-tocopherol, Sigma Chemical Co., 가 가 가 St. Louis, MO, U.S.A.) 가 , 16 , 32 60 5 가 E가 30 µ Ci 4 NF-kB [3H]thymidine 가 (Amersham Pharmacia Biotech, Buckinghamshire, England) (Fig. 1).5 2. cyclin D1, CDK (cyclin 1) CDK) 4, cyclin E, CDK2, Rb, dependent kinase, E2F-1 NF-kB (pronase) (collagenase)(Boehringer Mannheim, Mannheim, Germany) Hank's , 37 30 Е 가 Schwartz Nycodenz (Sigma Chemical Co., 3-4 St. Louis, MO, U.S.A.) (density (1,000 g, 4 , 20 gradient) 328 nm 1. oil red desmin 100 g Sprague-Dawley 95% . 1 -70 1:3 (vol:vol) 2 mL/kg, 2 2) 0.5% NP-40, 10 mM (NaF) 10 mM (Na pyrophosphate) 5% , 30% (citric acid) (sucrose) 1% (4,000 g, 4 , 30)-70 3) A LT Fig. 1. The classification according to the method. The Sprague-Dawley male rats each received a single intraperitoneal injection of 1 mLCCl4 in mineral oil (1:3, vol:vol) at a dose of 2 mL/kg only -70 ALT (Hitachi) (group 1) or with daily intraperitoneal injection of vitamin E at a

ALT

4) (block), 1 (Vector Laboratories, Burlingame, -SMA CA, U.S.A.) (biotin) 2 (DAKO, Glostrup, Denmark) (alkaline phosphatase)가 NBT-BCIP 가 2 400 -SMA 10 5) [3 H]thymidine (GIBCO BRL (Trizol) Life Technologies, Grand Island, NY, U.S.A.) (12,000 g, 4 , 15) 100% 가 DNA DNA (DNA pellet) 0.1 M (sodium citrate) 75% 8 mM(NaOH) OD 260 nm 10 (beta counter) μg 6) Western blot **RIPA** (10,000 g, 4 , 10) 10 μg (CDK2, CDK4, Rb) OD 595 nm (cyclin D1, cyclin E) , 10% (cyclin E, Rb) 16% (cyclin D1, CDK2, CDK4) (nitrocellulose membrane) 2 4 (nonfat milk) cyclin D1, CDK4, cyclin E, 1 Santa Cruz Biotechnology, Santa CDK2, Rb (Cruz, CA, U.S.A.) (DAKO) (chemiluminescence, NENTM Life Science,

7) Electrophoretic mobility shift assay (EMSA)

Boston, MA, U.S.A.)

NF-kB oligonucleotide (5'GGG GAC TTT CCC 3'), (band shift) polydIdC 6% $10\% \quad (acetic\ acid),\ 20\% \qquad ,$ $1 \qquad \qquad ,$ $1 \qquad \qquad ,$ $-70 \qquad \qquad \qquad ^{32}P$ oligonucleotide $_{4.5,7.8}$

1. ALT

2. [3H]thymidine

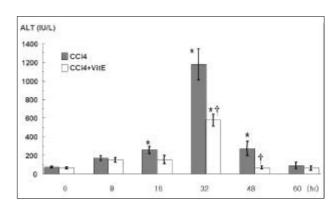


Fig. 2. The change of serum ALT level. Serum ALT level before injection of CCl₄, at 8 hr, 16 hr, 32 hr, 48 hr, and 60 hr after injection of CCl₄ showed 74 ± 20.7 , 170 ± 54.3 , 258 ± 83.5 , 1178 ± 381.3 , 274 ± 174.0 , 92 ± 74.0 (IU/L) in group 1, and 64 ± 21.9 , 152 ± 58.9 , 156 ± 62.3 , 576 ± 141.5 , 70 ± 35.4 , 62 ± 55.4 (IU/L) in group 2. Serum ALT level was significantly increased at 32 hr in group 1, but there was significant difference between group 1 and group 2.

*p<0.05 compared to time 0 (control) of each group.

[†] p<0.05 compared at the same time between group I & II.

