Korea red ginseng restores impaired endothelial function in aged mice through inhibition of arginase activity

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A Masters Thesis Submitted to the Department of Medicine and the Graduate School of Yonsei University in partial fulfillment of the requirements for the degree of Master of medicine

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January 2014

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January 2014

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ABSTRACT

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Korean red ginseng water extract (KRGE) has reported beneficial effects on the cardiovascular system. Cardiovascular disease is the leading cause of morbidity and mortality and the incidence of cardiovascular disease is predicted to increase as the population ages. There is accumulating evidence that arginase upregulation is associated with impaired endothelial function in aged vasculature. Here, we demonstrate that oral administration of KRGE to aged mice inhibits increased arginase activity, restores NO generation, and reduces ROS production via enhancement of endothelial nitric oxide synthase coupling. In vascular tension assay, attenuated vasorelaxation responses to acetylcholine and reduced vasoconstriction responses to phenylephrine in aged vessels were significantly improved following administering KRGE. Furthermore, KRGE showed a preventative effect on formation of peroxynitrite in plasma of aged mice. Taken together, these results suggest that KRGE may exert vasoprotective effects through augmentation of NO signaling by inhibiting arginase activity. Therefore, KRGE may be useful in the treatment of vascular diseases associated with aging.

Keywords: *Panax ginseng*, Korean red ginseng extract, Aging, Arginase, Endothelial Nitric Oxide Synthase, Nitric Oxide, Vasorelaxation

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I. INTRODUCTION

Cardiovascular diseases are the leading cause of morbidity and mortality in both industrialized and developing countries. The occurrence of cardiovascular disease is predicted to increases as population ages, although effective treatments for several established cardiovascular risk factors, such as hypertension and hypercholesterolemia. The hallmark of the aging cardiovascular system is decrease in nitric oxide (NO) bioavailability ^{1, 2} and increase in reactive oxygen species (ROS) production ^{3, 4}. Superoxide (O_2^-) is a free radical that rapidly scavenges NO, thereby decreasing NO bioavailability. NO and O_2^- may react to produce peroxynitrite (ONOO⁻), a highly damaging ROS molecule. Thus, simultaneous generation of NO and O_2^- can raise ONOO⁻ to levels potentially detrimental to vascular cell function and viability ⁵. This nitroso-redox imbalance contributes to age-related endothelial dysfunction and vascular stiffness ⁶.

Korean red ginseng water extract (KRGE) has been extensively studied and its consumption progressively increased. Ginseng has shown beneficial effects to various diseases, including thrombosis, hyperlipidemia, cancer, and atherosclerosis ⁷⁻¹¹. In vascularture, it is well documented that KRGE has vasoprotective effects by eliminating superoxide derived from NADPH oxidase ¹², promoting endothelial cells proliferation and protecting from H₂O₂-dependent cell death ^{8, 13}, and inducing heme oxygenase-1 expression ¹⁴. Ginseng extract exerts a direct vasodilatory effect by releasing NO in endothelium-dependent manner ¹⁵. Furthermore, the beneficial effect of ginseng on vascular system may be dependent on activation of phosphoinositide 3-kinase (PI3K)/Akt signal transduction in endothelial cells ¹⁶.

Vascular changes associated with aging have been investigated in humans and a number of other species ¹⁷. However, the relative contributions of dysregulated mechanisms to age-related vascular pathology remain to be elucidated, because the contribution of vascular control mechanisms in health, aging, and disease conditions is influenced by vessel type and size ¹⁷. In previous study, we demonstrated that KRGE inhibited arginase activity and reciprocally regulates NO production and enhanced NO-dependent vasorelaxation in wild type young mice ¹⁸. Therefore, we, here, investigated which mechanisms contribute to age-related endothelial dysfunction in mice and determined whether orally administered-KRGE improves impaired endothelial function in aged mice.

