

Serum and Cerebrospinal Fluid Neuron-Specific Enolase for Diagnosis of Tuberculous Meningitis

Tae-Jin Song, Young-Chul Choi, Kyung-Yul Lee, and Won-Joo Kim

Department of Neurology, Gangnam Severance Hospital, Yonsei University College of Medicine, Seoul, Korea.

Received: September 26, 2011

Revised: December 6, 2011

Accepted: December 8, 2011

Corresponding author: Dr. Won-Joo Kim,
Department of Neurology,
Gangnam Severance Hospital,
Yonsei University College of Medicine,
211 Eonju-ro, Gangnam-gu,
Seoul 135-720, Korea.
Tel: 82-2-2019-3324, Fax: 82-2-3462-5904
E-mail: kzoo@yuhs.ac

The authors have no financial conflicts of interest.

Purpose: Late diagnosis and treatment lead to high mortality and poor prognosis in tuberculous meningitis (TbM). A rapid and accurate diagnosis is necessary for a good prognosis. Neuron-specific enolase (NSE) has been investigated as a biochemical marker of nervous tissue damage. In the present study, the usefulness of NSE was evaluated, and a cut-off value for the differential diagnosis of TbM was proposed. **Materials and Methods:** Patient charts were reviewed for levels of serum and cerebrospinal fluid (CSF) NSE, obtained from a diagnostic CSF study of samples in age- and gender-matched TbM (n=15), aseptic meningitis (n=28) and control (n=37) patients. **Results:** CSF/serum NSE ratio was higher in the TbM group than those of the control and aseptic groups ($p=0.001$). In binary logistic regression, CSF white blood cell count and CSF/serum NSE ratio were significant factors for diagnosis of TbM. When the cut-off value of the CSF/serum NSE ratio was 1.21, the sensitivity was 86.7% and the specificity was 75.4%. **Conclusion:** The CSF/serum NSE ratio could be a useful parameter for the early diagnosis of TbM. In addition, the authors of the present study suggest a cut-off value of 1.21 for CSF/serum NSE ratio.

Key Words: Neuron-specific enolase, tuberculous meningitis

INTRODUCTION

The diagnosis of tuberculous meningitis (TbM) is difficult because early symptoms and laboratory results from cerebrospinal fluid (CSF) are unspecific to tuberculosis and overlap with those of other chronic diseases of the central nervous system. Late diagnosis can result in a poor prognosis and even death, and a confirmative culture study for TbM requires a long time before a diagnosis can be affirmatively made. Rapid diagnosis and proper treatment is essential for TbM and a lack of confidence in test results can delay the initiation of treatment with anti-tuberculosis medication in TbM patients.^{1,2} In addition, most of the tests developed for the early diagnosis of TbM are not sensitive³ and cannot be used routinely.^{4,5}

Neuron-specific enolase (NSE) is found in relatively large amounts in neurons, peripheral nervous system tissues and neuroendocrine cells, and is, therefore, one of the most often investigated biochemical markers of nervous tissue damage. In-

© Copyright:

Yonsei University College of Medicine 2012

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

creased NSE levels in serum, CSF, or both have been reported in several neurological diseases, including status epilepticus,⁶ Creutzfeldt-Jakob disease,⁷ acute ischemic stroke,⁸ head injury⁹ and hypoxic brain injury.¹⁰ NSE can also act as a marker of neuronal damage, especially in meningitis;¹¹ however, data for TbM is rare. The authors of the present study attempted to confirm the usefulness of NSE and to propose a cut-off value for the differential diagnosis of TbM.

