

**Induction of interferon-gamma  
secretion by HCV-specific epitopes  
and its clinical implication in  
patients with chronic hepatitis C**

Myung-Eun Lee

Department of Medical Science  
The Graduate School, Yonsei University

**Induction of interferon-gamma  
secretion by HCV-specific epitopes  
and its clinical implication in  
patients with chronic hepatitis C**

Directed by Professor Kwang-Hyub Han

The Master's Thesis  
submitted to the Department of Medical Science  
the Graduate School of Yonsei University  
in partial fulfillment of the requirements for the degree  
of Master of Medical Science

Myung-Eun Lee

December 2006

This certifies that the Master's Thesis  
of Myung-Eun Lee is approved.

-----  
Thesis Supervisor : Kwang-Hyub Han

-----  
[typed name: Thesis Committee Member#1)

-----  
[typed name: Thesis Committee Member#2)

The Graduate School  
Yonsei University

December 2006

## ACKNOWLEDGEMENTS

또 한번의 도약을 꿈꾸며 노력하고 애써온 지난 2년 남짓에 걸친 시간동안의 저의 작은 결실들을 정리하며 여러모로 부족했던 지난날 많은 도움을 주셨던 분들과 이 지면을 빌어 감사의 뜻을 전하고 싶습니다.

‘실험이란 무엇인가’에 대해서가 아니라 ‘연구란 무엇인지’에 대해 늘 스스로 생각하며 물게 만들어 주시고 이에 조언해 주신 한광협 교수님께 가장 먼저 감사의 뜻을 전합니다. 학문을 넘어선 많은 가르침들 너무 감사했습니다. 큰물에 던져져 어디로 나아가야 하는지 알지 못하고 있을 때마다 늘 세심하게 방향을 지시해 주셨던 안상훈 선생님께도 진심으로 감사드리며, 부족한 작은 결실을 큰 결실로 맺을 수 있도록 조언해 주신 박전한 선생님께도 너무나 감사하다는 말씀을 전하고 싶습니다.

본 연구를 수행하는데 있어서의 실험과 연구방향에 대한 제시, 결과해석에 대한 조언으로 끊임없이 도움을 주셨던 목암생명공학 연구소의 황유경 선생님을 비롯한 이현웅 선생님, 아마 선생님들의 도움이 아니었다면 지금의 결과는 만들어지지 않았을 거라는 것을 잘 알고 있습니다. 감사의 뜻 절대 잊지 않을 것입니다.

실험실을 위해 늘 응원의 힘을 보태어 주시며 작은 문제라도 있을까 늘 챙겨 주시고 신경 써 주신 김자경 선생님, 랩미팅 시간마다 안부의 인사를 물어 주시던 김도영 선생님과 김화숙, 이진형, 박준용, 운영준 선생님들께도 작은 감사의 마음을 전합니다.

pipetting 한번 해본 적 없던 제가 실험을 하겠다고 찾아왔을 때 부족함과 철없음도 늘 눈감아 주시며 선생님처럼, 선배님처럼 또는 언니처럼 같이해주신 장혜영 선생님, 실험을 비롯한 모든 것과, SOS 칠 때 마다 굶은일은 도맡아 해결해 주고 도와주셨던 용욱오빠, 동기라는 이름만으로도 힘이 되어주며 많은 것을 대신해주던 용광오빠, 대학원 생활의 추억을 같이 나눈 숙인언니, 그리고 이제 막 같은 배에 올라선 효정언니, 실험실 생활을 가르쳐 주어서 너무 감사했습니다.

또, 한 고비 한 고비 생길 때마다 늘 커피 한잔의 여유를 같이 해준 연정언니, 실험을 하는 사람의 자세가 무엇인지 무엇을 위해 공부해야 하는지를 늘 생각하고 반성하게 만들어 주시던 이인옥 선생님과, 은주언니, 양정민 선생님, 그리고 동문이란 이름으로 힘이 되어준 정원언니와 민주, 옆방 언니들, 임상의학연구소센터 선생님들 잘 배우고 갑니다.

할 수 있다는 칭찬이 늘 먼저이신 김균환 선생님과 실험실의 이름은 달라도 동기 같았던 거흔오빠와 지은언니, 공대방 오빠들 다들 더 나은 뜻으로 열심히 실험하시고 졸업 잘 하시길 바랍니다.

눈앞의 마시멜로를 보고 한 순간 입으로 넣어버리고 싶은 유혹에 흔들려 할 때 마다 친구라는 이름으로 곁에서 응원해 주고 기다려 주던 하나하나 떠오르는 이름들의 친구들, 같은 공부를 하면서 늘 서로의 걱정을 먼저 해주며 가까이에서 늘 보태준 힘 그 힘으로 버티어 왔다는 것을 이제야 말하게 되어 너무 미안하고 고맙습니다.

끝으로, 가장 큰 힘을 가진 사람들, 앞에서 끌어주고 뒤를 받쳐주고 계신 사랑하는 부모님과 하나뿐인 든든한 룸메이트, 알게 모르게 응원해 주시던 많은 친지분들에게도 이 작은 결실을 바치게 되어 기쁩니다.

보잘 것 없이 갈팡질팡 하고, 밀려오는 파도와 물결에 힘없이 뒤집힐 것만 같았던 제 작은 배에 성공을 향한 힘찬 닻을 올리게 되었습니다. 세상에서 가장 아름다운 유혹은 '성공' 이라는 마시멜로의 이야기처럼 성공 이상의 성공을 향한 항해를 다시 한번 시작해 볼까 합니다.

그 항해의 방향키가 되어 줄 이 논문을 여기서 마칩니다.

2006년 11월 26일

이 명 은

## Table of contents

Abstract .....	1
I. Introduction.....	3
II. Materials and Methods.....	7
1. Study subjects.....	7
2. Synthetic HCV-specific epitopes.....	7
3. Detection of intracellular cytokine staining (ICS).....	8
III. Results.....	9
1. The clinical characteristics of subjects.....	9
2. HCV-specific epitopes - the evidence base.....	10
3. Production of IFN- $\gamma$ by CD8+ T cells.....	11
4. ICS assay analysis of Chronic Hepatitis C (CHC) patients and healthy control samples to HCV-specific epitopes.....	12
5. Frequency of HCV-specific CD8+ T cells in the peripheral blood of sustained virological responders (SVR) and non responders (NR).....	13
6. Pattern of CD8+ T cells responses to HCV-specific epitopes.....	18
IV. Discussion.....	19
V. Conclusion.....	22
References.....	23
Abstract (in Korean).....	29

## LIST OF FIGURES

Figure 1. Detection of IFN- $\gamma$ producing cells in CD8+ T cells from HCV-infected patients.....	11
Figure 2. Epitope-specific average response by ICS assay.....	12
Figure 3. Comparison of IFN- $\gamma$ production in response to HCV-specific epitopes (a) C36, (b) C132, (c) NS4 1789, (d) NS5B 2510, (e) NS5B 2588, (f) C41, (g) C110 among patient group (SVR vs. NR).....	17

## LIST OF TABLES

Table 1. The clinical characteristics of subjects.....	9
Table 2. Synthetic HCV-specific epitopes sequence.....	10
Table 3. Pattern of CD8+ T cells responses to HCV-specific epitopes.....	18

Abstract

Induction of interferon-gamma secretion by  
HCV-specific epitopes and its clinical implication in  
patients with chronic hepatitis C

Myung-Eun Lee

*Department of Medical Science*  
*The Graduate School, Yonsei University*

(Directed by Professor Kwang-Hyub Han)

As interferon/ribavirin-based standard therapy is effective in only about half of HCV patients, there remain important needs for alternatives, including vaccines. Several of the cytokines (IFN- $\gamma$ ) that regulate of immune responses and play an important role in the control of virus-associated disease, including HCV. Therefore, differences in the productivity of IFN- $\gamma$  in patients with HCV infection may account for the outcome of interferon monotherapy.