[3 H] thymidine 5 1 76.4 \pm 14.7, 76.6 \pm 19.7, 78.8 \pm 23.8, 529.2 \pm 284.8, 299.0 \pm 161.6, 179.6 \pm 63.9 cpm , 71.6 \pm 19.9, 90.4 \pm 9.6, 85.0 \pm 24.0, 223.0 \pm 86.3, 171.2 \pm 47.8, 127.8 \pm 19.3 cpm 32 7 † 7 † (p<0.05), 1 2 [3 H]thymidine 7 † (p<0.05)(Fig. 3).

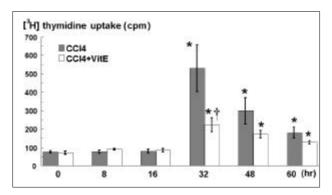


Fig. 3. The change of [3 H]thymidine level. [3 H]thymidine level before injection of CCl₄, at 8 hr, 16 hr, 32 hr, 48 hr, and 60 hr after injection of CCl₄ showed 76.4 ± 14.7 , 76.6 ± 19.7 , 78.8 ± 23.8 , 529.2 ± 284.8 , 299.0 ± 161.6 , 179.6 ± 63.9 (cpm) in group 1, and 71.6 ± 19.9 , 90.4 ± 9.6 , 85.0 ± 24.0 , 223.0 ± 86.3 , 171.2 ± 47.8 , 127.8 ± 19.3 (cpm) in group 2. [3 H]thymidine level was significantly increased at 32 hr in group 1, but there was significant difference between group 1 and group 2.

*p<0.05 compared to time 0 (control) of each group.

[†]p<0.05 compared at the same time between group I & II.

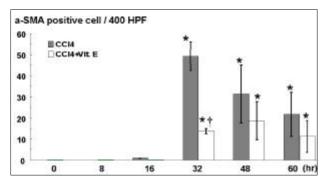


Fig. 4. The change of -SMA expression. -SMA in liver tissue before injection of CCl₄, at 8 hr, 16 hr, 32 hr, 48 hr, and 60 hr after injection of CCl₄ showed 0, 0, 2.0 ± 0.8 , 49.2 ± 6.8 , 31.2 ± 13.6 , 21.8 ± 10.1 (-SMA positive cell/400x high power field) in group 1, and 0, 0, 0, 14.8 ± 1.4 , 18.0 ± 9.1 , 12.2 ± 6.2 (-SMA positive cell/400x high power field) in group 2. -SMA activities was significantly increased at 32 hr in group 1, but there was significant difference between group 1 and group 2.

*p<0.05 compared to time 0 (control) of each group.

-SMA9 1 $0, 0, 2.0 \pm 0.8, 49.2$ ± 6.8 , 31.2 ± 13.6 , 21.8 ± 10.1 (-SMA) /400 $0, 0, 0, 14.8 \pm 1.4, 18.0 \pm$) , 2 9.1, 12.2 ± 6.2 16 가 32 (p<0.05),2 -SMA (p<0.05)(Fig. 4).

4. G1 cyclin

(Fig. 6).

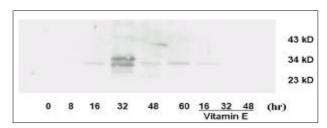


Fig. 5. The change of cyclin D1 expression. Cyclin D1 activities measured by Western blot were increased at 32 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2.

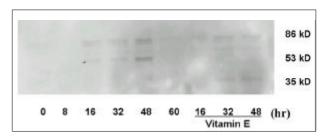
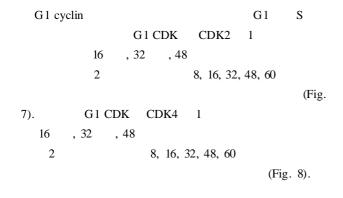


Fig. 6. The change of cyclin E expression. Cyclin E activities measured by Western blot were increased at 16 hr, 32 hr and 48 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2.

[†]p<0.05 compared at the same time between group I & II.

5. G1 CDK



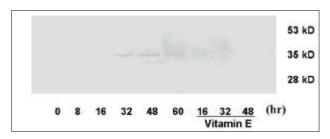


Fig. 7. The change of CDK2 expression. CDK2 activities measured by Western blot were increased at 16 hr, 32 hr and 48 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2.