II. MATERIALS AND METHODS

1. Materials

KRGE (solid extract 64%, gensenosideRg1+Rb1 4 mg/g) was obtained from Korea Ginseng Corporation (chuncheon, Korea) and directly dissolved in distilled water. MnTBAP(Mn(III) Tetra(4-benzoic acid) porphyrin chloride) and L-NAME(N^G-nitro-L-arginine methyl ester) were obtained from Calbiochem. All reagents were purchased from Sigma unless otherwise stated.

2. Animal protocol

Young (10±3 weeks) and aged (55±5 weeks) mice (C57BL/6J) were used for all experiments. Mice were housed at 23°C under a 12-h light/12-h dark cycle. The dark period was from10:00 to 22:00. All animals had access *ad libitum* to water and food (Nara Biotech.). The study protocols were in accordance with the Guide for the Care and Use of Laboratory Animals (Institutional Review Board, Kangwon National University). KRGE was orally administered for 4 weeks. Given that each mouse consumed approximately 10 ml water/day, this represented a daily dose of ~20 mg/mouse/day of KRGE

3. Arginase activity assay

Tissue lysates were prepared using lysis buffer (50 mM Tris-HCl, pH7.5, 0.1 mM EDTA and protease inhibitors) by homogenization at 4° C followed by centrifugation for 20 min at 14,000 x g at 4° C. The supernatants were used to assay for arginase activity as previously described ¹⁹.

4. Western blotting analysis

Aortic vessels from C57BL/6 mice were homogenized in the buffer (50 mM Tris-HCl, 150 mM NaCl, 1% Nonidet P-40, 1 mM EDTA, 1 μ g/ml of leupeptin, 1 μ g/ml of pepstatin, 1 μ g/ml of aprotinin, 1 mM phenylmethylsulfonylflouride, 1 mM sodium orthovanadate, and 1 mM NaF) and centrifuged for 30min at 14,000 x g. The protein

amount of the supernatant was analyzed by the Bradford method. Protein (100 µg) were separated in a 10% SDS-PAGE and then transferred to a nitrocellulose membrane (Bio-Rad). The blots were incubated with a polyclonal anti-arginase II (Santa Cruz), anti-endothelial nitric oxide synthase (eNOS, BD Bioscience), or anti-actin (Santa Cruz) antibodies followed by the secondary antibody (Amersham). The signals were detected using an enhanced chemiluminescence detection reagent with X-ray films.

5. Determination of eNOS dimerization

Dimers and monomers of eNOS were separated using low-temperature SDS-PAGE as previously described ²⁰. Band intensities were analyzed using NIH ImageJ Software.

6. Estimation of NO or ROS generation in isolated mice aorta using 4-amino-5methylamino-2',7'-difluorescein diacetate (DAF-FM) or dihydroethidine (DHE)

NO and ROS production were estimated using microscope by measuring change of fluorescence intensity at different time intervals as described previously ²¹.

7. Aortic vascular tension assay

The study was approved in accordance with Guide for the Care and Use of Laboratory Animals (Institutional Review Board, Kangwon national University). Male mice C57BL/6J were anesthetized using isoflurane and the thoracic aorta was rapidly removed. The aorta were placed on ice-cold oxygenated Krebs-Ringer bicarbonate solution (NaCl 118.3, KCl 4.7, MgSO4 1.2, KH2PO4 1.2, CaCl2 1.6, NaHCO3 25, glucose 11.1 (in mM)) and cleared off adherent connective tissues. The mouse aorta was cut into 1.5-mm rings and suspended between two wire stirrups (150 μ m) in a myograph (Multi myograph system DMT-620) in 10 ml Krebs-ringer (95% O2-5%CO2, pH7.4, 37 °C). One stirrup was connected to a three-dimensional micromanipulator, and the other to a force transducer. The rings were passively stretched at 10-minutes intervals in increments of 100 mg to reach optimal tone (600 mg). After the arterial rings had been stretched to their optimal resting tone, the contractile response to 100 mM KCl was determined. For constriction assay, phenylephrine (PE) was applied at different concentrations (10⁻⁹-10⁻⁵)