Thus, we investigated HCV specific IFN- $\gamma$  productivity by HCV-specific

epitopes in patients with sustained virological responders (SVR) and non responders (NR) to interferon therapy.

A total of 40 chronic hepatitis C (CHC) patients were enrolled: 21 SVR patients and 19 NR patients. Peripheral blood mononuclear cells (PBMCs) were collected from heparinized blood of 40 CHC patients by ficoll gradient centrifugation. After in vitro culture for 5 days with each HCV-specific peptide and rhIL-15, the IFN- $\gamma$  productivity were measured by enzyme- $\gamma$  linked immunospot (ELISPOT) analysis and flow cytometry. HLA typing and virus genotyping were performed by Micro SSP HLA class I DNA typing and Radioimmunoprecipitation (RIPA) assay, respectively.

Most of CHC patients had various HLA supertypes (HLA-A2 (27.5%), HLA-A3 (42.5%), HLA-B7 (30%)). The median age was 49.8 years (M:F=2.4:1). Our results showed high IFN- $\gamma$  productivity percentage that IFN- $\gamma$  produced by CD8<sup>+</sup> T cells against HCV-specific epitopes in some of patients with NR when compared with SVR group.

In conclusion, HCV-specific epitopes might be used as a new therapeutics to overcome suppressed cellular immune status in CHC patients with NR for interferon /ribavirin-based standard therapy

---

Key words : Hepatitis C Virus (HCV), HCV-specific epitopes, IFN- $\gamma$  productivity, Sustained Virological Responders (SVR), Non Responders (NR), Interferon.

# Induction of interferon-gamma secretion by HCV-specific epitopes and its clinical implication in patients with chronic hepatitis C

Myung-Eun Lee

*Department of Medical Science  
The Graduate School, Yonsei University*

(Directed by Professor Kwang-Hyub Han)

## **I. INTRODUCTION**

Since its discovery in 1989, hepatitis C virus (HCV) has been recognised as a major public health problem. According to the World Health Organization (WHO) approximately 170 million people worldwide suffer with chronic HCV infection <sup>1</sup>. HCV is a positive-stranded enveloped RNA virus belonging to the family of *Flaviviridae*. Its 10 kilobase genome of approximately 9,500 nucleotides that encodes a polyprotein of approximately 3,000 amino acids. That contains a single open-reading frame giving rise to a polyprotein which is posttranslationally processed into structural (Core, E1, E2/NS1) and non-structural proteins (NS2, NS3, NS4, and NS5) <sup>2,3</sup>. Analysis

of the HCV genome has demonstrated extremely high heterogeneity in both structural and nonstructural coding regions and has identified at least six different genotypes that have generally been divided into several subtypes. Differences in genotypes are significant and can be as much as 40% at the nucleotide level. Besides being restricted to different geographical areas, each genotype has its own unique pattern of disease progression and response to therapy <sup>4</sup>.

Currently there is no vaccine or small molecule therapy for this disease, which can lead to liver failure and cancer. The majority of those infected fail to clear the virus and 20% of persistently infected individuals will develop liver cirrhosis or hepatocellular carcinoma occurs in up to 2.5% <sup>5</sup>. Our current peginterferon (pegasys)/ribavirin combination therapies are effective in only a fraction of the patients and are plagued with adverse effects. Furthermore, the most effective treatment is pegylated interferon (IFN)-plus ribavirin, which has morbid side effects, variable cure rates, and high costs <sup>6</sup>. Thus, new treatment regimens are needed that are more efficacious and better tolerated by all patients.

Investigators have taken several different approaches to address this pressing medical need. Major research efforts have focused on the identification of agents that inhibit specific steps in the life cycle of the virus. The other thing that research modulate the host immune response are being investigated for their ability. Actually, There are investigational compounds <sup>7</sup>.

Also, rapidly emerging viral diseases, such as the hepatitis C virus (HCV), oblige the development of procedures for the fast development of vaccines.

Traditional vaccine developed efficient methods for epitope identification and vaccine design. This analysis is to predict peptides capable of binding to

specific major histocompatibility complex (MHC) molecules. In humans, MHC molecules are called human leukocyte antigen (HLA) and the main anchor residues of HLA Class I molecules occur at positions 2 and at the C-terminus of the peptides, which are usually 8 to 11 amino acid residues in length. T cell receptor (TCR) on the T cell surface recognizes the peptide presented on the self MHC molecule. TCR of CTLs recognizes only linear determinants of epitope peptides defined predominantly by primary amino acid sequences that assume extended conformation within the peptide-binding clefts of class I molecules. Previous studies containing a limited number of peptides will not be applicable to a large part of the human population, since most T cell epitopes are specific for single HLA allele<sup>8,9,10</sup>. Thus, our study used of HCV-specific epitopes capable of binding alleles representing as few as three HLA alleles, HLA-A2, HLA-A3, and HLA-B7 and these synthesized base on the referances<sup>11,12</sup>.

Primary HCV infection causes immune evasion of broad and multispecific CD4+ and CD8+ T cell responses. A variety of postulated mechanism are summarized in *David et al*<sup>13</sup>. HCV antigen-exposure to immune system in individuals who develop persistent infections is usually weak or absent when compared with spontaneously resolving infections<sup>14,15,16,17</sup>.

When exogenous antigen entered into cell as peptides, it could be also presented on MHC class I and recognized by CD8+ T cells. In particular, cytotoxic T lymphocytes (CTLs) paly an important role in the control of virus-associated diseases. Several groups have analyzed HCV-specific CTLs responses to identify CTLs-based epitopes within viral antigens<sup>18-23</sup>. It has been reported that stronger, broader and more sustained Th1/ Tc1 (IFN- $\gamma$ ) responses are associated with resolving infection. Although chronically infected patients also show some IFN- $\gamma$  responses, these tend to be weaker and directed against less epitopes. Thus, IFN- $\gamma$  producing CD8+ T cells may play an important role in the control of HCV antigen and correlation between IFN- $\gamma$  levels and T cell function in CHC<sup>24</sup>.