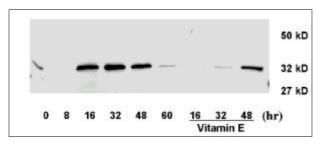


Fig. 8. The change of CDK4 expression. CDK4 activities measured by Western blot were increased at 16 hr, 32 hr and 48 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2.

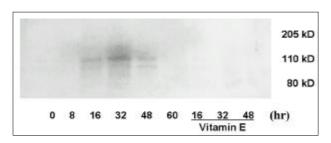


Fig. 9. The change of Rb expression. Rb activities measured by Western blot were increased at 32 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2.

6. Rb

	E2F	E2F	
Rb	CDK		E2F
,		Rb	Rb
1		32	
2		8, 16, 32, 48	8, 60
			(Fig. 9).
7. E2F-	1		
Rb		E21	F-1 Rb
	G1	S	
E2F-1 1		16	5 , 32

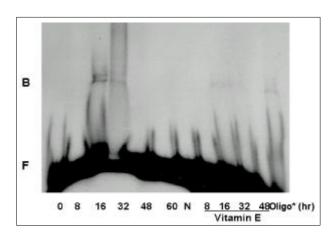


Fig. 10. The change of E2F-1 expression. E2F-1 measured by electrophoretic mobility shift assay were increased at 16 hr and 32 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2. The activities were decreased with unlabelled oligonucleotide (Oligo*).

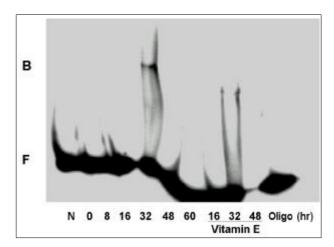


Fig. 11. The change of NF-kB expression. NF-kB measured by electrophoretic mobility shift assay were increased at 32 hr in group 1, but decreased or absent at 8 hr, 16 hr, 32 hr, 48 hr and 60 hr in group 2. The activities were decreased with unlabelled oligonucleotide (Oligo*).

2 8, 16, 가 32, 48, 60 oligonucleotide 가 가 (Fig. 10). 가 8. NF-kB 가 NF-kB NF-kB 1 32 2 8, 16, 32, 48, 60 NF-kB 가 oligonucleotide NF-kB (leukotriene), 가 (Fig. 11). 14,15 microsomal cyto-(trichloromethyl radical) chrome p-450 가 가 (trichloro-(cytokine) methyl peroxyl radical) (oxygen free radical) 가 (microtubule) 가 가 가 가 가 (glycoprotein) (proteoglycan) 가가 가 가 가 TGF-beta1 가 가 가 가 E가 가 가 E 가 NF-kB 가 ,11,12 .13 가 (myofibroblast) E (cytoskeletal filament) C В 가

G1 Cyclin

Rb-E2F

17,18

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E가
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         가
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                         cyclin
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                  G1
                                 cell division cycle
(CDC)
                                         (p53, Rb
                                                          Ε
                         (E1A, E2F1)
gene)
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                       cyclin A, B1, B2, C, D1, D2,
D3, E가
                          G1 G2
                                        S M
                                              G1
cyclin, G2 cyclin
                               cyclin
                                                                               cyclin D1, CDK4, cyclin E,
                        S
                              M
                                                      CDK2, Rb, E2F-1
                                                                        NF-kB
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                . G1 cyclin
                              A-type cyclin, D-type
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        E-type cyclin
                         CDK2 CDK4
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cyclin
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      D-type cyclin CDK4
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                    cyclin D
               cyclin D CDK4
                                   CDK6
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                                                                                   , 48
                     CDK
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       . Cyclin E-CDK2
                           cyclin D
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      E2F, Elf-1, c-Abl
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                             DNA
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                                                         ALT, [3H]thymidine
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            S
   G1
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                                                                          , cyclin D1, Rb, E2F-1, NF-kB
                                                      CDK4 16, 32, 48
cyclin G1 CDK
                                                      32
  cyclin D, cyclin E, CDK2, CDK4, Rb, E2F-1
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   16, 32, 48
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