and cumulative responses were recorded. For NO-dependent relaxation responses, vessels were pre-constricted with PE (10^{-6}) for 15 minutes and acetylcholine (Ach, $10^{-9}-10^{-5}$) was then added to determine relaxation activities. To further confirm the vasorelaxation activity in a NO-dependent manner, the inhibitor of guanylate cyclase (1H-[1,2,4]oxadizolo[4,3-a]quinoxalin-1-one, ODQ, 1 μ M) was added at the end of experiments.

8. Determination of thiobarbituric acid-reactive substances (TBARS)

Plasma samples were mixed with trichloroacetic acid (20%) and the precipitate was dispersed in H_2SO_4 (0.05 M). TBA (0.2% in 2 M sodium sulfate) was added and heated for 30 min in boiling water bath. TBARS adducts were extracted by *n*-butanol and absorbance was measured at 532 nm²². Malondialdehyde (MDA) was used as a standard.

9. Statistics

All data are represented as mean \pm S.D. of at least four independent experiments. An unpaired Student's *t*-test or 1–way ANOVA was used to assess significant differences. A value of p<0.05 was accepted as significant.

III. RESULTS

1. Effect of KRGE on arginase activity in aortas of aged mice

With the previous report that KRGE inhibits arginase activity ¹⁸, mice were orally administered KRGE at 10 or 20 mg/mouse/day for 4 weeks. At first, we measured arginase activity in isolated aortic vessels. Arginase activity in aorta of aged mice was significantly increased (* vs. young control, 140.0 ± 13.5 vs. $100\pm6.9\%$, p<0.01) that was blocked by KRGE (** vs. aged, 117.1 ± 10.2 (10 mg/mouse/day) and 110.0 ± 9.2 (20 mg/mouse/day) vs. $140.0\pm13.5\%$, p<0.05).

2. KRGE enhances NO production and decreases ROS generation in endothelium of aged mice

Given that arginase competes with nitric oxide synthase (NOS) for the common substrate, L-arginine, and negatively regulates NO production, we measured whether KRGE–dependent inhibition of arginase activity was associated with increase in NO production. As demonstrated in Fig. 2A, NO production was significantly attenuated in aged mice (* vs. young, 0.58 ± 0.16 vs. 0.90 ± 0.06 change of DAF fluorescence/second, p<0.01). It was restored in groups of KRGE administration (** vs. aged, 0.87 ± 0.11 (10 mg/mouse/day) or 0.91 ± 0.14 (20 mg/mouse/day) vs. 0.58 ± 0.16 change of DAF fluorescence/second, fluorescence/second, p<0.01).

The effect of KRGE on NO production was also observed in young mice (# vs. young, 1.13 ± 0.09 vs. 0.90 ± 0.06 change of DAF fluorescence/second, p<0.01). On the other hand, ROS generation in aged mice was markedly increased (* vs. young, 0.52 ± 0.13 vs. 0.31 ± 0.04 change of DHE fluorescence/second, p<0.01). This was significantly inhibited with KRGE administration (** vs. aged, 0.37 ± 0.13 or 0.28 ± 0.06 change of DHE fluorescence/second, p<0.01). In young mice, KRGE also attenuated ROS generation (# vs. young, 0.24 ± 0.02 vs. 0.31 ± 0.04 change of DHE fluorescence/second, p<0.01). Interestingly, treatment of NOS inhibitor, L-NAME, reduced ROS generation in aged

mice (## vs. aged, 0.29±0.11 vs. 0.52±0.13 change of DHE fluorescence/second, p<0.01).