Also, chronically infected patients identified consistent differences hepatic gene expression in patients who SVR and NR with interferon therapy<sup>25</sup>.

Thus, we have investigated the relationship between IFN- $\gamma$  productivity differences by HCV-specific epitopes and its clinical implication in patients with CHC according to the differences outcome of interferon therapy.

## **II. MATERIALS AND METHODS**

### **1. Study subjects**

A total of 40 Korean subjects having either present or past evidence of HCV infection were enrolled from the patients clinic of the liver unit or from Yonsei University Medical Center between May 2004 and December 2005.

Diagnosis of CHC was based on the following criteria: (1) exposure to HCV-infected blood, (2) detection of serum HCV RNA positive at least 6 months, and (3) elevation of serum alanine aminotransferase (ALT) levels at least 6-fold above the upper limit of normal value (22U/l), and absence of markers for other viral hepatitis and autoimmune hepatitis. Patients treated interferon/ribavirin-based standard therapy for 6 month were followed-up after the end of therapy. Subjects were classified according to the response to interferon into two groups as follows, "Sustained virological responders (SVR) group" contained 21 subjects, who have HCV-RNA negative for at least 6 months followed-up after the end of therapy, "Non responders (NR) group" contained 19 subjects who didn't showed sustained virological remission during IFN treatment.

HLA typing and virus genotyping were performed by Micro SSP HLA class I DNA typing and RIPA assay, respectively.

### **2. Synthetic HCV-specific epitopes**

HCV-specific epitopes were synthesized and purified by a solid-phase and pin-method using the Fmoc-based protocol. Synthetic peptides at 10 mg/ml were dissolved in dimethyl sulfoxide (DMSO) and diluted with phosphate buffered saline (PBS) to 1 mg/ml for the binding assay. The

HCV-specific peptides were obtained by Mogam Biotechnology Institute (synthesized by Peptron Inc., Daejeon, Korea), and each was dissolved in buffer according to the chemical properties of the peptides for *in vitro* stimulation of T cells. The peptides, were resuspended in distilled water and sonicated for homogenization. Each solution of peptide was adjusted to a final concentration of 1 mg/ml and neutral pH. The purity and integrity of the synthesized epitopes were more than 95% as confirmed by HPLC.

### **3. Detection of intracellular cytokine staining (ICS)**

Epitope-specific IFN- $\gamma$  productivity were measured by IFN- $\gamma$  secreting cells among CD8<sup>+</sup> T cells after *in vitro* culture for 5 days with each epitopes and rhIL-15 (rhIL-15, R&D system) by intracellular cytokine staining (ICS) analysis. Briefly, 40 mL of blood was drawn and PBMC isolated by Ficoll-Hypaque density gradient centrifugation. The cells were resuspended in RPMI 1640 plus 10% FCS plus 50u/ml penicillin, 50 $\mu$ g/ml streptomycin and 2mM L-glutamine (R10) to a concentration of  $5 \times 10^5$  PBMC/well (96-well plates). Separate wells were set up with  $5 \times 10^5$  PBMC/well to which was added HCV-specific epitopes at a final concentration of 20 $\mu$ g/ml and rhIL-15 at a concentration of 10 $\mu$ g/ml. At the end of the incubation, the cells were washed twice and stained for 30 min at 4°C with fluorescence isothiocyanate (FITC)-conjugated anti-mouse CD8 in FACS buffer [1% FBS, Gibco].The permeabilized cells were fixed with cytofix/cytoperm (BD Pharmingen), and washed with Perm/Wash buffer (BD Pharmingen). Cells were stained with R-pycoerythrin (R-PE)-conjugated anti-mouse IFN- $\gamma$  (BD Pharmingen) for 30 min at 4°C, and acquired with a FACSCalibur (Becton Dnckinson) and then analysed using CellQuest software (Becton Dnckinson).

### III. RESULTS

#### 1. The clinical characteristics of subjects

In a total of 40 patients with CHC were grouped into 21 patients of SVR and 19 patients of NR according to the interferon response (Table 1). In 21 SVR patients, median age was 52.4 years (range 36-63 years). In 19 NR patients, median age of subjects was 47.1 years (range 39-62 years). There is no considerable clinical characteristics difference between of the two groups.

Table 1. The clinical characteristics of subjects

Characteristics (Number of patients)	Sustained virological responders (SVR) (N=21)	Non responders (NR) (n=19)
Gender (M:F ratio)	1.8 : 1	2.2 : 1
Age (years)	52.4 (36-63)	47.1 (39-62)
HCV genotype (1b:Non-1b ratio)	1 : 1.29	8 : 1
HLA supertype		
A2	5	6
A3	10	7
B7	6	6

## 2. HCV-specific epitopes - the evidence base

Synthetic peptides with the length of 9-10 amino acids derived from HCV core, non-structural NS4, NS5B regions (Table 2). These peptides have been recognized by human PBMC's using cytolytic assays, and/or ELISPOT assays/ICS assays (IFN- $\gamma$ ).

Table 2. Synthetic HCV-specific epitopes sequence

Supertype	Peptides	Position	Protein	Sequence	Reference
A2	C36	36-44	core	LLPPRGPRL	[11]
	C132	132-140	core	DLMGYIPLV	[26]
	NS4-1789	1789-1797	NS4	SLMAFTA AV	[27]
A3	NS5B-2510	2510-2518	NS5B	SLTPPHSAK	[28]
	NS5B-2588	2588-2596	NS5B	RVCEKMALY	[39]
B7	C41	41-49	core	GPRLGVRAT	[30]
	C110	110-118	core	DPRRRSRNL	[12]

### 3. Production of IFN- $\gamma$ by CD8+ T cells

Antigen-experienced CD8+ T cells upregulated IFN- $\gamma$  production after encountering cells displaying their cognate HCV-specific epitopes ligand in association with a MHC complex class I molecule. The data is representation of intracellular cytokine staining (ICS) (Figure 1).