MATERIALS AND METHODS

Patients

The authors retrospectively reviewed charts from January 2000 to June 2008 to determine the levels of serum and CSF NSE obtained before starting anti-Tb or anti-viral treatment. We investigated NSE routinely for evaluating central nervous system involvement. The TbM groups were subdivided into a group of patients in whom *M. tuberculosis* was observed by direct smear with acid fast bacillus staining or in CSF culture (confirmative TbM) or a group of patients with CSF findings of pleocytosis with lymphocytic predominance, decreased CSF (blood glucose ratio less than 0.5), elevated protein, negative gram and India ink stains, no positive bacterial or fungal cultures, and one or more of the following: dramatic responsiveness to anti-tuberculosis medication that subsided neurologic symptoms, i.e. improvement in headache, fever, ocular movement and CSF findings; chest radiography consistent with active pulmonary tuberculosis or confirmed in another organ; typical brain CT or brain MRI findings such as hydrocephalus; edema; or basal meningeal enhancement (presumptive TbM). The age and sex matched control group included tension type headache patients diagnosed with normal neurological examination, brain imaging and CSF results. Lumbar puncture was performed in patients with meningitis-like symptoms with posterior neck pain or neck stiffness, even though tension type headaches were suspected. The aseptic meningitis group included patients who had acute onset of fever, symptoms and signs of meningeal irritation and supportive CSF findings such as a mild increase in protein, normal glucose level and predominantly lymphocytic pleocytosis, no history or clinical findings of intra or extra-cranial tuberculosis, no seizure and alteration of consciousness, no organismal growth in a culture study and conversion to normal CSF upon follow up CSF test at 3 days after the first lumbar puncture. Patients with fungal and bacterial meningitis, stroke, seizure, meningoencephalitis, de-

generative disease, autoimmune disease or traumatic brain damage were excluded because these conditions can increase serum and CSF NSE levels.

Serum NSE collection and quantification

All patient samples of NSE were collected intravenously upon admission. Samples were centrifuged for 30 minutes and stored at -80°C for later analysis. The NSE sample was analyzed using an enzyme immunoassay based on the sandwich technique including the solid-phase monoclonal antibody against NSE (Cobas Core II; Roche Diagnostics, Basel, Switzerland). Hemolytic specimens were excluded because blood cell lysis influences serum NSE level.¹²

Data analysis

Statistical analysis was performed with PASW Statistics software, version 18 (IBM, Chicago, IL, USA). To compare variables among the TbM, aseptic meningitis and controls groups, chi-square, one-way ANOVA and Scheffe's post-hoc analyses were used. A *p*-value of less than 0.05 was considered significant.

To investigate predictive factors for diagnosis of TbM, binary logistic regression was performed, where variables with a *p*-value of less than 0.1 in univariate analysis were entered into the multivariate analysis. To analyze the sensitivity, specificity and cut-off value of CSF/serum NSE ratio, NSE related variables significant in multivariate logistic regression were used to generate receiver operating characteristic (ROC) curves. A two-tailed value of *p*<0.05 was considered significant.

RESULTS

The demographic features and laboratory findings of the patient and control groups are presented in Table 1. There were no significant differences in demographic features among the three groups (Table 1). Fifteen patients were included in the TbM group, 28 in the aseptic meningitis group and 37 in the control group. The TbM group consisted of 3 patients with confirmed TbM via culture, one that was smear-positive and 11 presumptive (two patients showed dramatic responsiveness to anti-tuberculosis medication; one patient's chest radiography was consistent with active pulmonary tuberculosis; eight patients exhibited typical brain imaging findings for TbM). There were no seizures or altered mental states in TbM group and 8 patients showed basal meningeal

Table 1. Demographic Features and Laboratory Findings

	Control (n=37)	Tb meningitis (n=15)	Aseptic meningitis (n=28)	Mean±SD, n (%) <i>p</i> value
Age (yrs)	39.29±16.54	41.13±15.24	43.28±13.31	0.582
Male, n (%)	16 (43.2)	6 (40.0)	12 (42.9)	0.976
CSF WBC (count)	0.85±1.03	328.33±194.67* [‡]	100.93±122.02 [†]	0.001
CSF glucose (mg/dL)	65.59±12.89	45.53±12.25* [‡]	62.21±10.64	0.001
Serum glucose (mg/dL)	93.73±10.66	96.67±10.51	96.36±9.90	0.502
CSF protein (mg/dL)	29.39±7.12	243.00±99.87* [‡]	68.43±28.75 [†]	0.001
CSF NSE (ng/dL)	9.46±5.76	12.35±3.52	9.79±6.10	0.225
Serum NSE (ng/dL)	10.42±6.06	8.00±2.17	9.57±7.16	0.874
CSF/serum NSE ratio	0.92±0.30	1.62±0.59* [‡]	1.07±0.23	0.001
CSF/serum glucose ratio	0.68±0.09	0.47±0.12* [‡]	0.65±0.13	0.001

SD, standard deviation; WBC, white blood cell; NSE, neuron specific enolase; CSF, cerebrospinal fluid; Tb, tuberculosis.

* $p < 0.05$ (control versus Tb meningitis).

[†] $p < 0.05$ (control versus aseptic meningitis).

[‡] $p < 0.05$ (Tb meningitis versus aseptic meningitis).