3. KRGE administration enhanced eNOS coupling

Based on the data of NO production and ROS generation, we next assessed proteins expression to find underlying mechanism. Interestingly, eNOS protein expression in aged mice was significantly increased (young vs. aged, 1.0 ± 0.03 vs. 1.23 ± 0.04 , p<0.01) that was not affected by KRGE administration. On the other hand, arginase II protein expression was not changed in both groups (Fig. 3A). This result is not consistent with the above observation that NO production was decreased and ROS generation was increased in aged mice. Therefore, we tested whether eNOS coupling was reduced in aorta of aged mice. As shown in Fig. 3B, eNOS coupling was attenuated in aged mice, which was restored with KRGE administration (Fig. 3C). These results imply that endothelial function is impaired in aged mice through attenuated eNOS coupling despite increased eNOS expression and KRGE administration can restore augmented eNOS uncoupling in aorta of aged mice.

4. NO-dependent relaxation of aortic vessels was augmented with KRGE administration

With the above data that KGWE administration induced eNOS coupling and NO production, we measured vascular tension to determine whether increased NO production by KGWE contributes to vasorelaxation in aged mice. Mouse aorta were preconstricted with PE (10^{-6} M), and dose-response curves to endothelium-dependent vasodilator acetylcholine (Ach) were constructed. The vasorelaxation responses in aged mice were significantly attenuated compared with those from young (Fig. 4A, *, young vs. aged, 97.16±4.08 vs. 73.30±2.50% (E_{max}), p<0.01; 7.23±0.13 M vs. 6.81±0.09 M (-logEC₅₀), p<0.01). KRGE administration to aged mice enhanced vasorelaxation responses Next, vasoconstrictor responses to the agonist phenylephrine (PE) were measured. As shown in Fig. 4B, the responses to PE were markedly attenuated in aorta from aged mice (*, young vs. aged, 59.92±1.83% vs. 20.93±1.68% (E_{max}), p<0.01; 8.21±0.08 M vs. 7.72±0.20 M (-

logEC₅₀), p<0.01). However, KRGE administration to aged mice enhanced vasoconstrictor responses in E_{max} (Fig. 4B, **, aged vs. aged+KRGE (10 and 20 mg/kg/day, respectively), 20.93±1.68% vs. 40.06±3.32% and 55.84±1.54% (E_{max}), p<0.01), but not in -logEC₅₀ (**, aged vs. aged+KRGE (10 and 20 mg/kg/day, respectively), 7.72±0.20 M vs. 7.56±0.19 M and 7.83±0.07 M, not significance). On the other hand, the responses to the endothelium-independent vasodilator sodium nitroprusside (SNP) were not significantly changed in any group (Fig. 4C)

5. KRGE prevents peroxynitrite formation in aged mice

Given that enhanced oxidative species such as peroxynitrite formation increase arginase activity ²³ and enhanced peroxynitrite formation is associated with vascular aging ²⁴, we performed TBARS assay to measure lipid peroxidation. In plasma of aged mice, lipid peroxidation was significantly increased that was reduced to those of young by administration of KRGE (Fig. 5, *, young vs. aged, 0.23 ± 0.08 vs. 0.52 ± 0.13 µM MDA/ml plasma, p<0.01; #, aged vs. aged+20 mg/mouse/day, 0.52 ± 0.13 vs. 0.29 ± 0.09 µM MDA/ml plasma, p<0.01).

Figure 1



Fig. 1. Increased arginase activity in aged mice aorta was inhibited by administration of KRGE. Oral administration of KRGE for 4 weeks resulted in decrease in arginase activity. * vs. Young, p<0.01; ** vs. Aged, p<0.01.