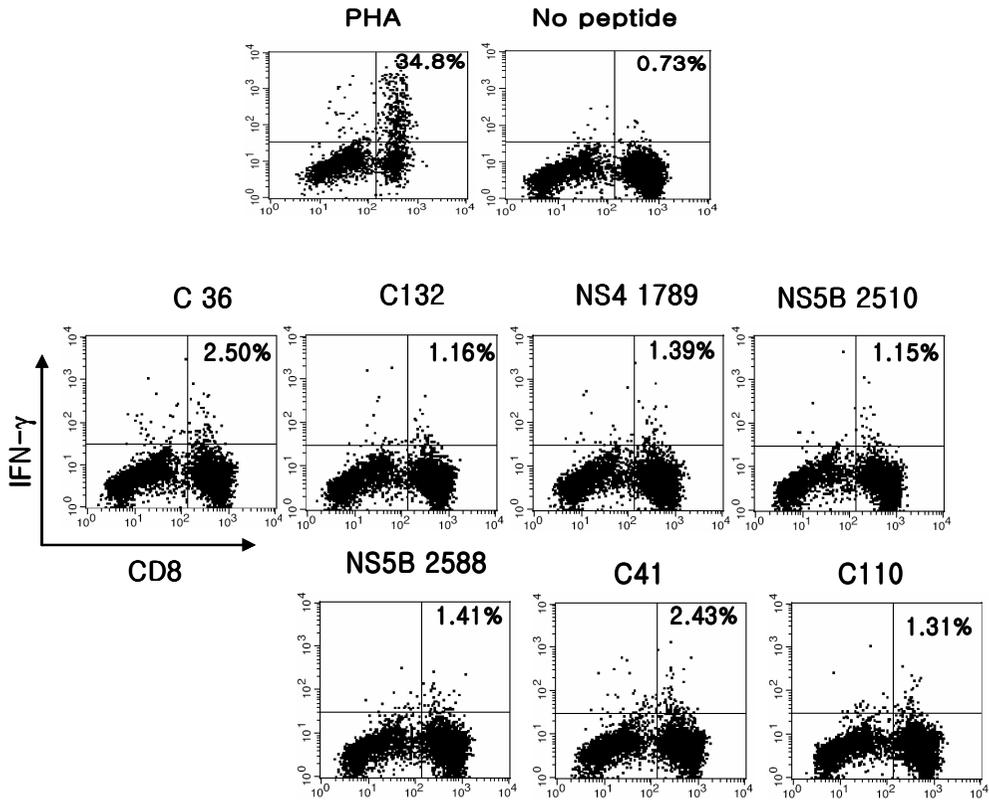
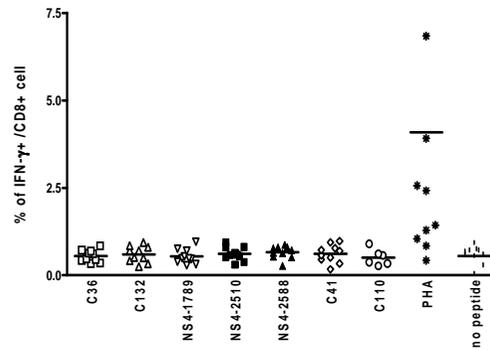


Figure 1. Detection of IFN- $\gamma$  producing cells in CD8+ T cells from HCV-infected patients (PBMCs). Single-cell suspensions were prepared. Cells were stained with anti-CD8-FITC and anti-IFN- $\gamma$ -PE. The percentage of CD8<sup>+</sup>IFN- $\gamma$ <sup>+</sup> cells to compared with control.

#### 4. ICS assay analysis of CHC patients and control samples to HCV-specific epitopes

##### A. Control samples



##### B. CHC patients

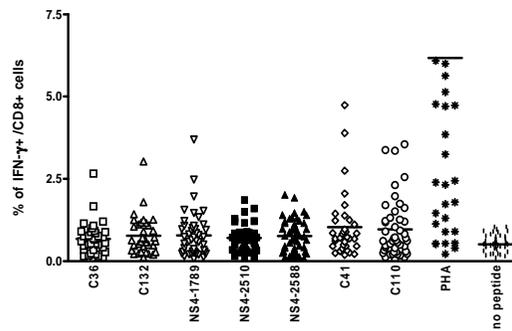
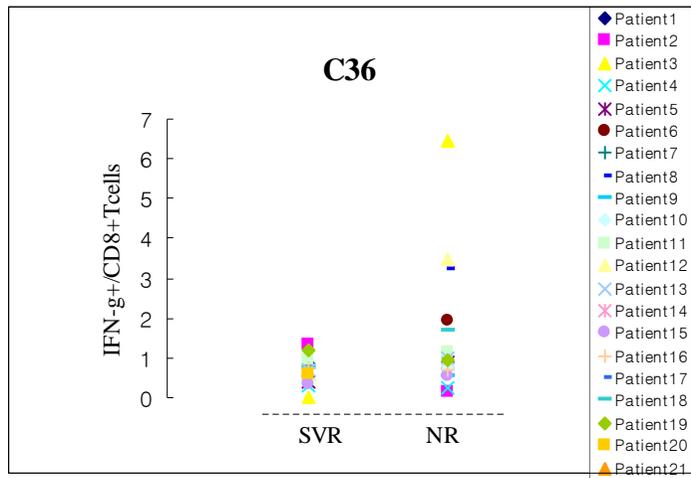


Figure 2. Epitope-specific average response by ICS assay. The IFN- $\gamma$  productivity percentage of healthy control responding to HCV-specific epitopes are shown (A), and CHC patients responding to HCV-specific epitopes are shown (B). A significant cut-off value percentage was more than 1% between (A) and (B). note. control samples : healthy persons.

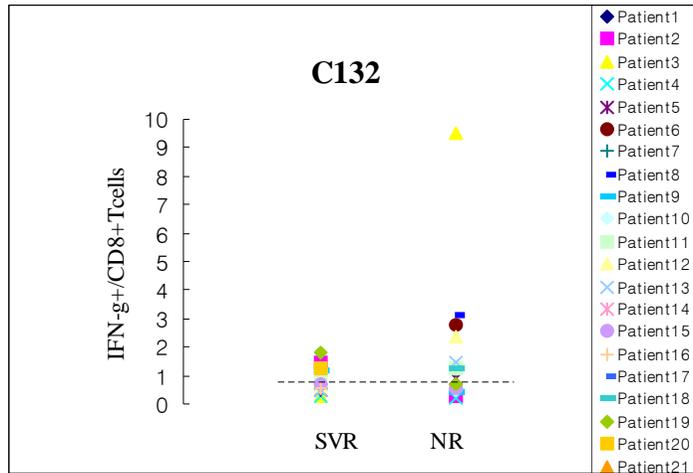
## 5. Frequency of HCV-specific CD8+ T cells in the peripheral blood of SVR and NR patients.

Expression of IFN- $\gamma$  producing cells in PBMCs from SVR and NR patients by various HCV-specific epitopes (C36, C132, NS4 1789, NS5B 2510, NS5B 2588, C41, C110). Comparison of IFN- $\gamma$  expression among SVR (n=21) and NR (n=19) (Figure 3). IFN- $\gamma$  productivity was rated positive if peptide-specific IFN- $\gamma$  expression was > 1%.