Table 2. The Results of Univariate and Multivariate Logistic Regression for Tuberculosis Meningitis Diagnosis

Variable	Univariate		Multivariate	
	Odds ratio	<i>p</i> value	Odds ratio	<i>p</i> value
Sex	1.135	0.828		
Age	1.001	0.978		
CSF WBC	1.011	0.001	1.012	0.004
CSF glucose	0.869	0.001	1.035	0.725
Serum glucose	0.541	1.017		
CSF protein	78.770	0.970		
CSF NSE	1.076	0.102		
Serum NSE	0.873	0.272		
CSF/Serum NSE ratio	91.504	0.001	51.418	0.022
CSF/Serum glucose ratio	0.231	0.001	0.01	0.204

CSF, cerebrospinal fluid; WBC, white blood cell; NSE, neuron specific enolase.

enhancement, 3 patients showed cerebral edema, and one patient showed hydrocephalus. There were significant differences in CSF white blood cell (WBC) count, CSF glucose level, CSF protein level, CSF glucose/serum glucose ratio and CSF/serum NSE ratio ($p=0.001$) among the three groups.

On the Scheffe's post-hoc analysis, CSF WBC count was higher in the TbM group than in the aseptic ($p=0.002$) and control groups ($p=0.001$); CSF glucose was lower in the TbM group than in the aseptic ($p=0.001$) and control groups ($p=0.001$); CSF protein was higher in the TbM group than in the aseptic ($p=0.001$) and control groups ($p=0.001$); CSF/serum NSE ratio was higher in the TbM group than in the aseptic ($p=0.001$) and control groups ($p=0.001$); and CSF/serum glucose ratio was lower in the TbM group than in the aseptic ($p=0.001$) and control groups ($p=0.001$). We performed subgroup analysis dividing the TbM group into confirmative and presumptive

groups, and there were no differences in laboratory findings between the two groups (confirmative and presumptive groups) by Mann-Whitney U test. Moreover, in one-Way ANOVA and Kruskal-Wallis test, CSF WBC count, CSF glucose level, CSF protein level, CSF NSE/serum NSE ratio, and CSF glucose/serum glucose ratio were significantly different among the control, aseptic and confirmative TbM group, as well as among the control, aseptic and presumptive group. In univariate logistic regression, higher CSF WBC count ($p=0.001$), lower CSF glucose ($p=0.001$), higher CSF/serum NSE ratio ($p=0.001$) and lower CSF/serum glucose ratio ($p=0.001$) were significant factors for diagnosis of TbM. In multivariate logistic regression, CSF WBC count ($p=0.004$) and CSF/serum NSE ratio ($p=0.022$) were significant prognostic factors for diagnosis of TbM (Table 2). In ROC curve, when the cut-off value of the CSF/serum NSE ratio was 1.21, the sensitivity was 86.7% and the specificity was 75.4%.

DISCUSSION

Late diagnosis and treatment of TbM leads to high mortality and poor prognosis.^{13,14} Therefore, rapid and accurate diagnosis is necessary for improved outcomes. Many laboratory results exist regarding diagnosis of TbM including the Ziel-Neelsen stain, CSF culture, radioactive bromide partition test,¹⁵ immunoassay,¹⁶ and latex particle agglutination.¹⁷ However, Ziel-Neelsen staining of CSF lacks sensitivity, the rate of positivity of which is approximately 40% in culture, and approximately six weeks are required to obtain a result.^{18,19} Our study showed lower detection rates than those of previous results^{18,19} and only a single positive result with Ziel-Neelsen staining (6.6%). The solid culture of *M. tuberculosis* from CSF is still the best confirmatory test for the diagnosis of TbM, but its sensitivity is limited. In addition, this procedure takes at least five to six weeks to demonstrate growth of *M. tuberculosis*. In the current study, *M. tuberculosis* culture was only positive in three patients (20.0%).