Fig. 2. KRGE restored impaired endothelial function in aged mice. Isolated aortic segments were incubated with DAF-AM (5 μ M) and fluorescence was measured in real-time (endothelium side up). The slope of DAF fluorescence was determined. (A) KRGE administration with young vessels increased the slope of DAF fluorescence (# vs. young, p<0.01). However, the decreased DAF fluorescence in aged mouse aorta was increased after KRGE ingestion (* vs. young, p<0.01; ** vs. aged, p<0.01; n=4 mice). L-NAME was used as a control. (B) ROS production in aortic endothelium was measured with DHE (5 μ M), and the slope of DHE fluorescence was determined using cumulative data. KRGE intake reciprocally regulated ROS production (# vs. young, p<0.01; * vs. young, p<0.01; ** vs. aged, p<0.01; ## vs. aged, p<0.01; n=4 mice). MnTBAP was used as a control.

Figure 3



Fig. 3. KRGE contributes to improvement of aging-dependent eNOS uncoupling. (A) The expression levels of proteins was not changed by KRGE in aortas of young and aged mice. (B). eNOS dimerization was analyzed by low temperature SDS- PAGE and Western blot analysis and eNOS was uncoupled in aged mice aorta. * vs. young, p<0.05. (C) KRGE induced increase of eNOS coupling in aged mice aorta. * vs. aged, p<0.05.





Fig. 4. KRGE improved impaired vascular reactivity in aged mice. (A) Endothelium-dependent relaxation responses to Ach were impaired in aged aortas (*, young vs. aged, p<0.01). Impaired relaxation was recovered by KRGE administration (**, aged vs. aged+KRGE, p<0.05). (B) Aged aortic vessels had attenuated contractile responses to PE compared to young mice (*, young vs. aged, p<0.01). KGRE restored the PE-mediated pressor responses of aged aortic vessels (**, aged vs. aged+KRGE, p<0.01). (C) Relaxation responses to NO donor (sodium nitroprusside, SNP) are not significantly different in all groups. n=4 mice per each group.

Figure 5



Fig. 5. KRGE administration prevented peroxynitrite formation. Peroxyntrite content in plasma were measured by TBAR assay. Increased peroxynitrite content in aged mice was significantly inhibited with administration of KRGE. young vs. aged, *p<0.01, n=6; aged vs. aged+KRGE, p<0.01, n=6.

IV. DISCUSSION

Endothelial arginase can constrain the activity of eNOS by depleting the critical substrate, L-arginine. In turn, increased arginase activity reduces NO bioavailability and contributes to vascular diseases such as aging, hypertension, and atherosclerosis ^{6, 25-27}. Here, we show that oral administration of KRGE for 4 weeks enhances NO generation, reduces ROS production by inhibiting arginase activity, and induces the vasorelaxation in aortic vessels from aged mice. Furthermore, KRGE reduces the formation of peroxynitrite in plasma.

Ginseng has been shown to have beneficial effects in the treatment of various diseases. In the vasculature, it is well documented that KRGE eliminates superoxide ¹², promotes endothelial cell proliferation ⁸, protects cell death by H₂O₂ stimulation ¹³, and induces heme oxygenase-1 expression ¹⁴. Furthermore, the beneficial effects of ginseng on vascular system may be dependent on the activation of Akt/PI3K signal transduction ¹⁶, inhibition of angiotensin converting enzyme ²⁸, and inhibition of calcium ion influx ²⁹. Here, we demonstrate that KRGE inhibits arginase activity and is associated with decreased peroxynitrite formation through increased NO production and decreased ROS generation in aged vasculature.

Aging is associated with changes in arterial wall structure and function. The most frequent modifications are luminal enlargement, vessel wall thickening due to intimal and medial expansion, elastin depletion and fragmentation, collegen and calcium deposition, glycation of proteins, and impaired vasomotor function associated with endothelial dysfunction ³⁰⁻³². These structural and functional alterations in aging contribute to increased vascular stiffness, which is an independent risk factor for cardiovascular morbidity and mortality ³³⁻³⁵.

Accumulating evidence indicates that arginase contributes to age-associated endothelial dysfunction and arterial stiffening. The detrimental effect of arginase in vascular remodeling is attributable to its ability to stimulate vascular smooth muscle cell and endothelial cell proliferation, and collagen deposition by promoting the synthesis of polyamines and L-proline, respectively.