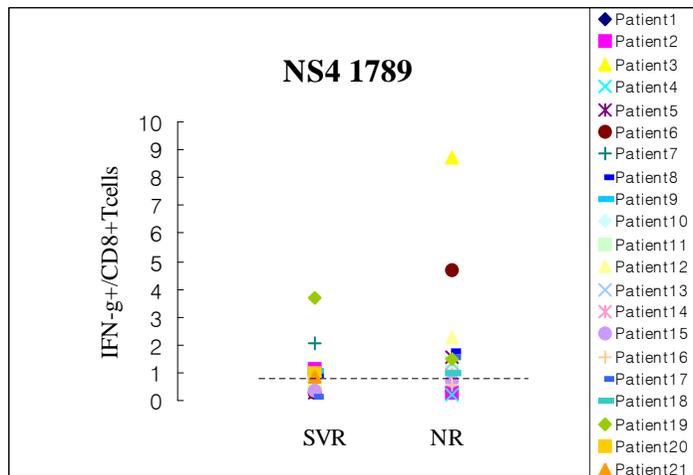
### (a) HCV-specific epitope C36



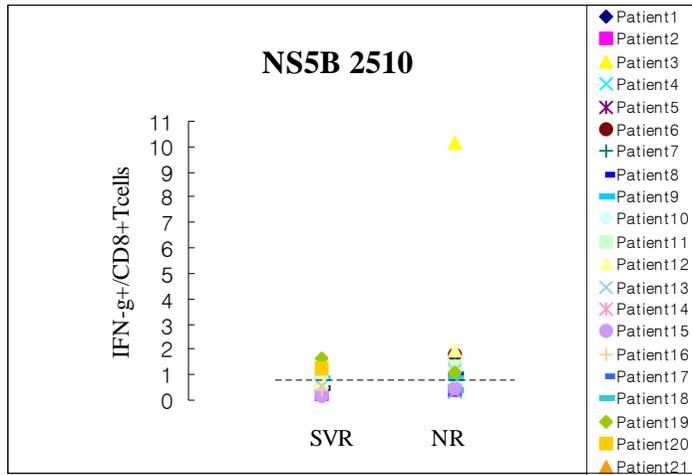
**(b) HCV-specific epitope C132**



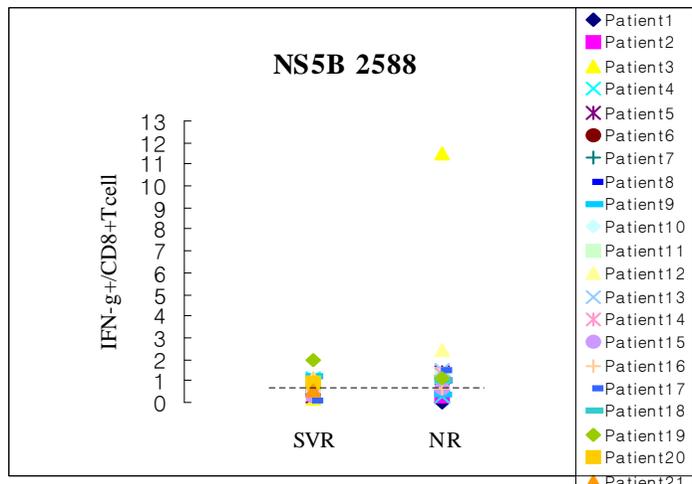
**(c) HCV-specific epitope NS4 1789**



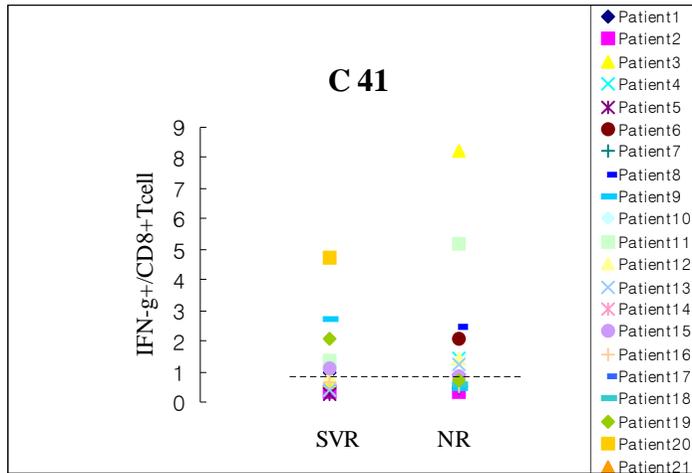
**(d) HCV-specific epitope NS5B 2510**



**(e) HCV-specific epitope NS5B 2588**



**(f) HCV-specific epitope C41**



**(g) HCV-specific epitope C110**

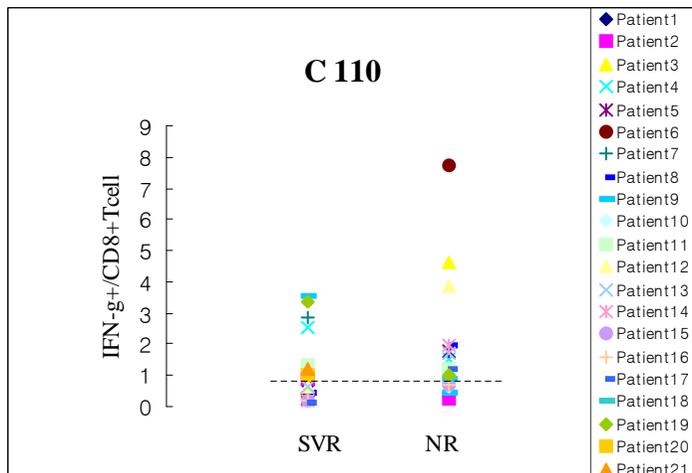


Figure 3. Comparison of IFN- $\gamma$  production in response to HCV-specific epitopes (a) C36, (b) C132, (c) NS4 1789, (d) NS5B 2510, (e) NS5B 2588, (f) C41, (g) C110 among patient group (SVR *vs* NR). Each dot represents the individual SVR patients (n=21) and NR patients (n=19).

## 6. Pattern of CD8+ T cells responses to HCV-specific epitopes

The positive percentage of HCV-antigen-specific IFN- $\gamma$  production assays is shown for patients with SVR and NR in relation to the protein location of the HCV-specific epitopes (Table 3).

Table 3. Pattern of CD8+ T cells responses to HCV-specific epitopes

	HCV-specific epitopes						
	C36	C132	NS41789	NS5B2510	NS5B2588	C41	C110
<b>SVR</b>							
RN/TN (%)	3/15 (20)	6/15 (40)	6/21 <b>(28.6)</b>	4/17 (23.5)	6/19 (31.6)	7/15 (46.7)	8/21 (38.1)
<b>NR</b>							
RN/TN (%)	4/15 (25)	8/16 (50)	11/19 <b>(57.9)</b>	7/16 (43.8)	8/19 (42.1)	6/16 (37.5)	9/19 (47.4)

Note. Percentage was calculated by the ratio of patient number showing the response to each epitope to total patients number (RN / TN).

## IV. DISCUSSION

The outcome of many viral infections is determined by the nature of the antiviral immune response. However it is well known that the cellular immune responses to be too weak to eliminate HCV and a functional suppression has been reported in chronic HCV infection<sup>31,32,33,34,35</sup>.