In our study, eight patients showed typical brain imaging findings for TbM in the presumptive group, exhibiting hydrocephalus, cerebral edema or basal meningeal enhancement on brain CT or MRI. These brain image findings suggested a high possibility of brain neuronal cell damage via vasculitis and tissue hypoxic damage. The elevation of CSF NSE in our study could be explained by the mechanism of neuron damage due to vasculitis or tissue hypoxia. Contrary to our data, a previous study²⁰ revealed the CSF NSE level in the viral meningitis group only was higher than that in other groups, while there was no difference between the TbM and control groups. This discrepancy may be due to the small number of TbM cases in this study. In addition, high NSE level in the mumps meningitis patients increased the overall mean level.²⁰ Moreover, in our study, we discerned that aseptic meningitis does not influence neuron damage or tissue hypoxia. Our study sample was small and there were no mumps, herpes zoster or herpes simplex infections diagnosed by PCR. In addition, there were no seizures or mental changes in the aseptic meningitis group. For these reasons, we think there was no brain damage in the aseptic meningitis group in our study.

The polymerase chain reaction (PCR) has entailed a relatively better detection rate for tuberculous agents; however, the test has not been completely validated and is often not available in developing countries where TbM is common.^{21,22}

Although immunoassays based on CSF antigen-antibody reactions, including enzyme-linked immunosorbent assay (ELISA), have been reported with variable sensitivities and specificities^{16,17,23} these diagnostic methods cannot be readily used just anywhere. CSF adenosine deaminase activity (ADA) has been studied for diagnosis of TbM.^{24,25} CSF ADA has shown a relatively high validity, with a sensitivity ranging from 44% to 100% and a specificity of 75% to 99%. However, previous study that showed a highly reliable result with a sensitivity of 100% and specificity of 99% in only three cases.²⁴ In addition, CSF culture for TbM requires two months to produce results. Overall, radioactive bromide partition test,¹⁵ immunoassay,¹⁶ latex particle agglutination,¹⁷ PCR, ADA and ELISA are not always accessible, and their reliabilities have not been thoroughly investigated.

In the present study, the sensitivity and specificity for CSF/serum NSE ratio were comparable with those of previous studies, especially the results of PCR, ELISA or ADA tests.^{15-18,23-25} Moreover, CSF and serum NSE results can be acquired more quickly and simply.

There are limitations in our study. First, our results showed a relatively low sensitivity and specificity for diagnosis of TbM. Neuron-Specific enolase estimation can non-specifically be raised in any brain disorder. Second, other chronic meningitis and partially treated bacterial meningitis cannot be differentiated by NSE, even though we excluded meningitis patients who showed suspected bacterial or other meningitis on clinical, laboratory and brain image findings. Third, the small sample size is an important limitation of our study. With only 15 cases in the disease group, the statistical reliability is weak.

In conclusion, CSF/serum NSE ratio could be a useful parameter for the early diagnosis of TbM. In addition, the authors of the present study suggest a cut-off value of 1.21 for CSF/serum NSE ratio. However, because the sensitivity and specificity were not high, clinical correlation and brain image study have to be considered for the diagnosis of TbM.

REFERENCES

1. Kashyap RS, Agarwal NP, Chandak NH, Taori GM, Biswas SK, Purohit HJ, et al. The application of the Mancini technique as a diagnostic test in the CSF of tuberculous meningitis patients. *Med Sci Monit* 2002;8:MT95-8.
2. Kashyap RS, Kainthla RP, Biswas SK, Agarwal N, Chandak NH, Purohit HJ, et al. Rapid diagnosis of tuberculous meningitis using the Simple Dot ELISA method. *Med Sci Monit* 2003;9:MT123-6.