Vascular ROS production is enhanced in aged blood vessels ^{3, 24, 36, 37}. One of enzymatic systems that contribute to increased ROS production in pathophysiological states may be eNOS. Although eNOS normally produces vasoprotectant molecule NO, it can also produce O₂⁻⁻ in the absence of either L-arginine or BH₄ because electrons flow from the reductase domain in the heme to molecular oxygen rather than L-arginine in uncoupled eNOS. Actually, we showed that arginase activity and peroxynitrite formation was increased, and eNOS was uncoupled in aged vessels. Furthermore, increased ROS production in the endothelium of aged mice was prevented with preincubation of the eNOS inhibitor L-NAME that is consistent with previous observations ³⁸. Peroxynitrite may stimulate arginase II activation via RhoA-dependent ROCK activation without a change in mRNA and protein levels ²³. One study reported that peroxynitrite can react with the redox-active cysteine (Cys¹⁸) of RhoA, which enhances GDP release from RhoA and thus modulates their activity ³. Indeed, we found peroxynitrite formation as measured by the thiobarbituric acid reactive substance assay, was significantly increased in plasma from aged mice, which is consistent with previous publication ²⁴.

Increased expression of eNOS may depend on shear stress and hemodynamic forces ^{39, 40} because of the presence of a shear stress-responsive element in the promoter region of the eNOS gene ⁴¹. Consistent with the demonstration, eNOS expression was increased in aorta ⁴² and not changed in artery ⁴³ and decreased in arteriole ². Furthermore, increased expression of eNOS protein in aged aorta may be one of the compensatory mechanisms to counterbalance endothelial dysfunction by increased arginase activity.

In summary, we demonstrate that KGRE, in aged aortic vessels, inhibits increased arginase activity that is associated with enhanced eNOS dimerization and increased NO production. Furthermore, KRGE augments vasorelaxation in NO-dependent manner and attenuates peroxynitrite formation. These finding suggest that KGRE possesses therapeutic potential for cardiovascular diseases associated with aging.

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혈관기능이 약화된 노화생쥐에서 arginase 길항작용을 통한 홍삼의 효과

국내산 홍삼 추출 액은 심혈관계에 도움 되는 효과가 있는 것으로 알려져 있 다. 심혈관계 질환은 사망률과 이환율에 큰 부분을 차지하며 이런 심혈관계 질환의 발생률은 나이, 즉 노화와 밀접한 관계가 있다. 노화된 혈관에서 agmiase의 발현 증가는 혈관 내피세포의 기능 부전을 일으킨다는 연구가 많이 진행 되었다. 본 연구에서는 홍삼 추출 액을 노화된 생쥐에 구강 투여 하였을 때 일산화질소 합성효소(nitric oxide synthase)의 결합을 통하여 arginase의 활성 도를 낮추고, 일산화질소의 생성을 복구하고, 활성산소의 생성을 억제함을 보 이고 있다. 홍삼 추출 액의 구강 투여로 혈관 탄성 분석을 통해서는 나이든 혈관의 아세틸콜린에 의한 혈관이완능력과 페닐레프린에 의한 혈관수축능력이 비약적으로 상향되는 것을 알 수 있었으며, 세포 내에서 peroxynitrite의 생성이 저해되는 것을 발견하였다. 종합해보면 이런 결과들을 통해 홍삼 추출 액이 일산화질소의 신호전달 체계를 통하여 argniase의 활성도를 저하시켜 혈관보호 효과를 나타낸다고 할 수 있다. 따라서 홍삼 추출 액은 노화에 따른 혈관질환 의 치료에 효과적일 수 있다.

Keywords: 인삼, 홍삼추출액, 노화, arginase, 일산화질소, 내피세포 일산화질소 생성제, 혈관이완