We identified that IFN- $\gamma$  productivity to HCV-specific epitopes that distinguish patients with SVR and NR after the end of IFN therapy. IFN- $\gamma$  producing CD8+ T cells was confirmed by ICS assays in figure 1 (Mogam Biotechnology Institute).

These data showed the most of patients recruited strong IFN- $\gamma$  productivity after *in vitro culture* with these epitope peptides and IL-15, while freshly collected bloods from 40 CHC patients secreted little amount of IFN- $\gamma$  to selected epitopes stimulation (data not shown). These results thought about antigens are provided by professional antigen presenting cell expressing co-stimulatory molecules or producing cytokines of IL-15. Actually, recent studies have shown that IL-15 can positively regulate the maturation processes as well as functions of DCs to generate efficient pathogen or tumor-specific CTLs responses<sup>36,37,38</sup>. IL-15 also mediates pleiotropic actions by driving the proliferation and maintenance of Ag-specific memory T lymphocytes as well as the production of TH1-driven cytokines<sup>39,40,41</sup>.

We have found that the differences of IFN- $\gamma$  productivity between in some of patients with NR and in some of patients with SVR. The IFN- $\gamma$  productivity showed higher frequency in NR group than in SVR group. Also, some patients with NR have that more recognize NS4 1789 peptide than SVR, relation to the protein location of the HCV antigen (28.6% vs 57.8%).

Interestingly, however, these results indicated that strong IFN- $\gamma$  induced

by CD8+ T cells to HLA-restricted A2, A3, B7 peptides in patients with NR. These results are consistent with recent studies showing higher frequency of IFN- $\gamma$  producing peripheral CD4+ and CD8+ T cells in HCV RNA-positive patients than in HCV RNA-negative patients or healthy controls<sup>42</sup>. Also, *Fareed Rahman and Barbara Rehermann et al*<sup>43</sup> compared to patients sustained treatment response and patient with viral breakthrough. HCV-specific T cells responses showed some fluctuation with significant peaks of IFN- $\gamma$  producing T cells according to ALT levels and HCV-RNA. These results suggest that HCV-specific T cells responses may have been stimulated and maintained by low levels of remaining antigen and that HCV may not have been completely cleared.

However, all of the patients didn't response to HCV-specific epitopes. One of the candidate answer of this question is a diversity host factors. For examples, there are HCV genotype, gender, sex, ethnic, and age. Actually, specific HLA alleles have been reported to be associated with response to therapy in chronic HCV infection<sup>44</sup>. In addition, the role of single nucleotide polymorphisms (SNPs) in some specific genes related to disease and treatment response, in considering the host genetic factors, has become increasingly important in a variety of diseases.

It is possible that other host protein with different functions may also be associated, through different mechanisms, with the IFN responsiveness of the HCV-infected patients. The MxA protein is one of such candidats, because it is a protein that is induced by IFN and influences the IFN-induced antiviral activities of host cells against several other viruses<sup>45,46,47</sup>. Also, a polymorphic site identified in the promoter region of MxA was involved in a region which shows a considerable resemblance to the IFN-stimulated response element (ISRE)<sup>48</sup>.

The traditional attempts to develop single peptide vaccines, based on a limited number of peptides face two problems : HLA polymorphism and the escape mutant. Thus, single peptide based vaccine containing a limited

number of peptides will not be applicable to a large part of the human population, since most T cell epitopes are specific for a single HLA allele <sup>49</sup>. However, the use of supertype-restricted multi-peptides, those capable of binding with significant affinity to multiple related HLA alleles. Thus, we used of HCV-specific multi-peptides capable of binding alleles representing as few as three HLA alleles, HLA-A2, HLA-A3, and HLA-B7, results in predicted recognition in nearly 90% of the global population, regardless of ethnicity.

Actually, we performed HLA typing assay by Micro SSP HLA class I DNA typing respectively from patients. The most of patients have a various HLA supertype (HLA-A2 (27.5%), HLA-A3 (42.5%), HLA-B7 (30%)) showed that in Table 1. But, we didn't check up association with between HLA type and IFN- $\gamma$  productivity. Thus, whether HLA type might be related to the IFN- $\gamma$  productivity and differences outcome of interferon therapy or not, further study may be needed.

Our experiment is detection IFN- $\gamma$  producing CD8+ T cells *in vitro culture*, the IFN- $\gamma$  productivity showed higher frequency in NR group than in SVR group. Taken together, these HCV-specific multi-peptides might contribute to overcome suppressed cellular immune status in CHC patients with NR for interferon /ribavirin-based standard therapy. It also gives the possibility that multi-peptide-based DNA vaccine vectors could be design. But, our experiment is just detection of patients with the end of treated interferon/ribavirin-based standard therapy. Therefore, further study may be needed a before starting drug treatment and during the therapy. In addition, further detailed study is needed to evaluate the regulatory mechanism, including specific CD4+ T cells and regulatory T cells.

## **V. CONCLUSION**

We have found that IFN- $\gamma$  was produced by CD8<sup>+</sup> T lymphocytes against our HCV-specific epitopes in patients with SVR and NR. The IFN- $\gamma$  productivity showed higher frequency in NR group than in SVR group to each HCV-specific epitopes (C36, C132, NS4 1789, NS5B 2510, NS5B 2588, C41, C110).

## VI. REFERENCES

1. WHO. Global surveillance and control of hepatitis C. Report of a WHO Consultation organized in collaboration with the Viral Hepatitis Prevention Board, Antwerp, Belgium. *J Viral Hepat* 1999;6:35-47.
2. Houghton M, Weiner A, Han J, Kuo G, Choo QL. Molecular biology of the hepatitis C viruses: implications for diagnosis, development and control of viral disease. *Hepatology* 1991;14(2):381-8.
3. Lohmann V, Koch JO, Bartenschlager R. Processing pathways of the hepatitis C virus proteins. *J Hepatol* 1996;24(2 suppl.):11-9.
4. Sobail A. Qureshi. Medicinal Research Reviews; Hepatitis C virus-Biology, Host Evasion Strategies, and Promising New Therapies on the Horizon.
5. Bowmen, D.G.& Walker, C.M. the origin of quasispecies: Cause or consequence of chronic hepatitis C viral infection. *J. Hepatol* 2005;42, 408-417.
6. McHutchison, J.G., Gordon, S.C., Schiff, E.R., et al. Interferon alfa-2b alone or in combination with ribavirin as initial treatment for chronic hepatitis C. Hepatitis Interventional Therapy Group. *New England Journal of Medicine*, 1998;339(21):1485-1492.
7. Raffaele De Francesco and Giovanni Migliaccio. Challenges and successes in developing new therapies for hepatitis C. *Nature* .Vol 436.18 August 2005.
8. Kast, A. M., Roux, L., Curren, J., Blom, H.J., Voordouw, A.C., Meloea, R.H., Kolakofsky, D., Melief, C.J. Protection against lethal Sendai virus infection by in vivo priming of virus-specific cytotoxic T lymphocytes with a free synthetic peptide. *Proc. Natl. Acad. Sci. U.S.A.* 1991;88:2283-2287.