3. Kilpatrick ME, Girgis NI, Yassin MW, Abu el Ella AA. Tuberculous meningitis--clinical and laboratory review of 100 patients. *J Hyg (Lond)* 1986;96:231-8.
4. Kashyap RS, Kainthla RP, Satpute RM, Agarwal NP, Chandak NH, Purohit HJ, et al. Differential diagnosis of tuberculous meningitis from partially-treated pyogenic meningitis by cell ELISA. *BMC Neurol* 2004;4:16.
5. Radhakrishnan VV, Mathai A. Detection of Mycobacterium tuberculosis antigen 5 in cerebrospinal fluid by inhibition ELISA and its diagnostic potential in tuberculous meningitis. *J Infect Dis* 1991;163:650-2.
6. DeGiorgio CM, Correale JD, Gott PS, Ginsburg DL, Bracht KA, Smith T, et al. Serum neuron-specific enolase in human status epilepticus. *Neurology* 1995;45:1134-7.
7. Zerr I, Bodemer M, Racker S, Grosche S, Poser S, Kretzschmar HA, et al. Cerebrospinal fluid concentration of neuron-specific enolase in diagnosis of Creutzfeldt-Jakob disease. *Lancet* 1995;345:1609-10.
8. Butterworth RJ, Wassif WS, Sherwood RA, Gerges A, Poyser KH, Garthwaite J, et al. Serum neuron-specific enolase, carnosinase, and their ratio in acute stroke. An enzymatic test for predicting outcome? *Stroke* 1996;27:2064-8.
9. Ross SA, Cunningham RT, Johnston CF, Rowlands BJ. Neuron-specific enolase as an aid to outcome prediction in head injury. *Br J Neurosurg* 1996;10:471-6.
10. Bassetti C, Bomio F, Mathis J, Hess CW. Early prognosis in coma after cardiac arrest: a prospective clinical, electrophysiological, and biochemical study of 60 patients. *J Neurol Neurosurg Psychiatry* 1996;61:610-5.
11. Inoue S, Takahashi H, Kaneko K. The fluctuations of neuron-specific enolase (NSE) levels of cerebrospinal fluid during bacterial meningitis: the relationship between the fluctuations of NSE levels and neurological complications or outcome. *Acta Paediatr Jpn* 1994;36:485-8.
12. Oh SH, Lee JG, Na SJ, Park JH, Choi YC, Kim WJ. Prediction of early clinical severity and extent of neuronal damage in anterior-circulation infarction using the initial serum neuron-specific enolase level. *Arch Neurol* 2003;60:37-41.
13. Girgis NI, Sultan Y, Farid Z, Mansour MM, Erian MW, Hanna LS, et al. Tuberculosis meningitis, Abbassia Fever Hospital-Naval Medical Research Unit No. 3-Cairo, Egypt, from 1976 to 1996. *Am J Trop Med Hyg* 1998;58:28-34.
14. Verdon R, Chevret S, Laissy JP, Wolff M. Tuberculous meningitis in adults: review of 48 cases. *Clin Infect Dis* 1996;22:982-8.
15. Mann MD, Macfarlane CM, Verburg CJ, Wiggelinkhuizen J. The bromide partition test and CSF adenosine deaminase activity in the diagnosis of tuberculosis meningitis in children. *S Afr Med J* 1982;62:431-3.
16. Watt G, Zaraspe G, Bautista S, Laughlin LW. Rapid diagnosis of tuberculous meningitis by using an enzyme-linked immunosorbent assay to detect mycobacterial antigen and antibody in cerebrospinal fluid. *J Infect Dis* 1988;158:681-6.
17. Krambovitis E, McIlmurray MB, Lock PE, Hendrickse W, Holzel H. Rapid diagnosis of tuberculous meningitis by latex particle agglutination. *Lancet* 1984;2:1229-31.
18. Thwaites G, Chau TT, Mai NT, Drobniewski F, McAdam K, Farrar J. Tuberculous meningitis. *J Neurol Neurosurg Psychiatry* 2000;68:289-99.
19. Mathai A, Radhakrishnan VV, Sarada C, George SM. Detection of heat stable mycobacterial antigen in cerebrospinal fluid by Dot-Immunobinding assay. *Neurol India* 2003;51:52-4.
20. Rodrguez-Nnuez A, Cid E, Rodrguez-Garca J, Camina F, Rodrguez-Segade S, Castro-Gago M. Neuron-specific enolase, nucleotides, nucleosides, purine bases, oxypurines and uric acid concentrations in cerebrospinal fluid of children with meningitis. *Brain Dev* 2003;25:102-6.
21. Pfyffer GE. Nucleic acid amplification for mycobacterial diagnosis. *J Infect* 1999;39:21-6.
22. Wilson SM. Application of molecular methods to the study of diseases prevalent in low income countries. *Trans R Soc Trop Med Hyg* 1998;92:241-4.
23. Kashyap RS, Dobos KM, Belisle JT, Purohit HJ, Chandak NH, Taori GM, et al. Demonstration of components of antigen 85 complex in cerebrospinal fluid of tuberculous meningitis patients. *Clin Diagn Lab Immunol* 2005;12:752-8.
24. Pettersson T, Klockars M, Weber TH, Somer H. Diagnostic value of cerebrospinal fluid adenosine deaminase determination. *Scand J Infect Dis* 1991;23:97-100.
25. Choi SH, Kim YS, Bae IG, Chung JW, Lee MS, Kang JM, et al. The possible role of cerebrospinal fluid adenosine deaminase activity in the diagnosis of tuberculous meningitis in adults. *Clin Neurol Neurosurg* 2002;104:10-5.