9. Wang, Y.E., Zhang, C., Berzofsky, J., DeLisi, C. Selecting stable molecular targets for treatment and prevention of AIDS. *Genome Inform.* 2005;16:254-261.
10. Missale, G. et al. Different clinical behaviors of acute hepatitis C virus infection are associated with different vigor of the anti-viral cell-mediated immune response. *J. Clin. invest.* 1996;98:706-714
11. Hanns F., Daniel S., Monika A., Sandra W., Guido, G and Karl-Hermann Meyer zum Buschenfelde. The viral clearance in interferon-treated chronic hepatitis C is associated with increased cytotoxic T cell frequencies. *Journal of Hepatology* 1999;31:407-415.
- 12 S. WARD, G. LAUER, R. ISBA, B., WALKER & P.KLENERMAN. Cellular immune responses against hepatitis C virus: the evidence base 2002. *Clin Exp Immunol* 2002; 128:195-203.
13. David G. Bowen and Christopher M. Walker. Adaptive immune responses in acute and chronic hepatitis C virus infection. *Nature* vol 436 18 August 2005.
14. Diepolder,H.M. et al. Possible mechanism involving T-lymphocyte response to non-structural protein 3 in viral clearance in acute hepatitis C virus infection. *Lancet* 1995;346:1006-1007.
15. Gerlach, J.T. et al. Recurrence of hepatitis C virus after loss of virus-specific CD4+ T-cell response in acute hepatitis C virus. *Gastroenterology* 1999;117:933-941.
16. Thimme, R et al. Determinants of viral clearance and persistence during acute hepatitis C Virus infection. *J. Exp. Med* 2001;194:1395-1406.
17. Cerny, A., McHutchison, J. G., Pasquinelli, C., Brown , M. E., Brothers, M. A., Grabscheid, B., Fowler, P., Houghton, M. and Chisari, F. V. Cytotoxic T lymphocyte response to hepatitis C virus-derived peptides containing the HLA A2.1 binding motif. *J. Clin. Invest.* 1995;95:521-530.

18. Battegay, M., Fikes, J., Di Bisceglie, A. M., Wentworth, P.A., Sette, A., Celis, E et al. Patients with chronic hepatitis C have circulating cytotoxic T cells which recognize hepatitis C virus encoded peptides binding to HLA-A2.1 molecules. *J. Virol.* 1995;69:2462-2470.
19. Shirai M., Arichi, T., Nishioka. M., Nomura, T., Ikeda, K., Kawanishi, K., Engelhard, V. H., Feinstone, S. M. and Berzofsky, J. A., CTL response of HLA-A2.1-transgenic mice specific for hepatitis C viral peptides predict epitopes for CTL of humans carrying HLA-A2.1. *J. Immunol.* 1995; 154:2733-2742.
20. Kurokohchi, K., Akatsuka, T., Pendleton, C. D., Takamizawa, A., Nishioka, M., Battegay, M., Feinstone, S. M. and Berzofsky, J.A., Use of recombination protein to identify a motifnegative human cytotoxic T cell epitope presented by HLA-A2 in the hepatitis C virus NS3 region. *J. Virol.*1996;70:232-240.
21. Wentworth, P.A., Sette. A., Celis, E., Sidney, J., Southwood, S., Crimi, C., Stitely, S., Keogh, E., Wong, N. C., Livingston, B.,Alazard, D. et al. Identification of A2-restricted hepatitis C virus- specific cytotoxic T lymphocyte epitopes from conserved regions of the viral genome. *Int. Immunol.* 1996;8:651-659.
22. Scognamiglio, P., Accapezzato, D., Casciaro, M.A., Cacciani, A., Artini, M., Bruno, G., Chicu, M., sidney, J., Southwood, S., Abrignani, S., Sette, A. and Barnaba, A., Presence of effector CD8+ T cell in hepatitis C virus-exposed healthy seronegative donors. *J. Immunol.*1999;162:6681-6689.
23. Limin, C., Ivan, B., Jordan, F., Jing, S., Laura-Lee, T., Catalina, C., Jenny, H., Aled, M., et al. Hepatic gene expression discriminates responders and nonresponders in treatment of chronic hepatitis C viral infection. *Gastroenterology* 2005;128:1437-1444.
24. Nelson DR.,Marousis CG., Davis GL., et al. The role of hepatitis C

- virus-specific cytotoxic T lymphocytes in chronic hepatitis C. *J Immunol* 1997;158:1473-1481.
25. Moser JM, Altman JD, Lukacher AE. Antiviral CD8<sup>+</sup> T cell responses in neonatal mice: susceptibility to polyoma virus induced tumors is associated with lack of cytotoxic function by viral antigen-specific T cells. *J. Exp Med* 2001;193:595-606.
  26. Alexander J, Del Guercio MF, Fikes JD, Chesnut RW, Chisari FV, Chang KM, Appella E, Sette A. Recognition of a novel naturally processed, A2 restricted, HCV-NS4 epitope triggers IFN-gamma release in absence of detectable cytopathicity. *Hum Immunol* 1998;59:776-82.
  27. Cerny A, McHutchison JG, Pasquinelli C et al. Cytotoxic T lymphocyte response to hepatitis C virus-derived peptides containing the HLA A2.1 binding motif. *J Clin Invest* 1995;95:521-30.
  28. Wong DK, Dudley DD, Afdhal NH et al. Liver-derived CTL in hepatitis C virus infection. breadth and specificity of responses in a cohort of persons with chronic infection. *J Immunol* 1998;160:1479-88.
  29. Koziel MJ, Dudley D, Afdhal N, Grakoui A, Rice CM, Choo QL, Houghton M, Walker BD. HLA class-I-restricted cytotoxic T lymphocytes specific for hepatitis C virus. Identification of multiple epitopes and characterization of patterns of cytokine release. *J Clin Invest* 1995; 96:2311-21.
  30. Gruener NH, Gerlach TJ, Jung MC et al. Association of hepatitis C virus-specific CD8<sup>+</sup> T cells with viral clearance in acute hepatitis C. *J Infect Dis* 2000;181:1528-36.
  31. Welsh RL. Assessing CD8 T cell number and dysfunction in the presence of antigen. *J Exp Med* 2001;193:F19-F22.
  32. Chisari FV. Cytotoxic T cells and viral hepatitis. *J Clin Invest* 1997;99:1472-7.

33. Gruener NH, Lechner F, Jung MC et al. Sustained dysfunction of antiviral CD8<sup>+</sup> T lymphocytes after infection with hepatitis C virus. *J Virol* 2001; 75:5550-8.
34. Wedemeyer H, He XS, Nascimbeni M et al. Impaired effector function of hepatitis C virus-specific CD8<sup>+</sup> T cells in chronic hepatitis C virus infection. *J Immunol* 2002;169:3447-58.
35. Rubinstein, M.P., A. N. Kadima, M. L. Salem, C. L. Nguyen, W.E. Gillanders, and D.J. Cole. Systemic administration of IL-15 augments the antigen-specific primary CD8<sup>+</sup> T cell response following vaccination with peptide- pulsed dendritic cells. *J. Immunol.* 2002;169:4928.
36. Mahamadzadeh, M., F.Berard, G.ESSERT,C.Chalouni,B. Pulendran, J. Davoust, G.Bridges, A.K. Palucka, and J. Bachereau. Interleukin 15 skews monocyte differentiation into dendritic cells with features of Langerhans cells. *J. Exp. Med.* 2001;194:1013.
37. Ohteki, T., K. Suzue, C. Maki, T. Ota, and S. Koyasu. Critical role of IL-15-IL-15R for antigen-presenting cell functions in the innate immune response. *Nat. Immun.* 2001;12:1138.
38. Fehniger, T. A., and M, A. Caligiuri. Interleukin 15: biology and relevance to human disease. *Blood* 2001;97:14.
39. Schluns, K. S., K. Williams, A. Ma, X. X. Zheng, and L. Lefrancois. Requirement for IL-15 in the generation of primary and memory antigen-specific CD8 T cells. *J. Immunol.* 2002;168:4827.
40. Becker, T. C., E. J. Wherry, D. Boone, K. Murali-Krishna, R. Antia, A. Ma, and R. Ahmed. Interleukin 15 is required for proliferative renewal of virus-specific memory CD8 T cells. *J. Exp. Med* 2002;195:1541.
41. Timm, J., Lauer, G. M., Kavanagh, D.G., Sheridan, I., Kim, A.Y., Lucas, M., Pillay, T., Ouchi, K et al. CD8 epitope escape and reversion in acute HCV infection. *J. Exp. Med* 2004;200:1593-1604.

42. Fenyu R., Keisuke H., Yuhki Y., Muneko O., Akira K., Yasuhiro S., Masaaki K., Michiari O., Kiwamu O. Hepatitis C virus infection upregulates expression of the type I infection receptor in human peripheral blood mononuclear cell. *Hepatology Research* 2003;26:15-22.
43. Fareed R., Theo H., Yuji S., Eishiro M., Michelina N., Harvey A., Steven H., Jay H., T.jake Liang., Barbara Reherrmann. Effects of antiviral therapy on the cellular immune response in acute hepatitis C. *Hepatology* ,Vol. 40, No. 1, 2004.
44. Robinson, J., Waller, M. J., Parham, P., Bodmer, J.G., Marsh, S.G.E. IMGT/HLA Database-a sequence database for the human Major Histocompatibility Complex. *Nucleic Acids Res.* 2001;29:210-213.
45. Pavlovic J, Zürcher T, Haller O, Staeheli P: Resistance to influenza virus and vesicular stomatitis virus conferred by expression of human MxA protein. *J Virol* 1990;64:3370-3375.
46. Zhao H, De BP, Das T, Banerjee AK: Inhibition of human parainfluenza virus-3 replication by interferon and human MxA. *Virology* 1996;220:330-338.
47. Landis H, Simon-Jodieke A, Kloti A, Di Paolo C, Schnorr JJ, Schneider-Schaulies S, Hefti HP, Pavlovic J: Human MxA protein confers resistance to Semliki Forest virus and inhibits the amplication in the absence of viral structural protein. *J Virol* 1998;72:1516-1522.
48. Nakade K, Handa H, Nagata K: promoter structure of the MxA gene that confers resistance to influenza virus. *FEBS Lett* 1997;418:315-318.
49. Yu ML, Dai CY, Chen SC et al. Human leukocyte antigen class I and II alleles and response to interferon-alpha treatment, in Taiwanese patients with chronic hepatitis C virus infection. *J Infect Dis* 2003;188: 62-65.

Abstract (In Korean)

만성 C형 간염 환자를 대상으로 **HCV-specific epitopes**을  
이용한 인터페론-감마 유도능과 그것과의 임상학적인  
연관성

<지도교수 한 광 협>

연세대학교 대학원 의과학과

이 명 은

만성 C형 간염 환자의 경우 약 50% 의 경우만이 인터페론/리바비린에 기초한 치료방법에 의해 효과를 나타내기 때문에 백신을 포함해 치료 변화에 대한 요구가 중요시된다. 몇몇의 cytokines (**IFN- $\gamma$** ) 은 면역반응의 조절과 만성 C형 간염 바이러스를 포함하여 바이러스와 관련되어진 질병을 조절하는데 있어서의 중요한 역할을 한다. 그러므로 만성 C형 간염 환자의 서로 다른 **IFN- $\gamma$  productivity** 는 인터페론 치료를 극복하는데 있어 연관되어 있을 것으로 보인다.

따라서, 우리는 인터페론 치료에 의한 sustained virological responders (SVR) 과 non responders (NR) 환자들을 대상으로 HCV-specific epitopes을 이

용하여 그들의 IFN- $\gamma$  productivity 대해 조사하였다.

만성 C형 바이러스 감염환자 40명을 대상으로 21명의 SVR 과 19명의 NR 환자로 구분하였다. 40명의 만성 C형 바이러스 감염환자로부터 그들의 말초단핵핵세포 (PBMC)를 ficoll을 이용한 원심분리기의 층 분리로 분리한 뒤에 각각의 peptide 와 인터루킨 15 (IL-15)을 처리하여 5일 동안 세포배양 한다. IFN- $\gamma$  productivity은 ELISPOT 분석과 flow cytometry 방법에 의해 측정되었다. HLA typing 과 virus genotyping에 대한 측정은 각각 Micro SSP HLA class I DNA typing 과 Radioimmunoprecipitation (RIPA) assay 방법을 수행하였다.

만성 C형 바이러스 감염환자의 대부분은 다양한 HLA supertypes 을 갖고 있는 것으로 조사되었으며(HLA-A2 (27.5%), HLA-A3 (42.5%), HLA-B7 (30%)), 그들의 평균나이는 49.8세로 남녀간의 성 비율은 2.4:1을 보였다 (M:F=2.4:1). 우리의 결과는 HCV-specific epitopes 에 반응하여 CD8+ T cell에서 유도된 인터페론-감마의 IFN- $\gamma$  productivity 가 NR 환자의 경우 SVR 과 비교하였을 때 높게 나타났다.

결론적으로, 이러한 HCV-specific epitopes 이 만성 C형 바이러스에 감염되고 인터페론치료에 대한 반응이 없던 NR 환자의 저하되어 있는 세포성 면역반응을 유도하는 새로운 치료로써 이용될 가능성이 있다는 사실을 확인하였다.

---

핵심되는 말 : C형 감염 바이러스, HCV에 특이적인 epitopes, IFN- $\gamma$  productivity sustained virological responders (SVR) , non responders (NR) 인터페론